



fNIRS Data Analysis with Homer3

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Neurophotonics Center, Boston University

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Outline

O Homer file formats

Basic processing :

- ♦ Intensity to hemoglobin concentrations
- ♦ **Filtering**
- ♦ Block-averaging

Our Advanced processing :

- **Other useful Homer functions**
- **Exporting results**
- Our Plot probe
- ♦ Group analysis

Probe source-detector geometry : .sd files

- fNIRS experimental data
- fNIRS experimental data*
- Data processing stream
- Processed data files

- : .nirs files
- : .snirf files (new)
- : .cfg files
- : groupResults.mat

.SD file format

Describes the 2D source/detector geometry

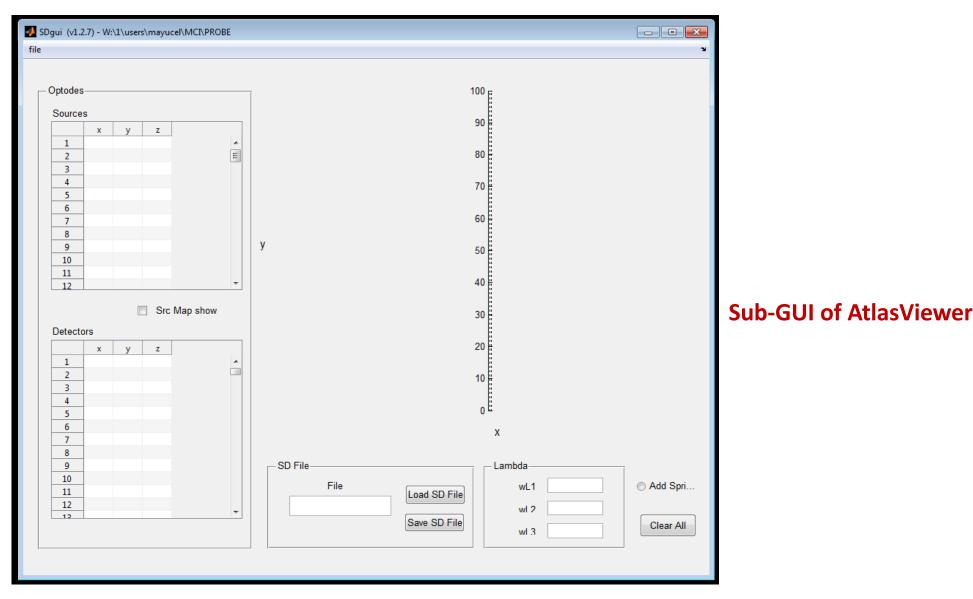
Matlab .sd file containing the structured variable **SD** with the following fields:

Field	Definition	Size	Example
Lambda	Wavelengths used for data acquisition	$1 \times n_{Wavelengths}$	[690 830]
nSrcs	Number of sources	1×1	1
nDets	Number of detectors	1×1	2
SrcPos	Array of source coordinates	nSrcs × 3	[0 0 0]
DetPos	Array of detector coordinates	nDets × 3	[30 0 0; 0 30 0]
MeasList	List of measurement channels <u>column 1 column 2 column 3 column 4</u> source idx detector idx unused wavelength idx	n _{Channels} × 4	[1 1 1 1; 1 2 1 1; 1 1 1 2; 1 2 1 2]

Note: The SD structure is also incorporated in the .nirs file format we will see in a few slides

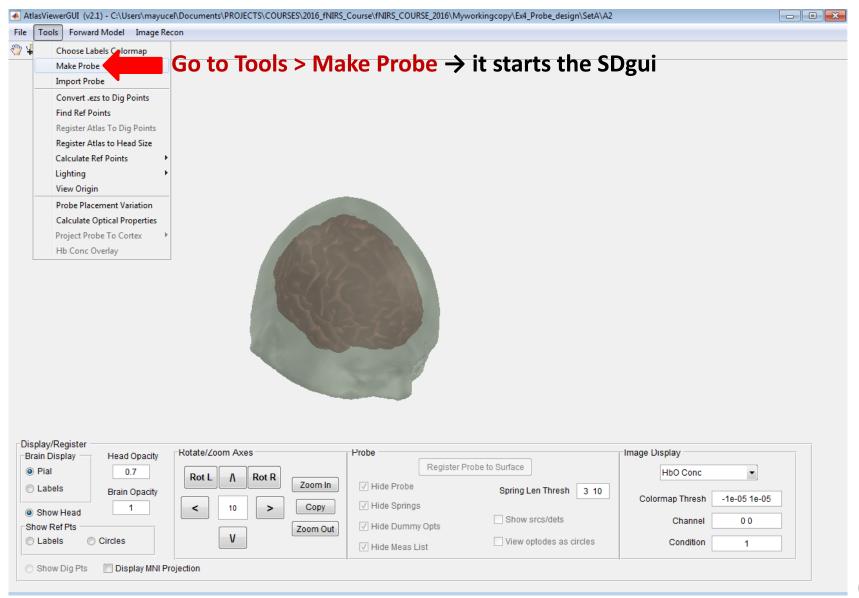
.SD file format: the SDgui

Enables you to create your own .sd files

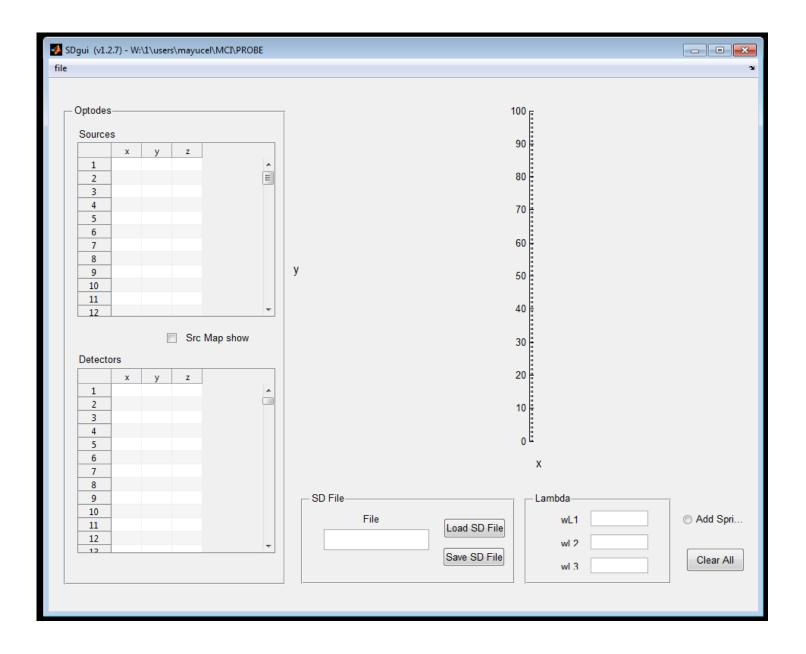


.SD file format: the SDgui

Start AtlasViewer



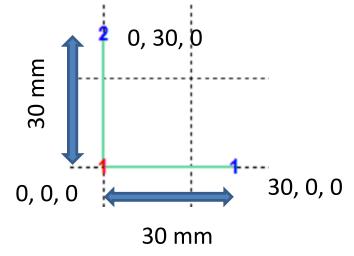
SDgui



How to start

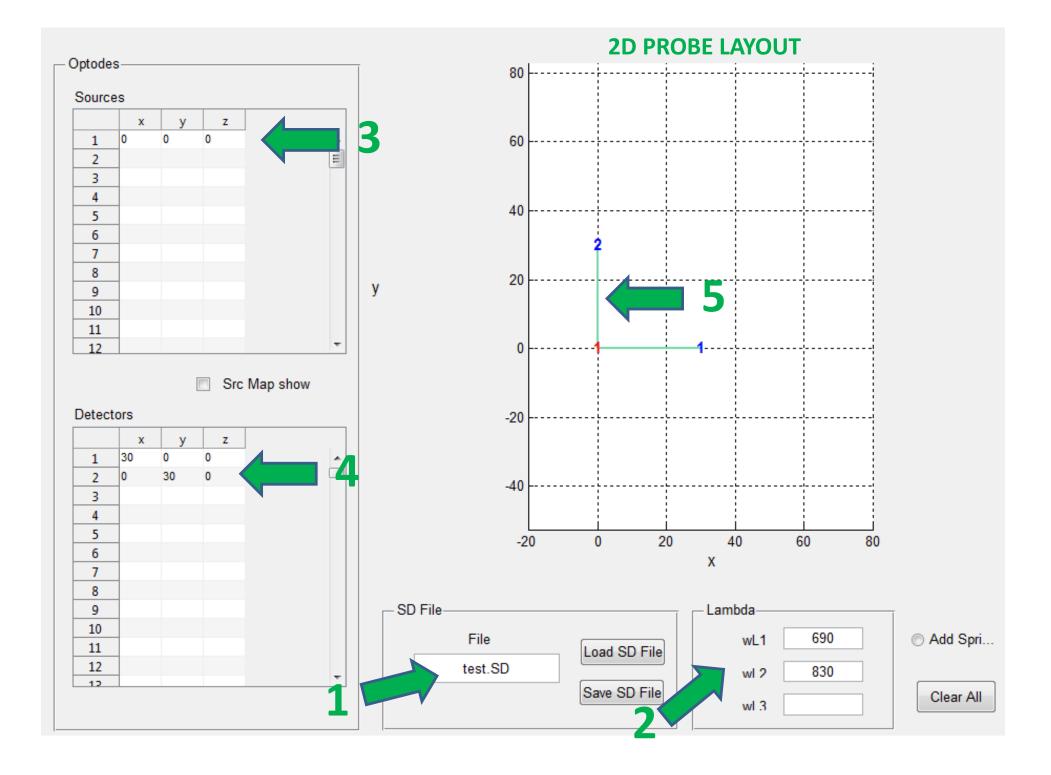
- 1. Begin by naming the new probe using the text box at the bottom-center of the GUI
- 2. Enter the wavelengths of light that will be used in the experiment (690 830)

Hint: easier if you have your 2D design on a piece of paper with the distances eg:



Adding sources, detectors and channels

- 3. Place a source at (0, 0, 0)
- 4. Add detectors at (30, 0, 0) and (0, 30, 0)
- 5. Connect measurement channels by clicking on optodes
 - to remove a channel do the same
 - you cannot connect two detectors or two sources



.NIRS file format

Contains an fNIRS dataset

Matlab .nirs file containing the following variables:

Field	Definition	Size
SD	Probe geometry, same as .sd files	Structured variable
t	Array of data time points	$N_{Time_{points}} \times 1$
d	Array of raw intensity time course	$N_{Time_{points}} \times N_{Channels}$
S	Array of stimulus onsets. Different conditions are a way to separate stimuli into different groups	$N_{Time_points} \times N_{Conditions}$
aux	Array of auxiliary signals	$N_{Time_{points}} \times N_{Aux channels}$
	In addition to the above parameters, additional param nirs file as a result of processing the data set in HOME.	
<u>Note 2</u> :	Conversion scripts are available for some other data fo	rmats (Hitachi, NIRx,

Shimadzu, ...), see *http://homer-fnirs.org/faq/*

.SNIRF file format

- many different fNIRS data file format
- difficult to use specific data analysis software and share data with collaborators
- Common format: Shared Near InfraRed File Format http://github.com/fNIRS/snirf
- SNIRF provides numeric fNIRS data with time stamps and a so-called 'measurement list' for detailed measurement information (in HDE5 format)

Finally, we have launched a SNIRF list server. The relevant addresses are here:

Mail list:	<u>snirf@fnirs.org</u>
Help address:	<u>snirf-help@fnirs.org</u>
Subscription address:	<u>snirf-subscribe@fnirs.org</u>
Unsubscription address:	snirf-unsubscribe@fnirs.org

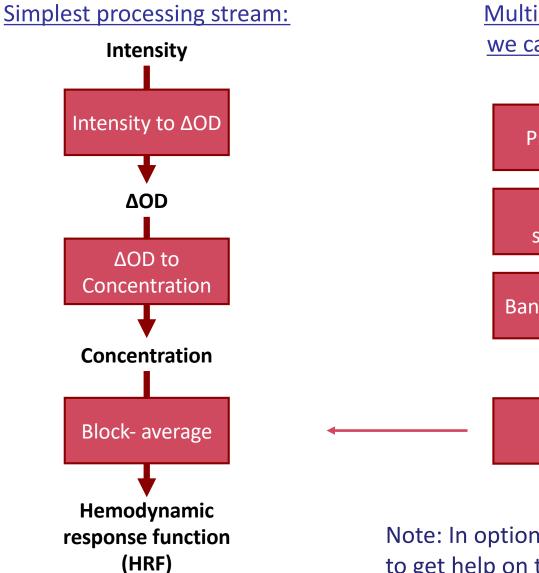
Shared Near Infrared File Format V1.0 Specification

- Document Version: Draft 3
- License: This document is in the public domain.

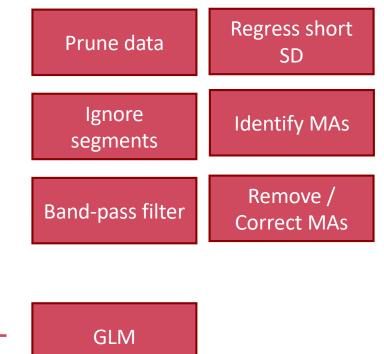
Table of Content

- Introduction
- SNIRF file specification
 - SNIRF data format summary
 - SNIRF data container definitions
 - formatVersion
 - nirs
 - metaDataTags
 - data
 - data.dataTimeSeries
 - data.time
 - data.measurementList
 - data.measurementList.sourceIndex
 - data.measurementList.detectorIndex

A .cfg file describes the steps of data analysis, a sequence of HOMER3 functions



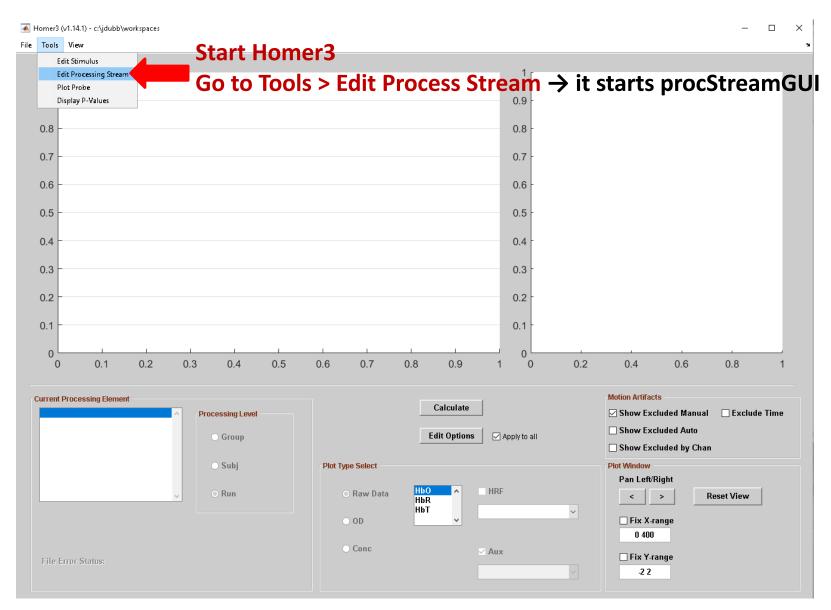
<u>Multiple more advanced functions</u> we can incorporate in the stream:



Note: In options, you can hover over each function to get help on the function and the parameters

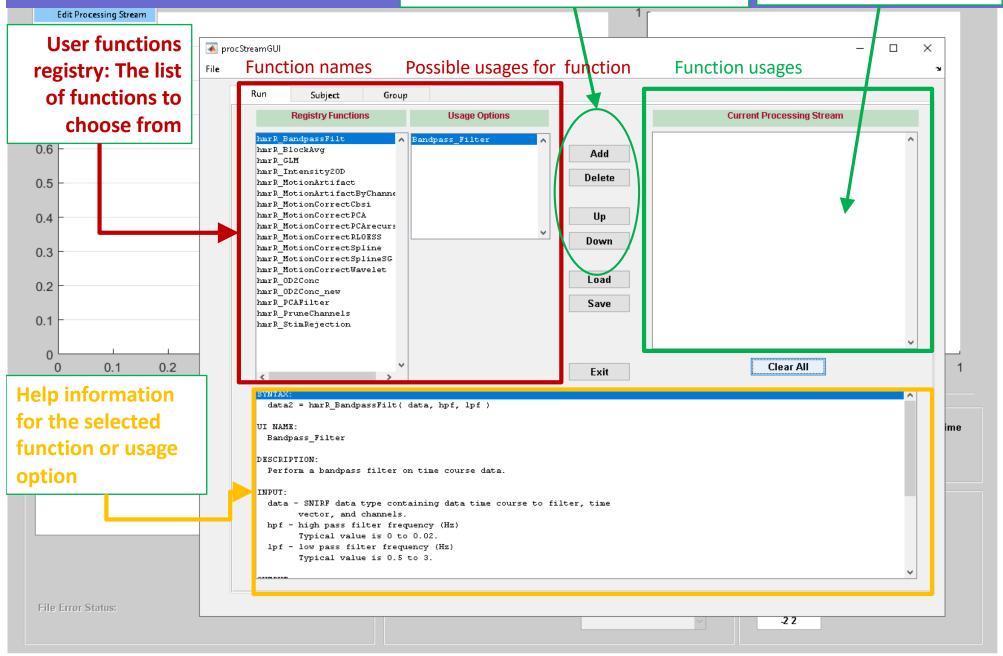
.CFG file format: the Process Stream GUI

The Process Stream GUI enables you to create your own .cfg files

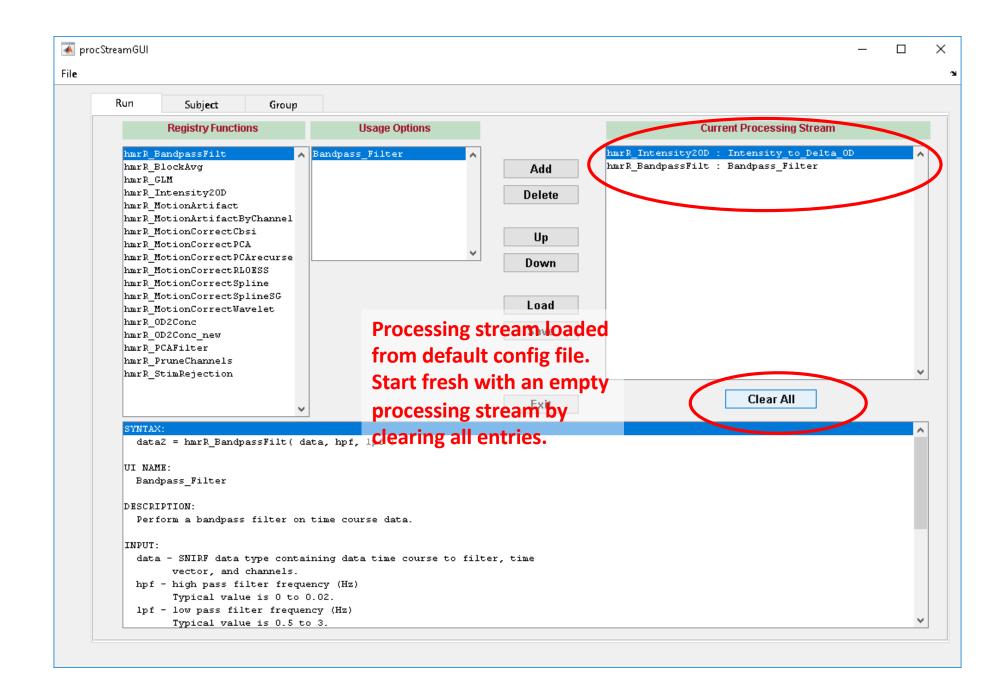


Add, remove, and move function usages in processing stream

The processing stream you're creating

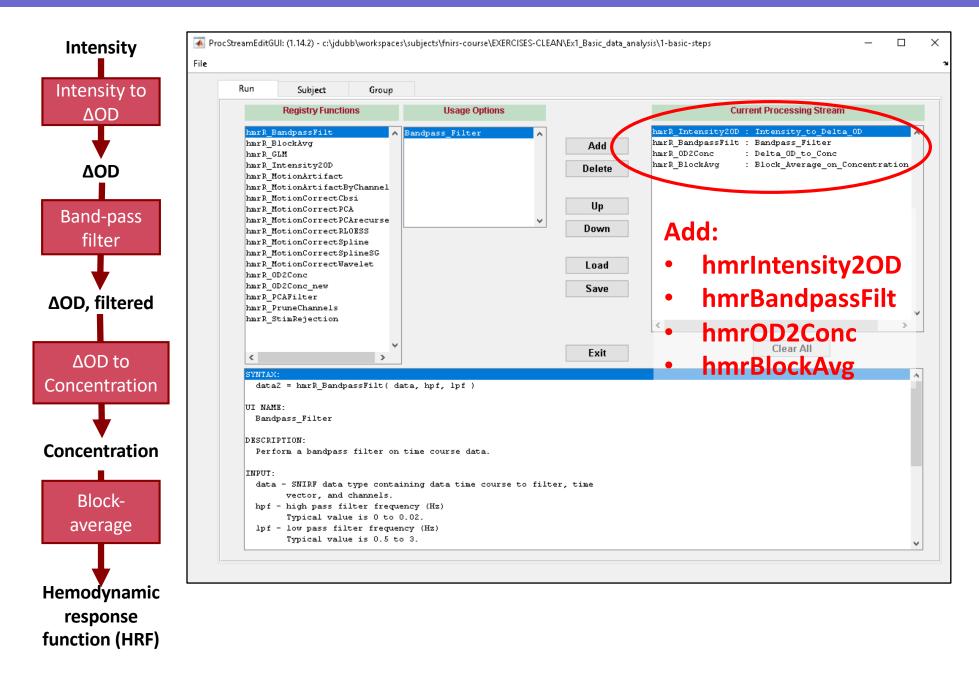


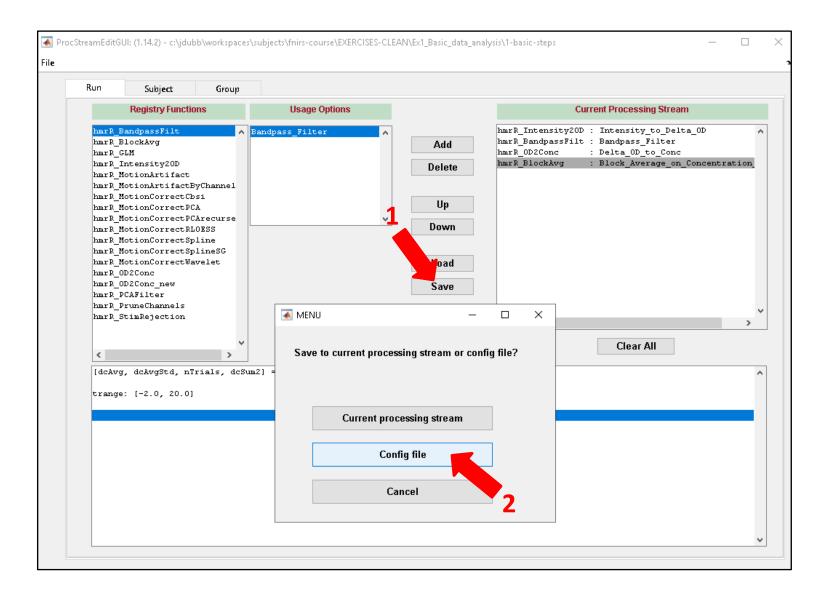
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ntensity to	Run Subject Group	
ΔOD	Registry Functions Usage Options Current Processing Stream	
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ΔOD	hmrR_GLM hmrR_Intensity20D hmrR_MotionArtifact	
	hmrR_MotionArtifactByChanne] hmrR_MotionCorrectCbsi	
	hmrR_MotionCorrectPCA hmrR_MotionCorrectPCArecurse hmrR_MotionCorrectRLOESS	
	hmrR_MotionCorrectSplineSG	
	hmrR_MotionCorrectWavelet hmrR_0D2Conc hmrR_0D2Conc_new Load	
	hmrR_PCAFilter hmrR_PruneChannels Save	
	hmrR_StimRejection	
		~
	SYNTAX: dod = hmrR_Intensity20D(intensity)	^
	UI NAME: Intensity_to_Delta_OD	
	DESCRIPTION:	
	Converts intensity data to optical density	
	INPUT: intensity - SNIRF data type where the d matrix is intensity	
	OUTPUT: dod - SNIRF data type where the d matrix is change in optical density	
	USAGE OPTIONS:	
	Intensity_to_Delta_OD: dod = hmrR_Intensity20D(data)	,

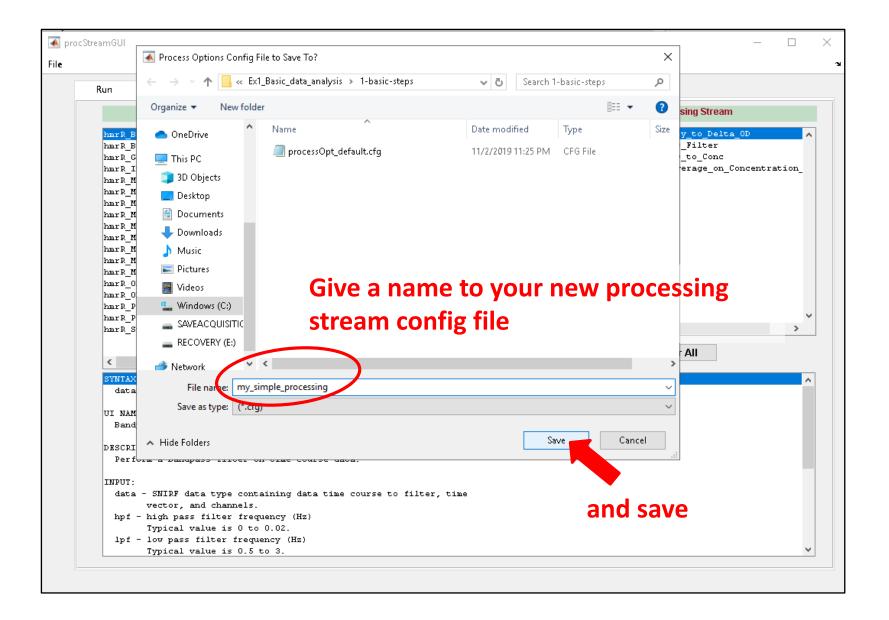


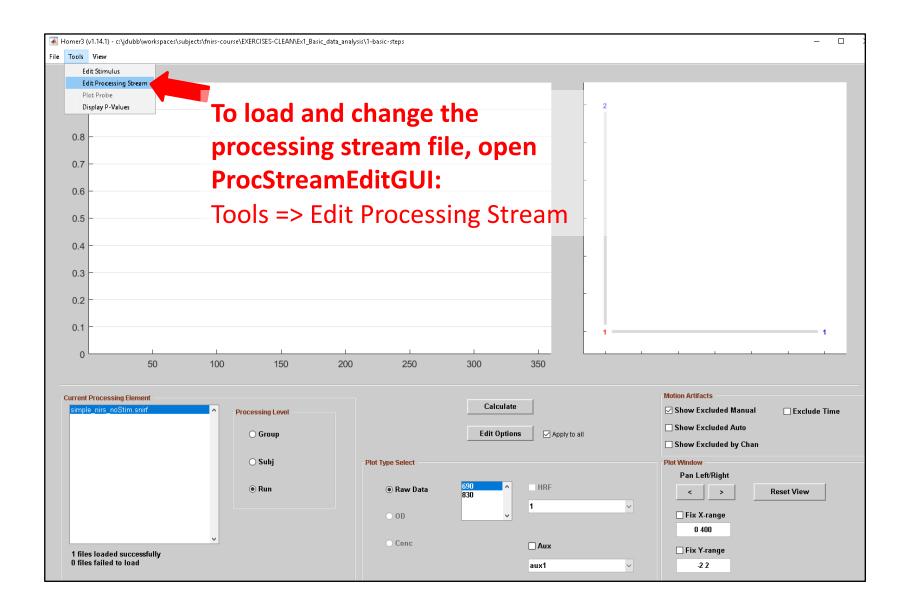
Your GUI should look like this now, with no entries in Current Processing Stream window

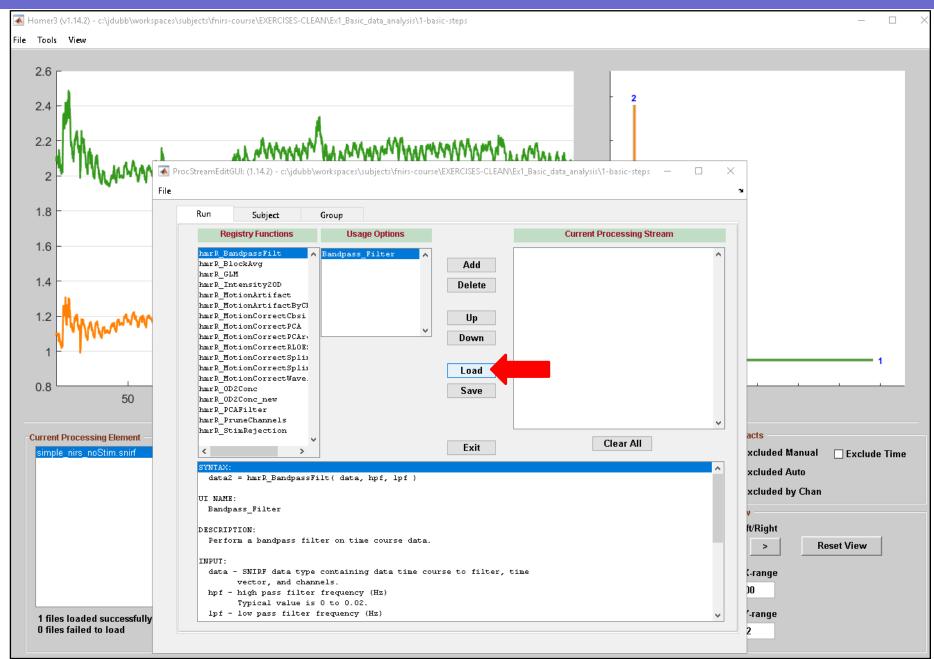
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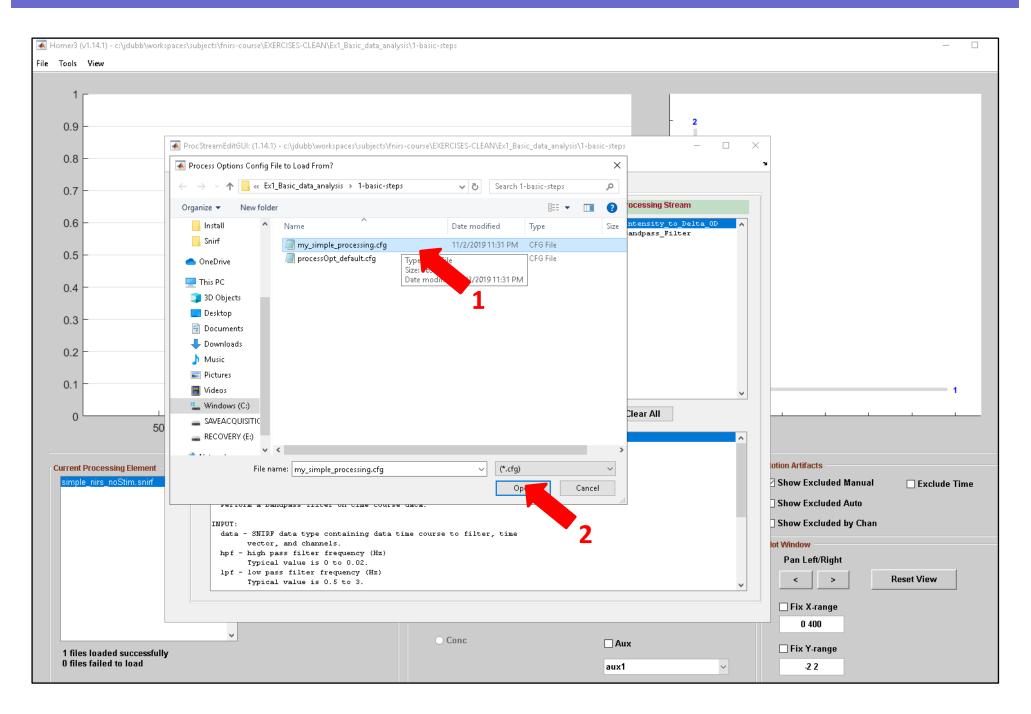


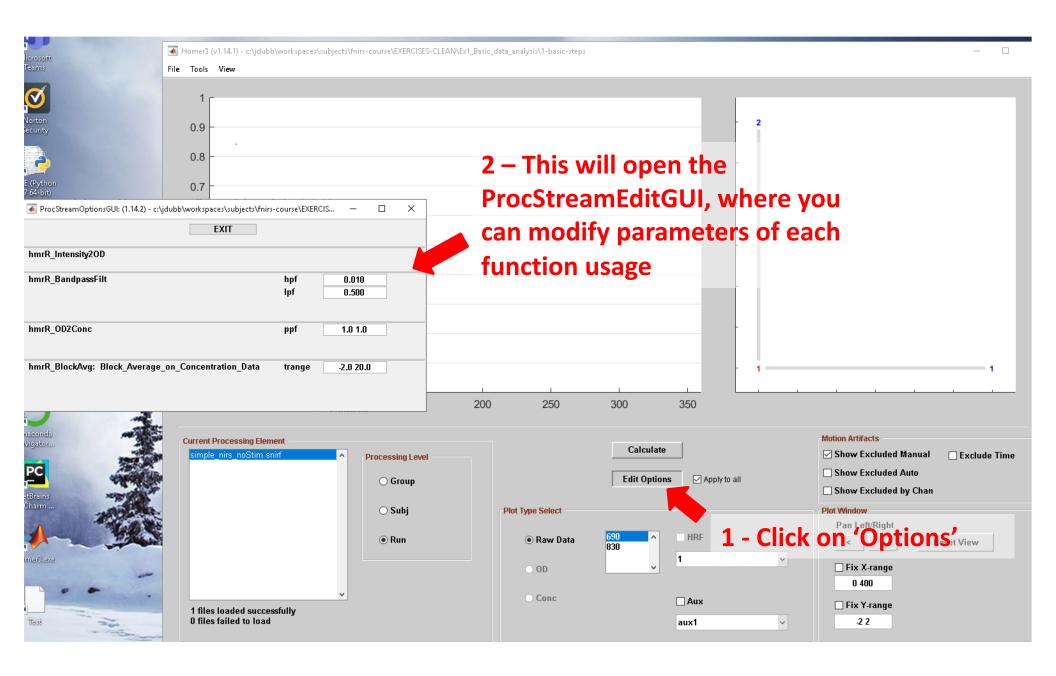












Let's analyze some fNIRS data: HOMER3_UI

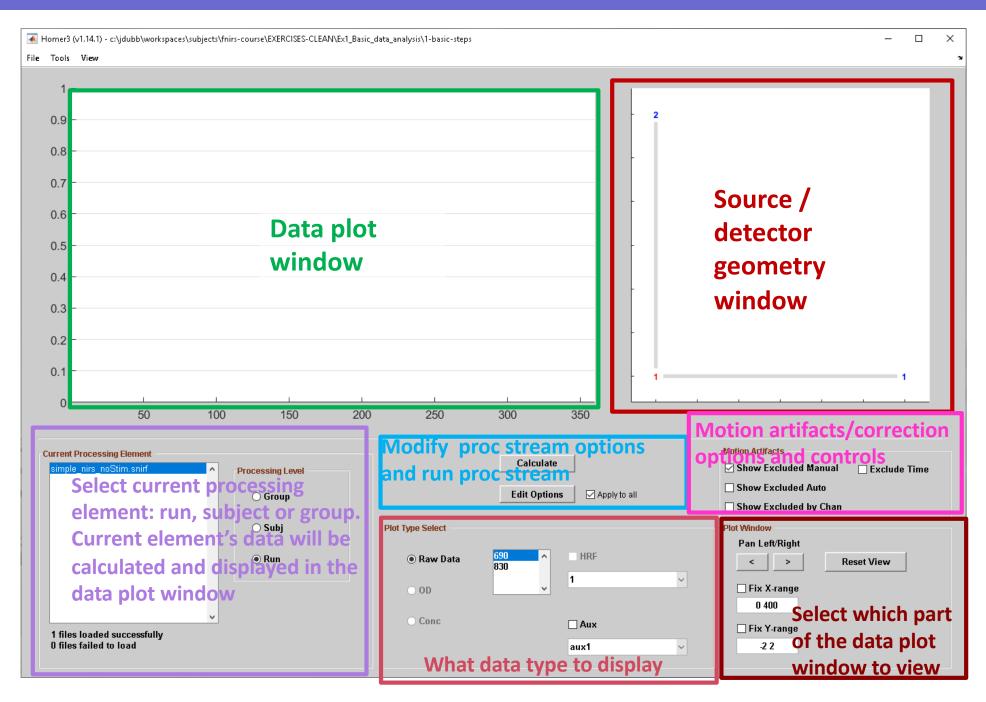
When opening Homer3, you need to select:

- The dataset(s): a folder containing at least one .snirf (or .nirs) file. If all you have are .nirs files, Homer3 will convert the files to the .snirf format.
- The data processing stream: a .cfg file. If you click 'Cancel', a default process stream will load.
- Start Homer3
- When prompted to select a .nirs folder, navigate to and select folder /Exercises/Ex1_Basic_data_analysis/
- When prompted to select a .cfg file, select my_simple_processing.cfg file we just created.

Note: If you're already in Homer3 and need to change the data directory or the .cfg file:

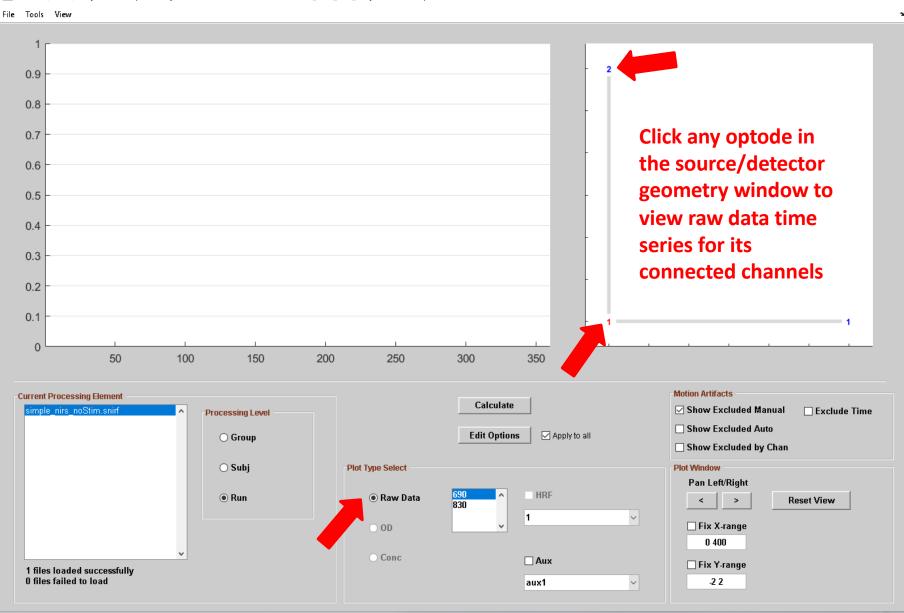
- Go to File > Change Directory, and select your nirs folder
- Go to **Tools > Edit Processing Stream** and click the Load button to select .cfg file

Homer3 MainGUI



Displaying raw fNIRS data

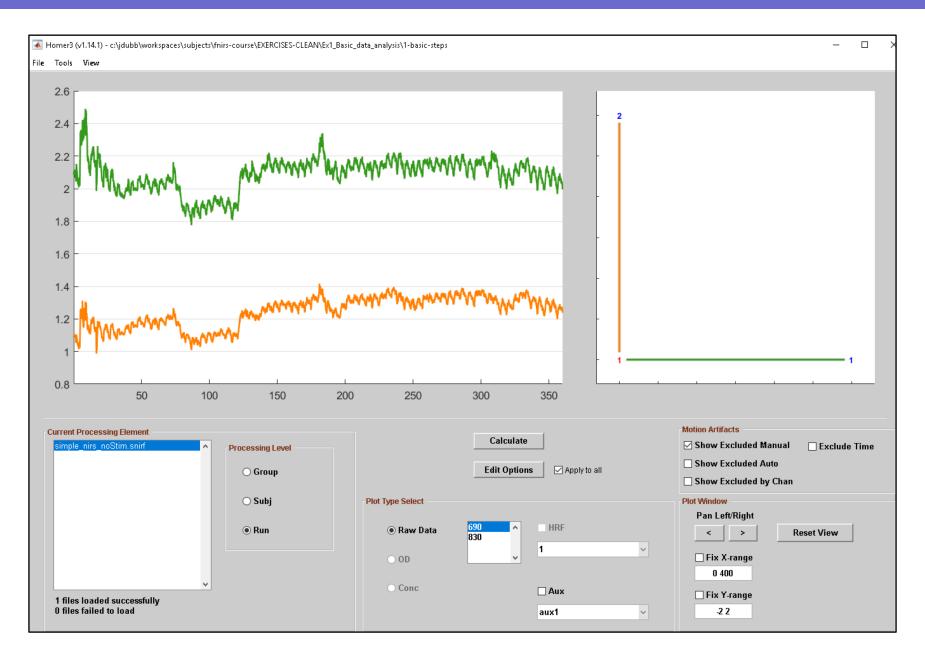
💽 Homer3 (v1.14.1) - c:\jdubb\workspaces\subjects\fnirs-course\EXERCISES-CLEAN\Ex1_Basic_data_analysis\1-basic-steps



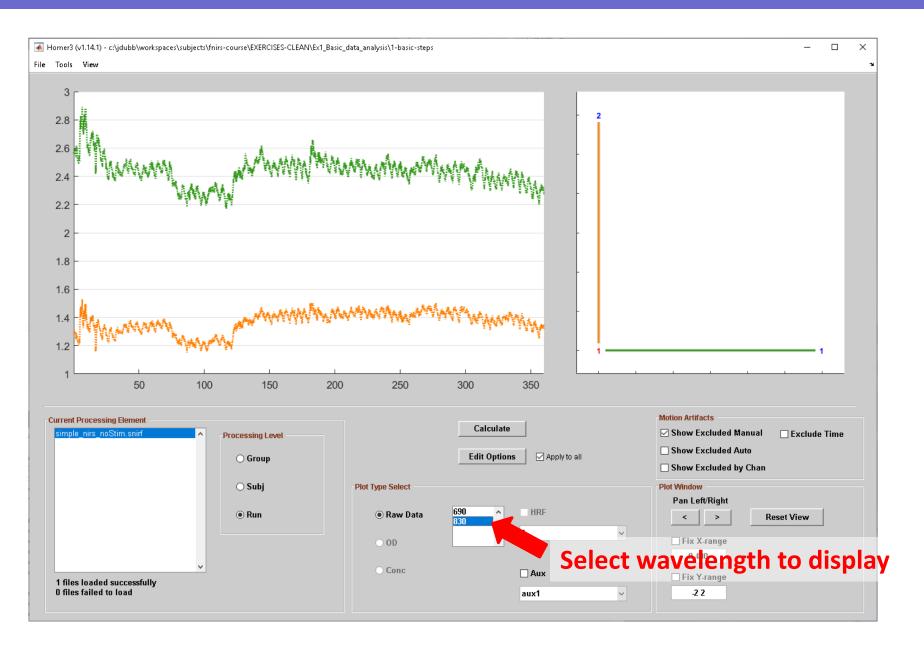
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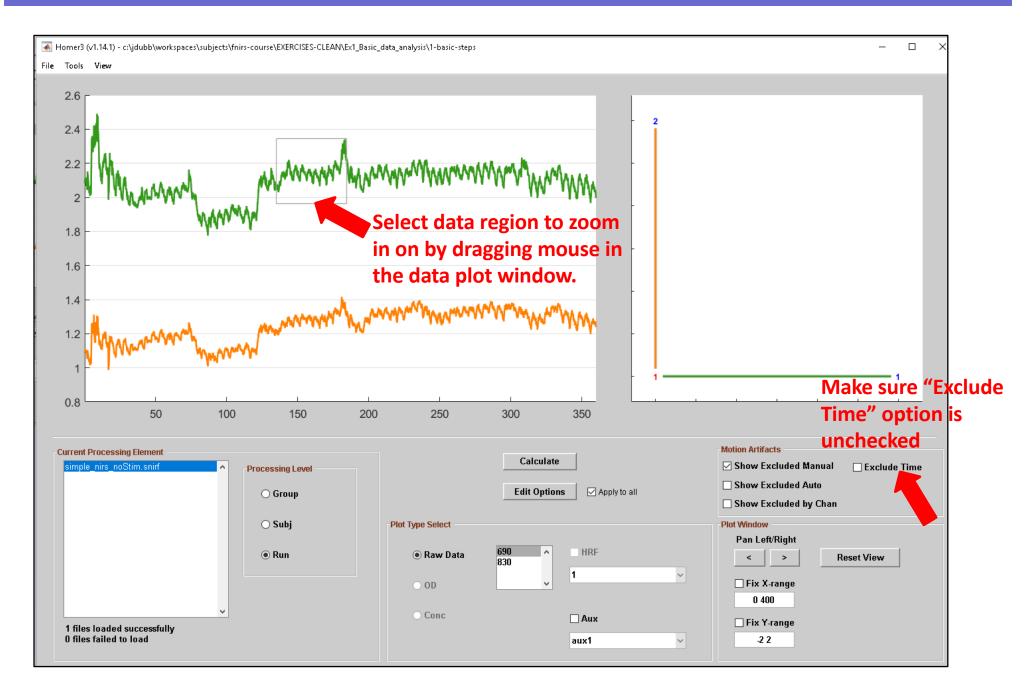
Displaying raw fNIRS data



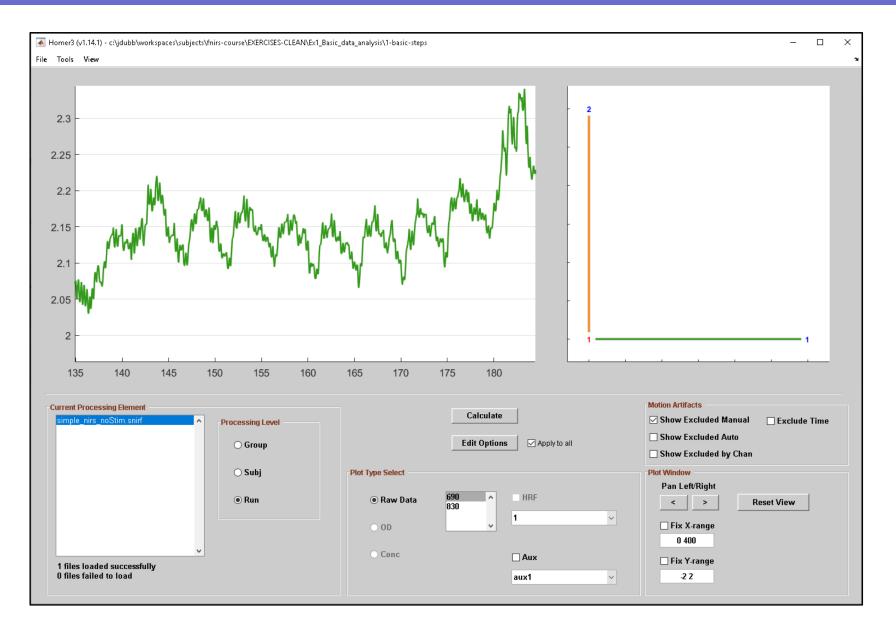
Displaying the raw fNIRS data



Zooming in on data

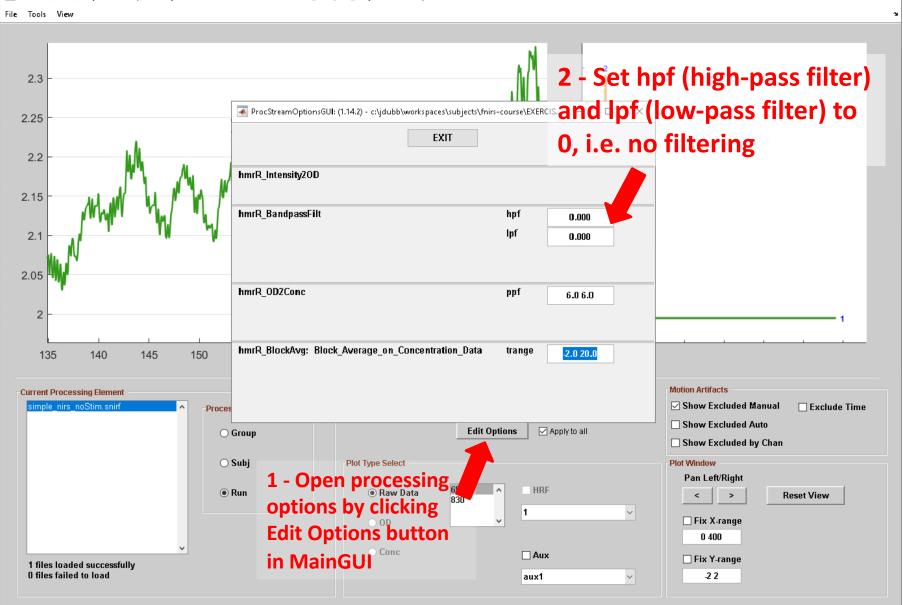


Zooming in



Modifying processing stream options

\overline Homer3 (v1.14.1) - c:\jdubb\workspaces\subjects\fnirs-course\EXERCISES-CLEAN\Ex1_Basic_data_analysis\1-basic-steps



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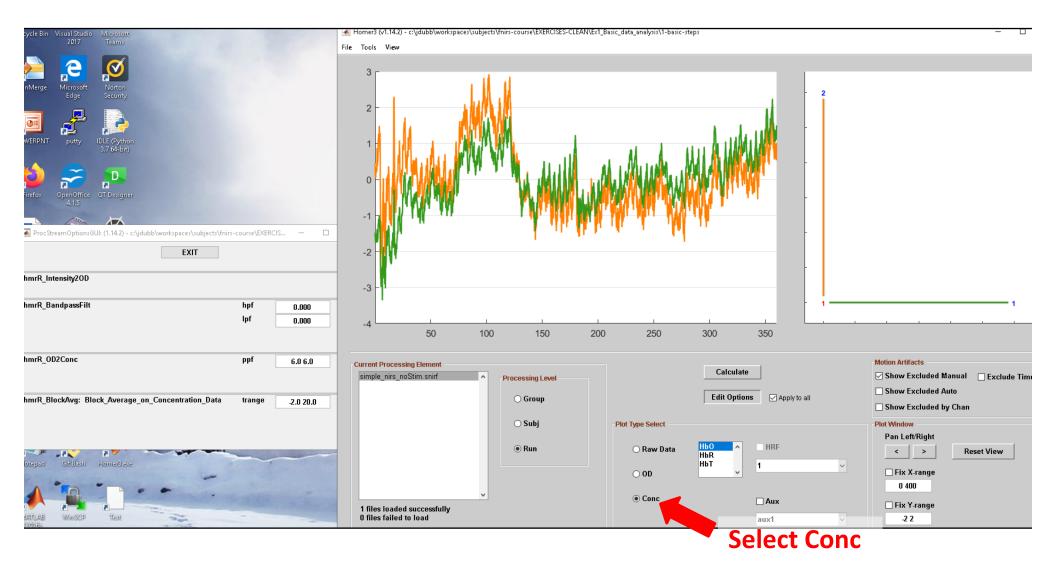
Running the processing stream



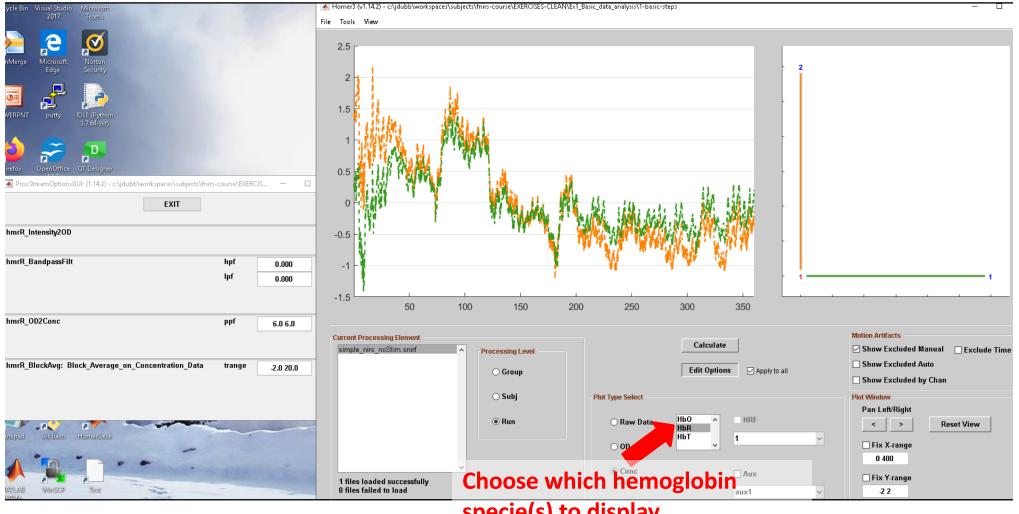
Intensity to ΔOD



△OD to Hb concentrations

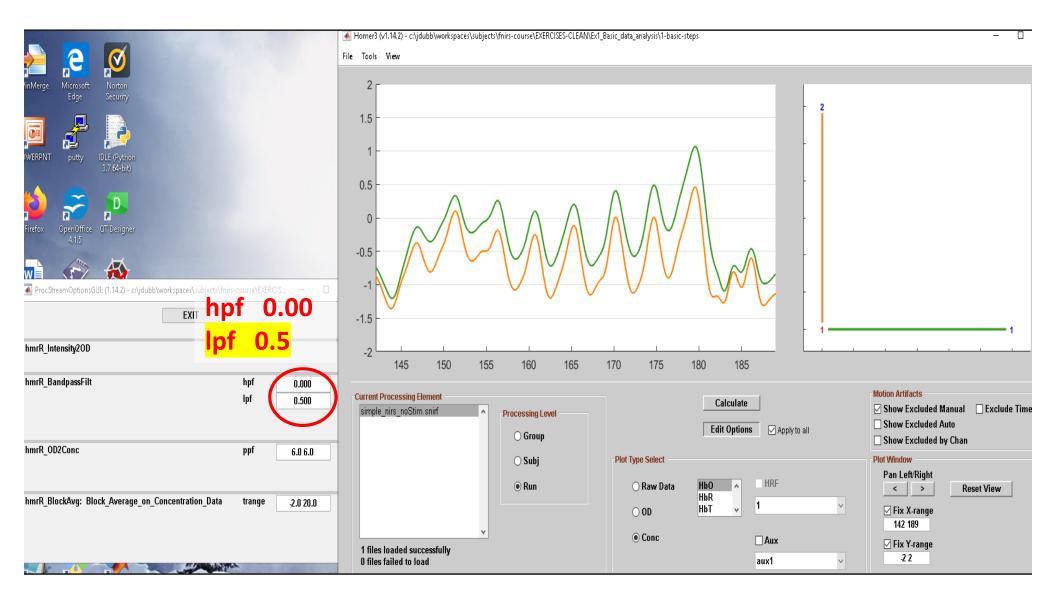


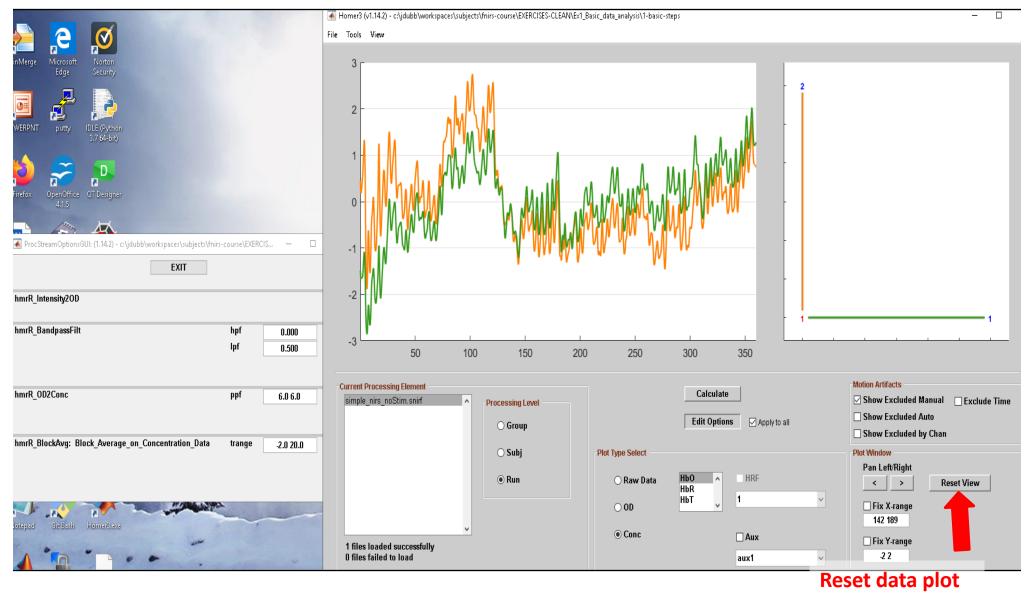
AOD to Hb concentrations



specie(s) to display





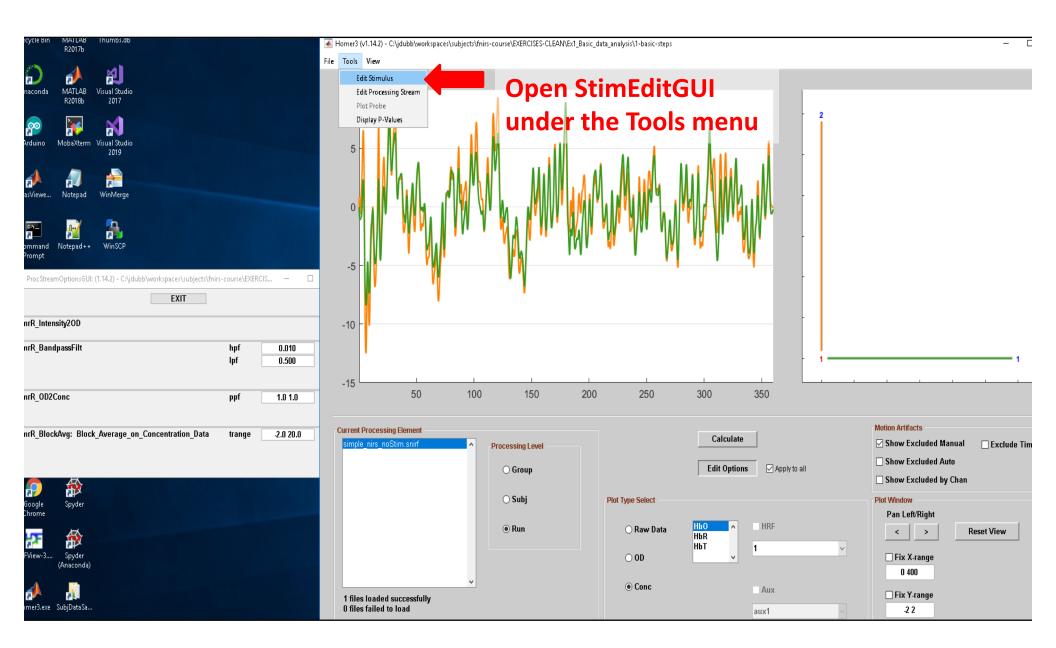


view



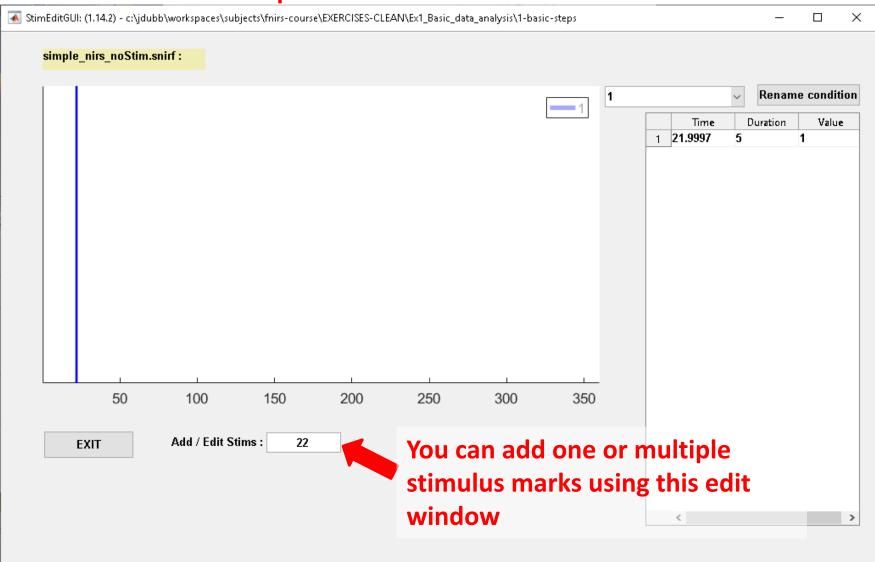


This dataset doesn't have stimulus onsets, so we will enter them manually



StimEditGUI

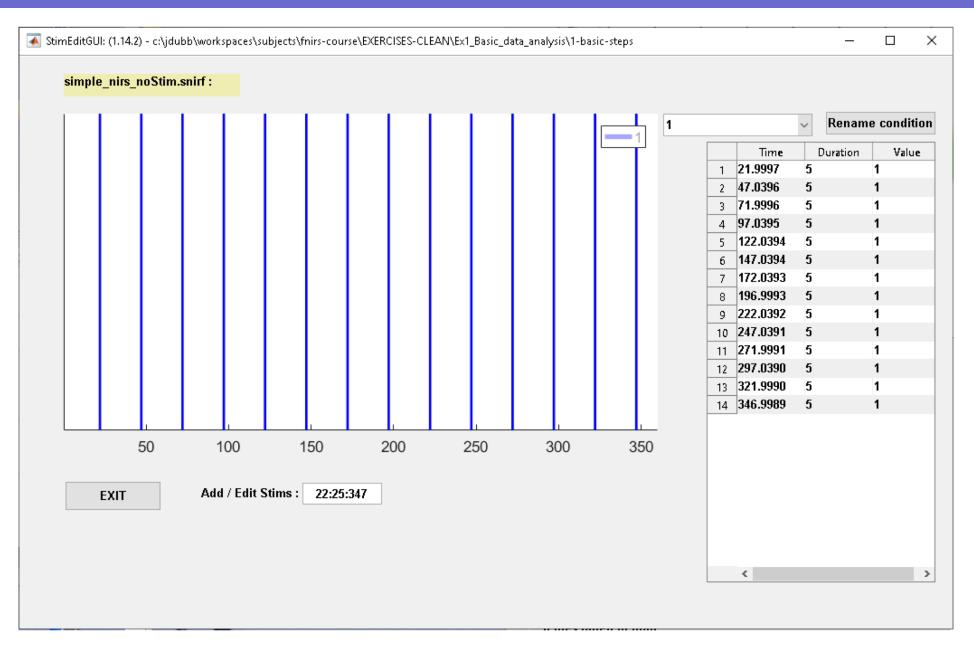
NOTE: Any edits to stims you make in StimEditGUI will automatically update the data plot window in MainGUI



StimEditGUI

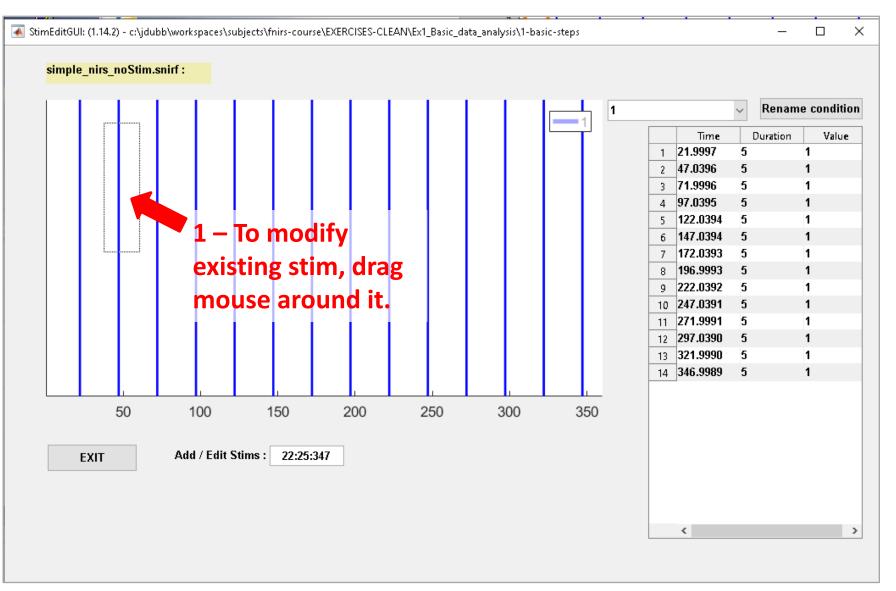
📣 StimEditGUI: (1.14.2) - c:\jdubb\workspaces\subjects	\fnirs-course\EXERCISES-CLEA	N\Ex1_Basic_data_	analysis\1-bas	sic-steps			_	
<pre>simple_nirs_noStim.snirf :</pre>								
				1	1	Time	Renam Duration	e condition Value
	MENU		- 0	×				
	Add stim mark at t=2	2.0						
	1 ···· New condition		– Sel	ect C	ondit	tion 1		
50 100	Cancel)	-			
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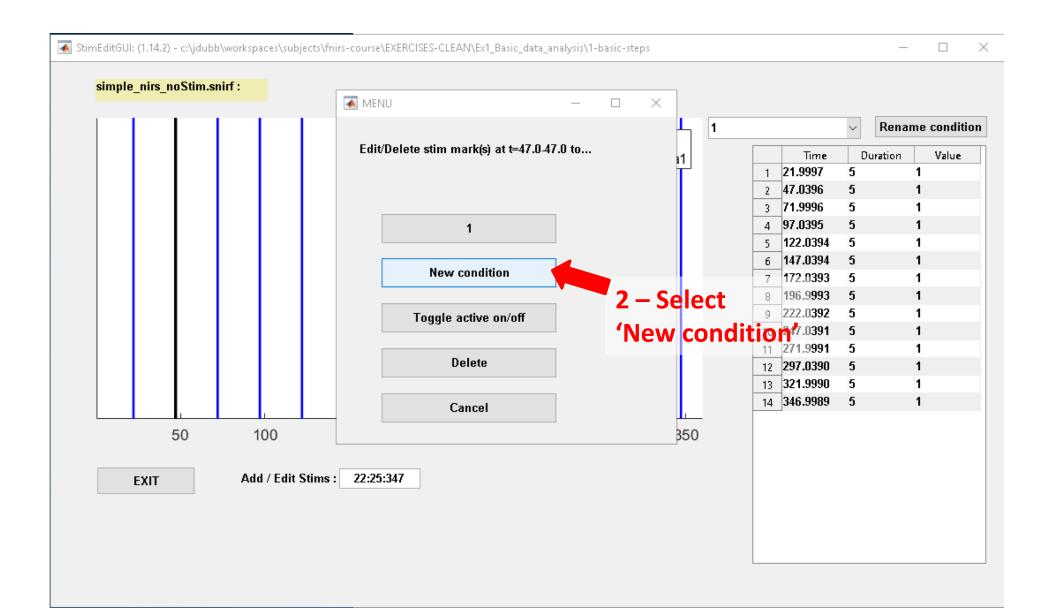
StimEditGUI

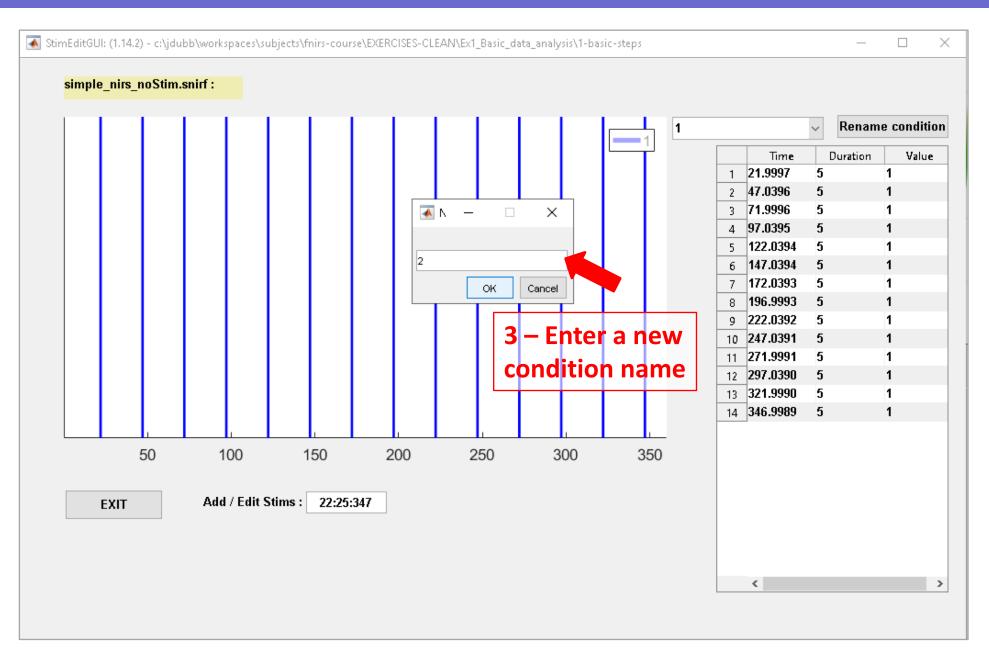


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	Add stim mark at t=22.0			
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50 100 EXIT Add / Edit Stims	: 22:25:347	р		
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... or by modifying existing stim marks









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simple_nirs_noStim.snirf : 4 – We now have two conditions

100

Add / Edit Stims :

50

EXIT

150

22:25:347

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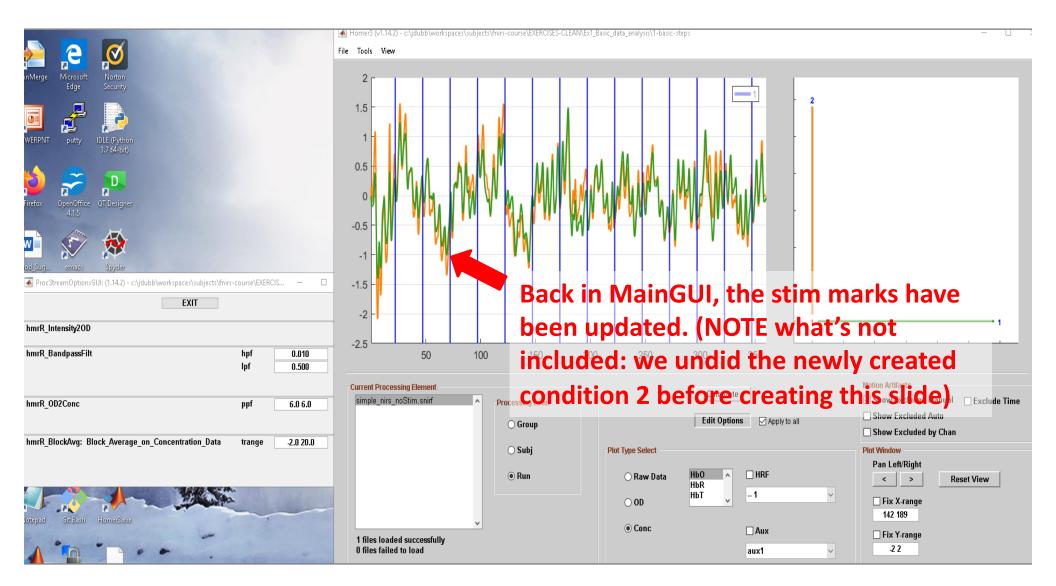
350

1 21.9997 5 1 2 71.9996 5 1 3 97.0395 5 1 4 122.0394 5 1 5 147.0394 5 1 6 172.0393 5 1 7 196.9993 5 1 8 222.0392 5 1 9 247.0391 5 1 10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1 13 346.9989 5 1		Time	Durat	ion	Value
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4 122.0394 5 1 5 147.0394 5 1 6 172.0393 5 1 7 196.9993 5 1 8 222.0392 5 1 9 247.0391 5 1 10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1	2	71.9996	5	1	
5 147.0394 5 1 6 172.0393 5 1 7 196.9993 5 1 8 222.0392 5 1 9 247.0391 5 1 10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1	3	97.0395	5	1	
6 172.0393 5 1 7 196.9993 5 1 8 222.0392 5 1 9 247.0391 5 1 10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1	4	122.0394	5	1	
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9 247.0391 5 1 10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1	7	196.9993	5	1	
10 271.9991 5 1 11 297.0390 5 1 12 321.9990 5 1	8	222.0392	5	1	
11 297.0390 5 1 12 321.9990 5 1	9	247.0391	5	1	
12 321.9990 5 1	10	271.9991	5	1	
	11	297.0390	5	1	
13 346.9989 5 1	12	321.9990	5	1	
	13	346.9989	5	1	

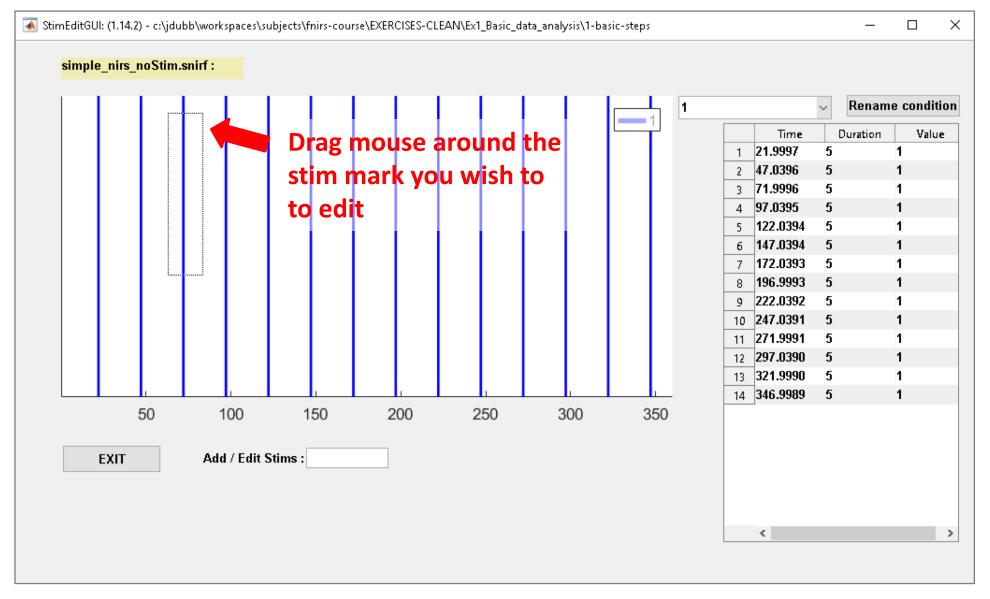
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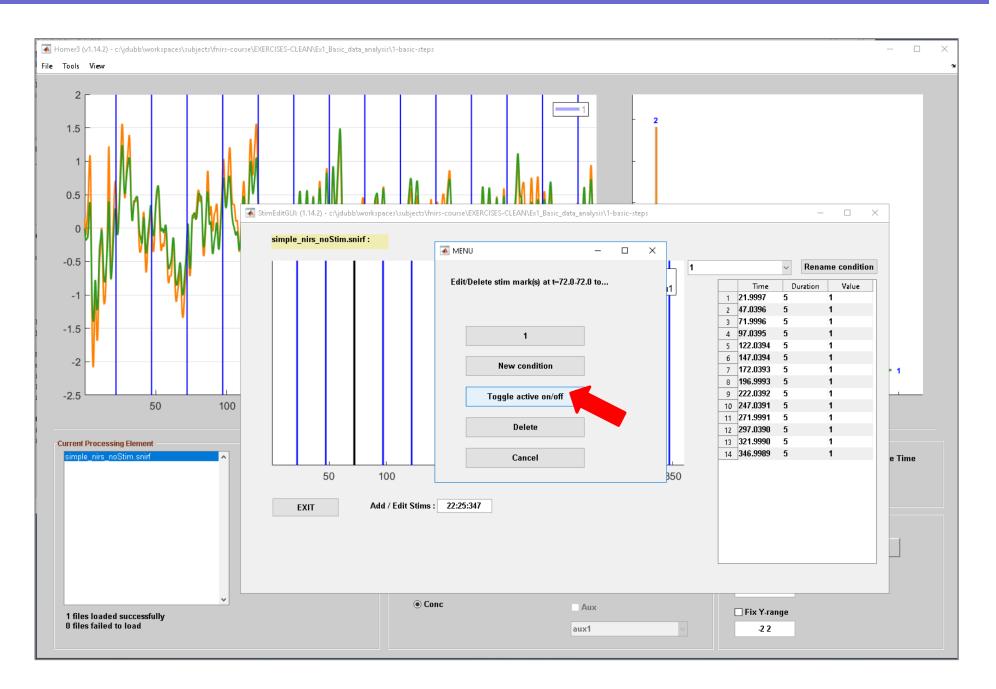
>

Stimulus onsets

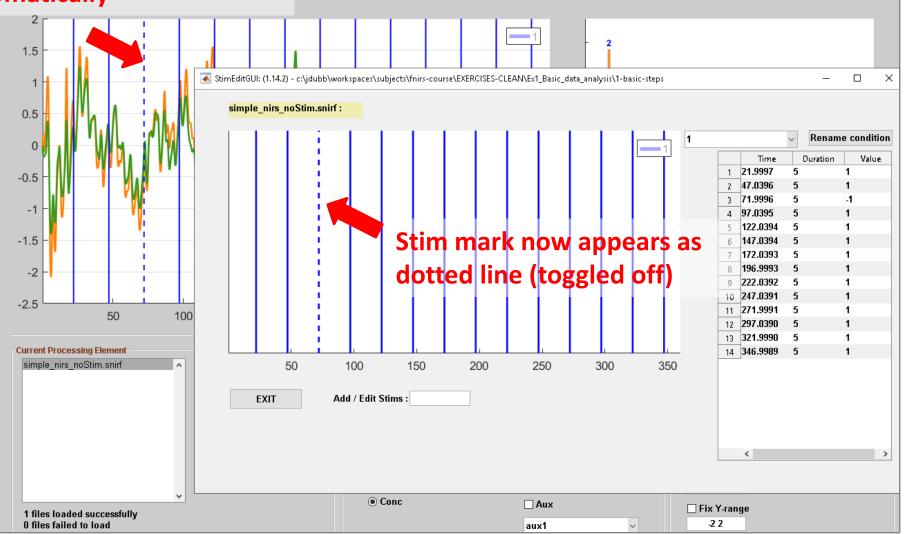


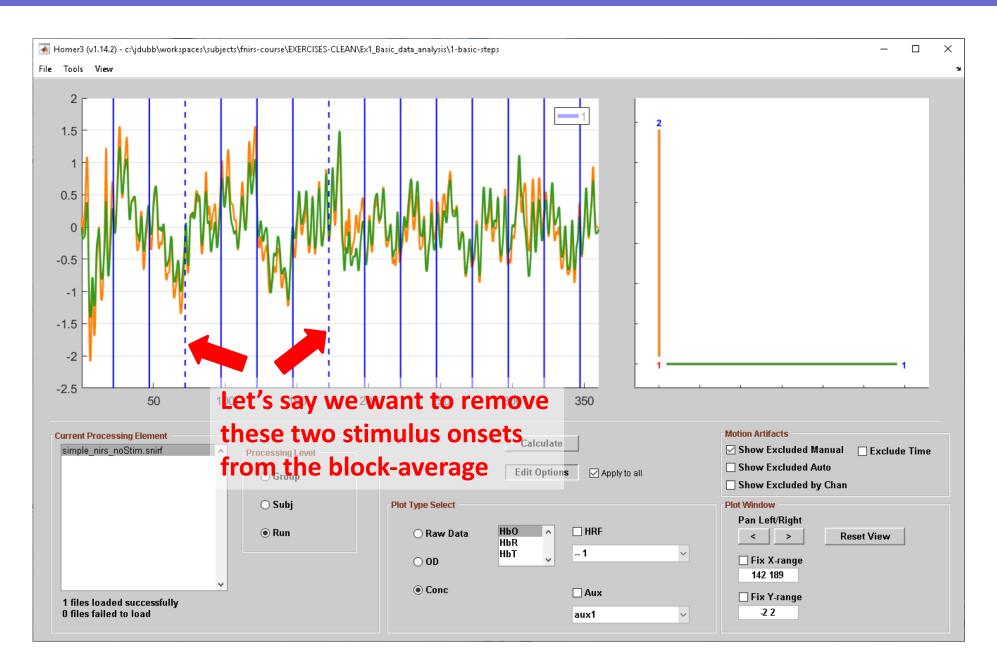
To reject stim go back to StimEditGUI



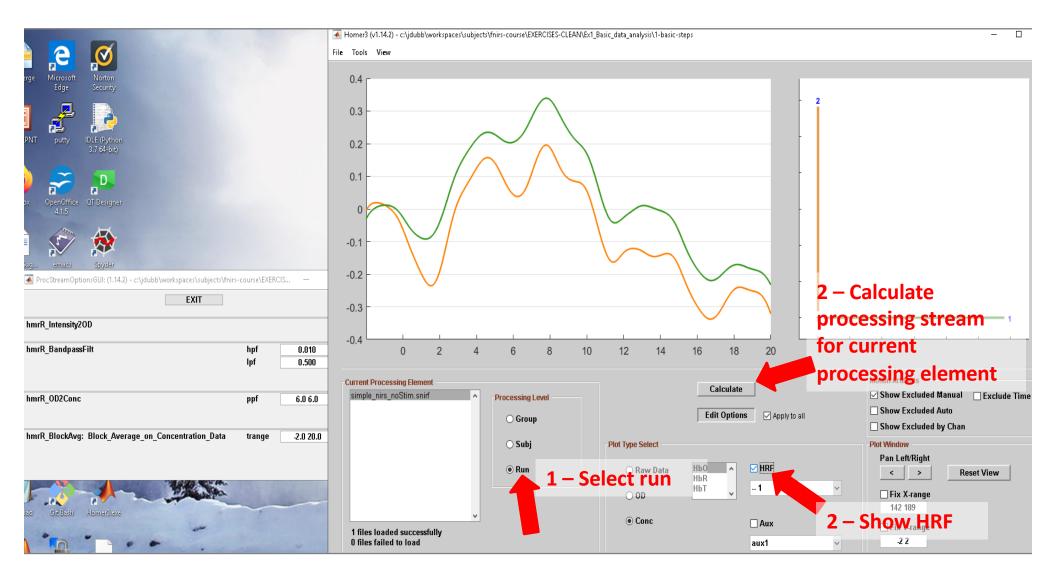


Updates stim in MainGUI Homers (v1.14.c) - crydubb (workspaces (subjects (fnils-course \EXERCISES-CLEAN\Ex1_Basic_data_analysis\1-basic-steps automTaltically





Calculating and displaying the HRF

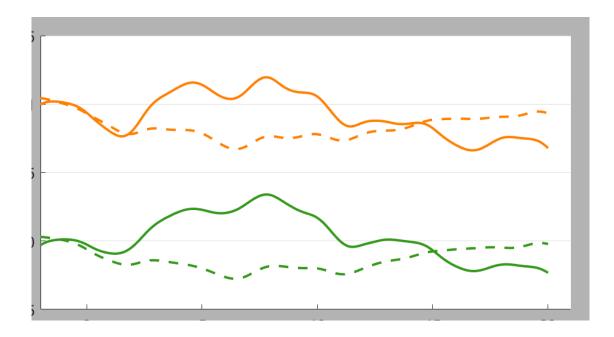


Displaying the HRF

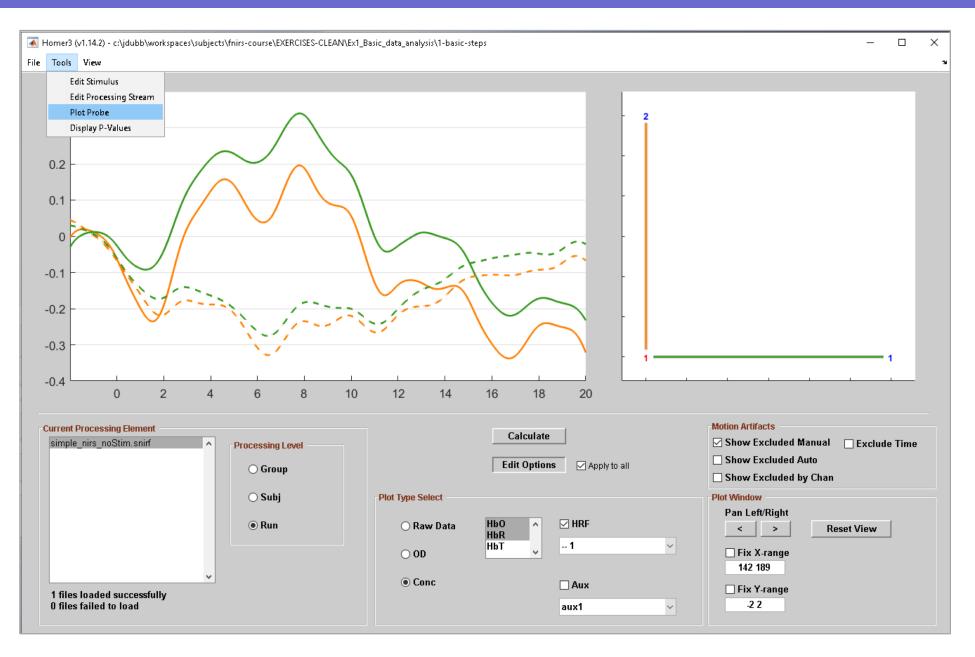


Display options: waterfall

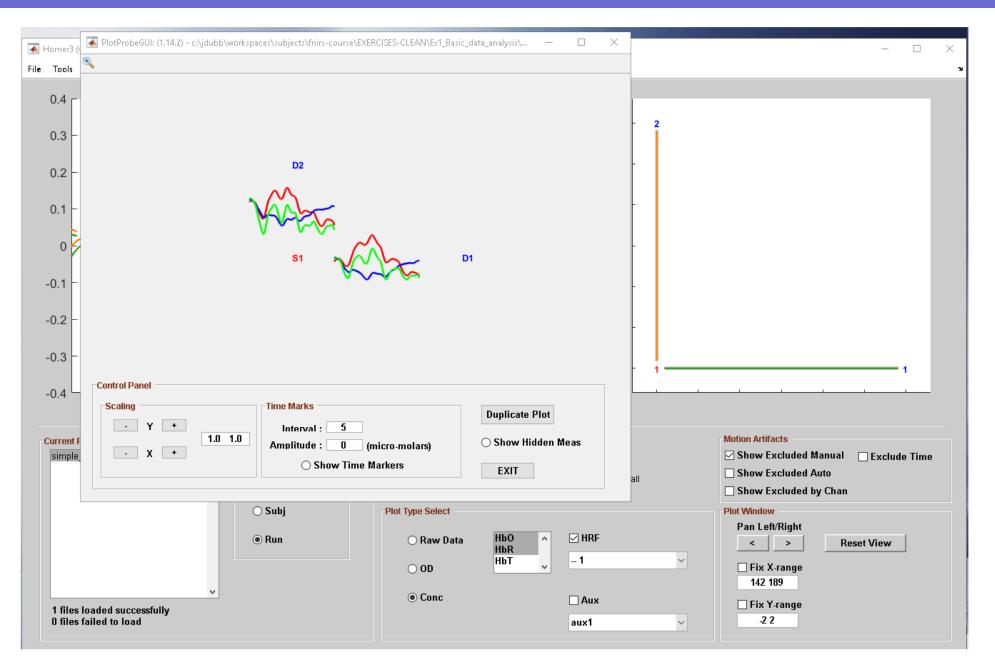
TO-DO, coming soon: Waterfall display of channels as shown below



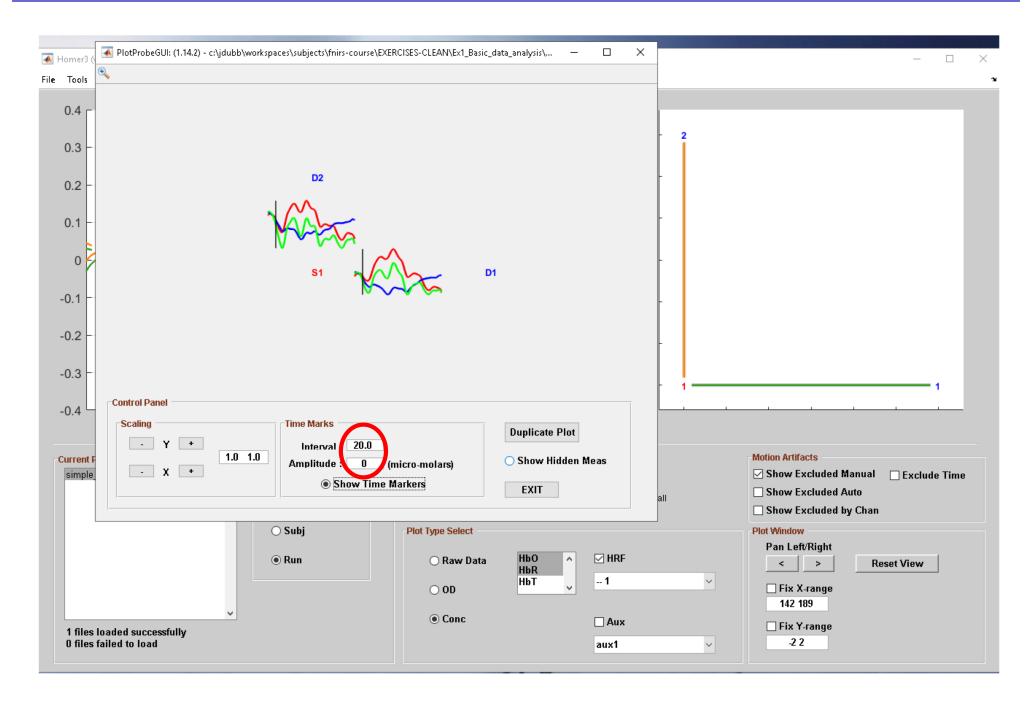
Visualize all HRFs: the Plot Probe feature



Visualize all HRFs: the Plot Probe feature



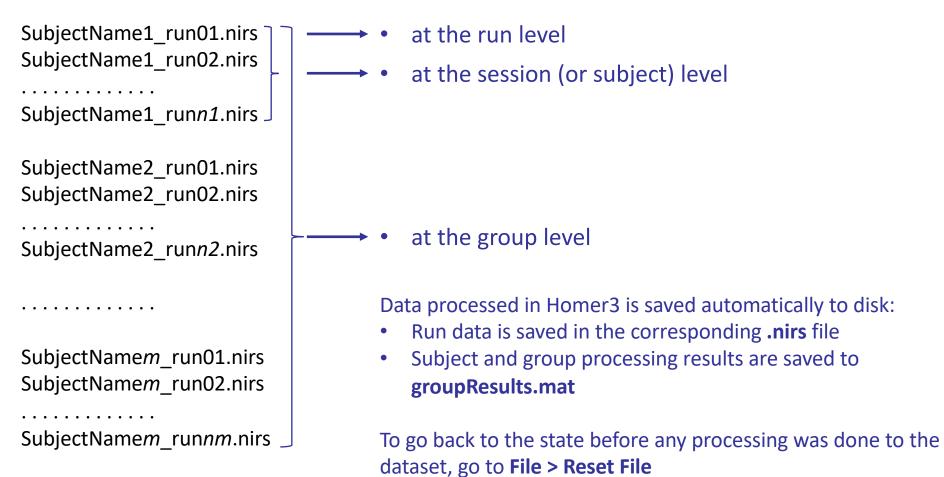
Visualize all HRFs: the Plot Probe feature



Group average: file names

Any directory that contains one or more files with the .nirs extension is considered by Homer3 to be a **group**, organized into sub-groups called **sessions** (or subjects).

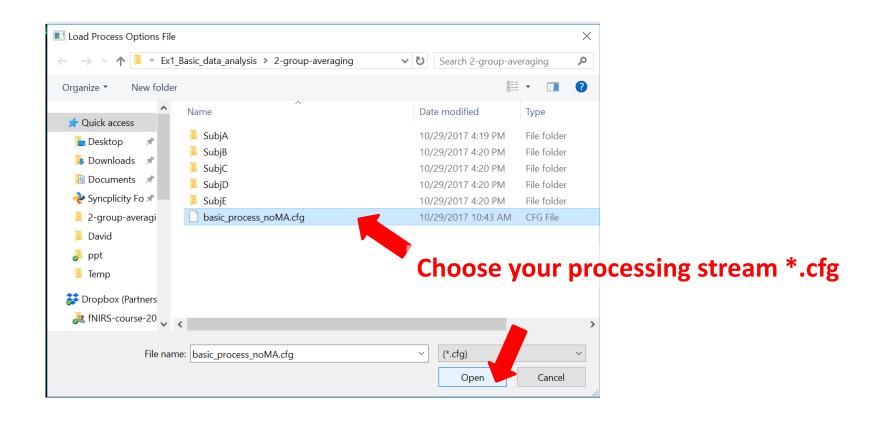
You can compute the HRF:

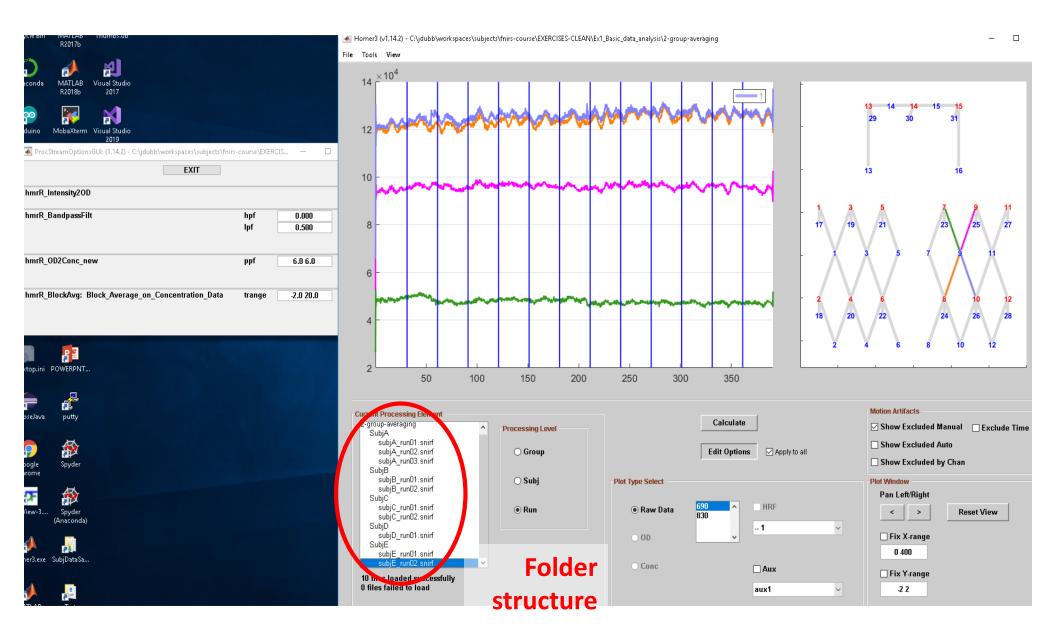


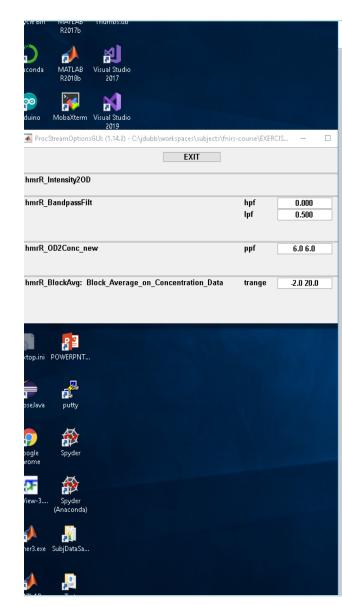
File > Change directory to Exercises / Ex1_Basic_data_analysis / 2-group-averaging

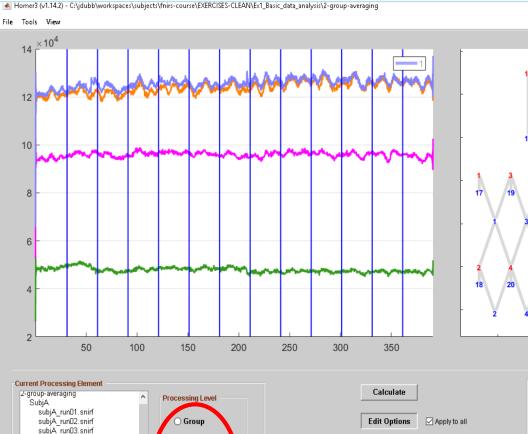
Homer3 (v1.14.2) - c:\jdubb\workspaces\subjects\	fnirs-course\EXERCISES-CLEAN\Ex1_Basi	ic_data_analysis\1-basic-steps				- 🗆 X
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← → ✓ ↑ 📙 « fnirs-course → EXERCISES	-CLEAN » Ex1_Basic_data_analysis »	✓ Ö Search Ex1_E	Basic_data_analysis	<i>۹</i>		
Organize 🔻 New folder				•	2	
🧞 fNIRS_workshop ^ Name	^ Date modif	ied Type	Size			
ProcStream 🧖 1-basic-steps	11/3/2019 6			-		
Snirf 🥏 2-group-averagir	ng 11/4/2019 6	32 AM File folder				
 OneDrive 				-		
💻 This PC						
🗊 3D Objects				-		
E. Desktop						
Documents						
- Downloads						
Music						
Pictures Videos						
Windows (C:)						
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RECOVERY (E:)						· _ · _ · _ · _ ·
- Notacyk Y						
Folder: Ex1_Basic_data_analy	sis				-Motion Arti	facts
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×		O Conc	ſ	Aux		Y-range
1 files loaded successfully O files failed to load				aux1		2

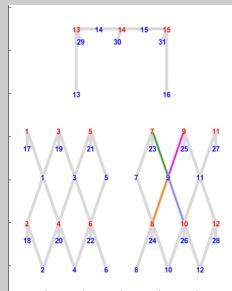
File > Load Process Options (*.cfg)



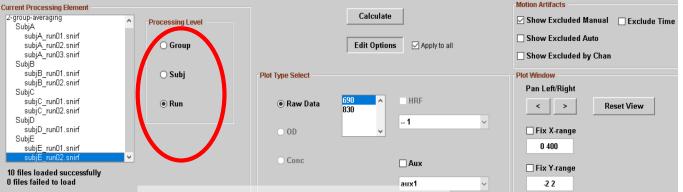






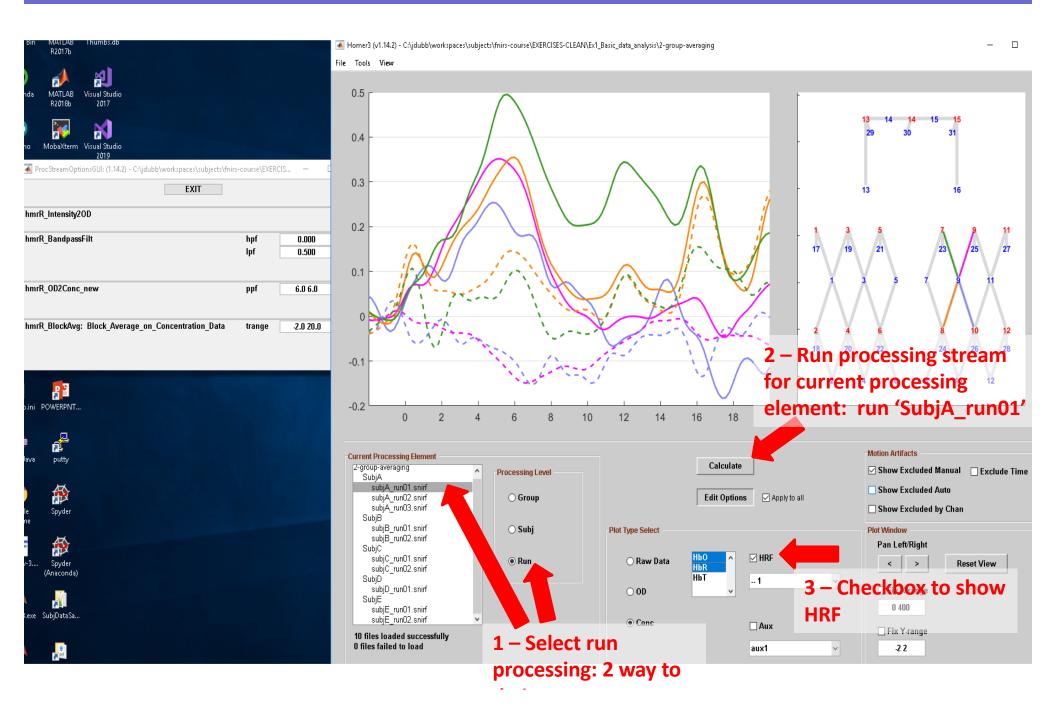


<u>- П</u>

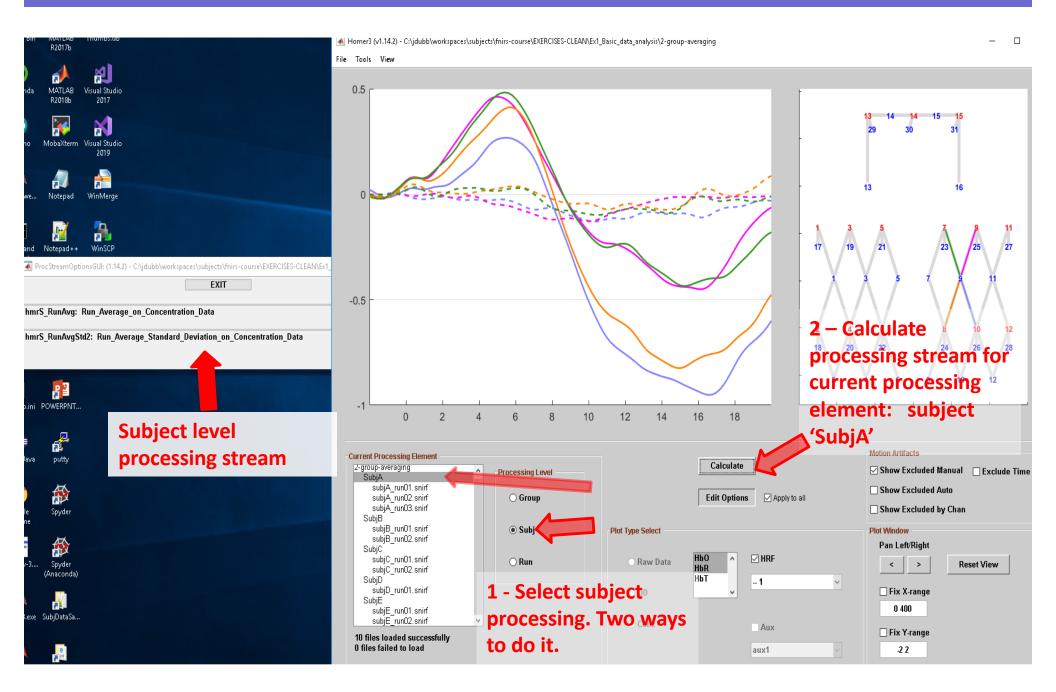


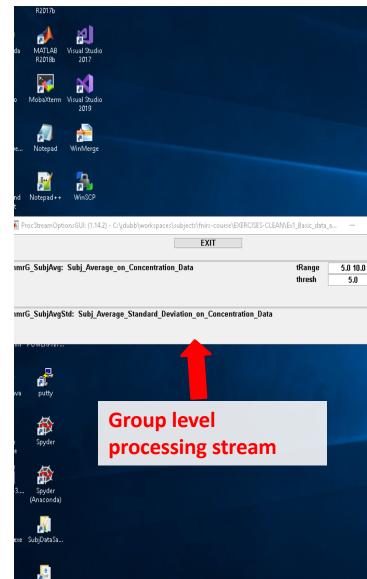
You can run the analysis at the run level, session level, or group level

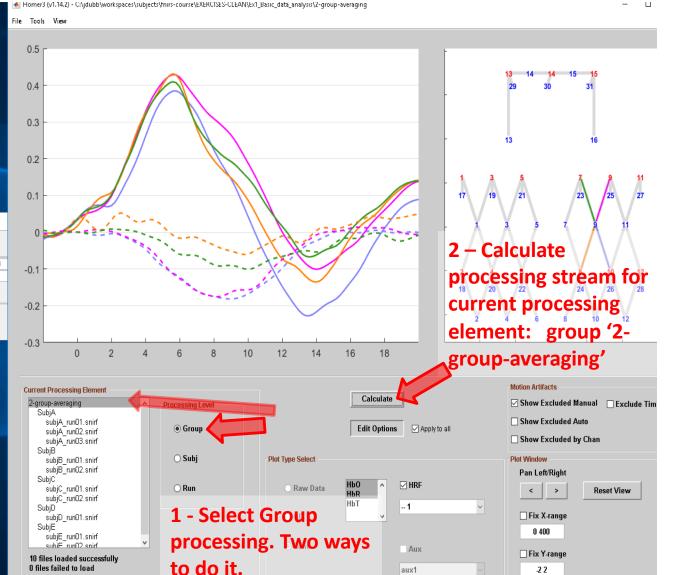
Run average: example

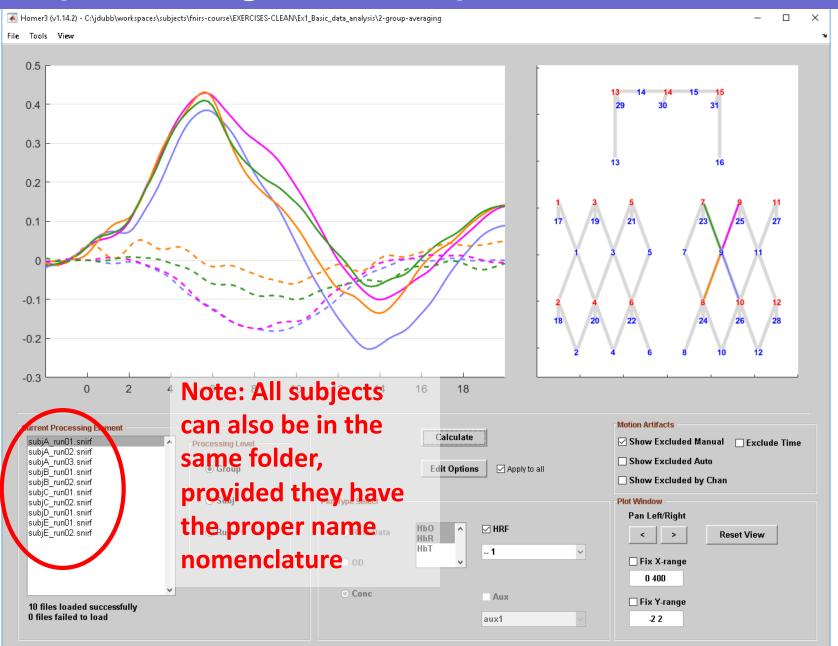


Subject average: example









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We will get back to this dataset tomorrow and perform more advanced data analysis