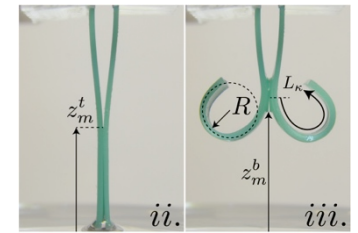
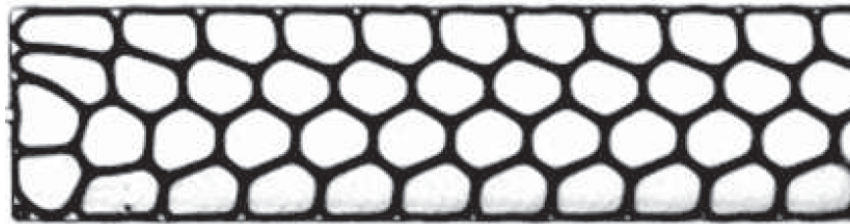
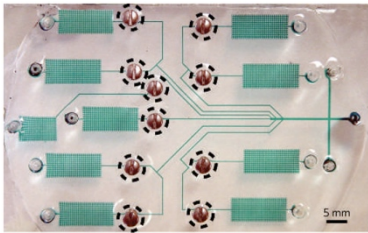


# Confined Fluid Flow: Microfluidics and Capillarity

Douglas P. Holmes

Mechanical Engineering  
Boston University



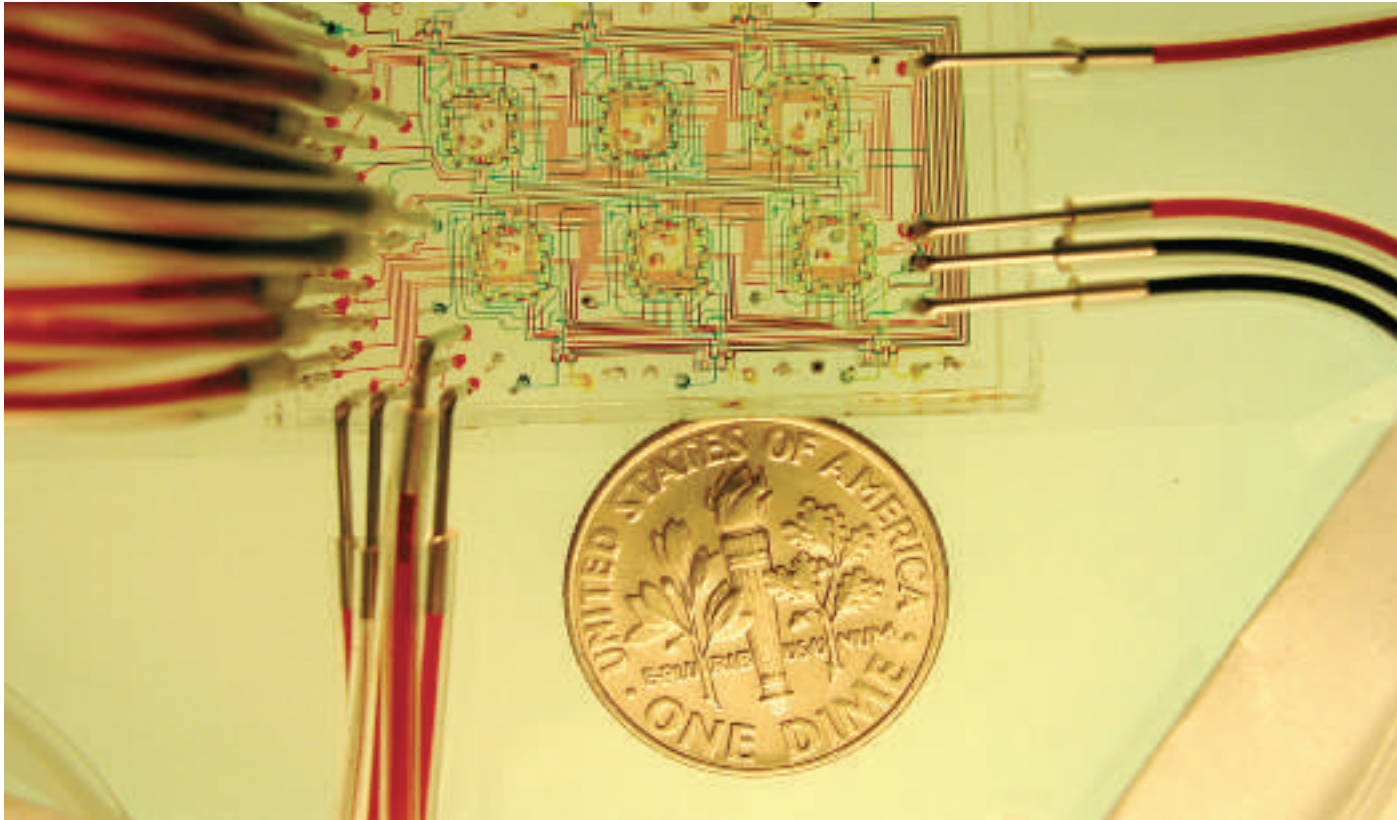


# Why Small?

## Microfluidic chemostat

...static chemical environment used as bioreactor...

fresh medium is continuously added, culture liquid is continuously removed



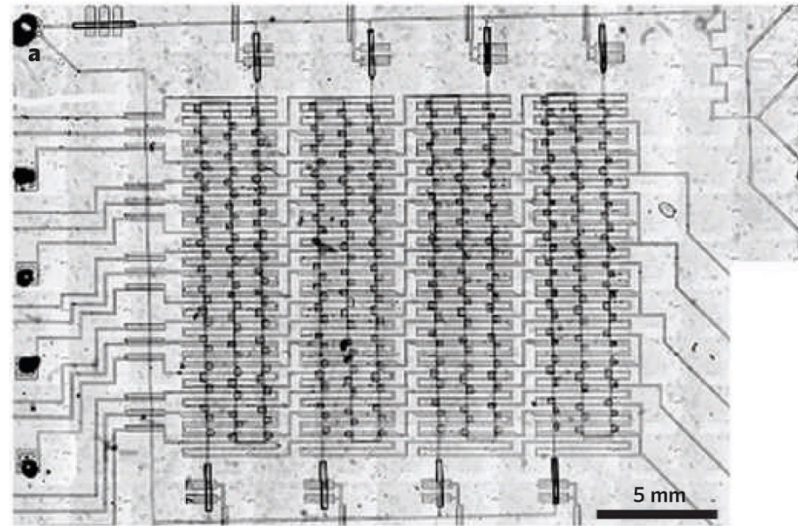
Device provided single-cell resolution to study microbial population growth.



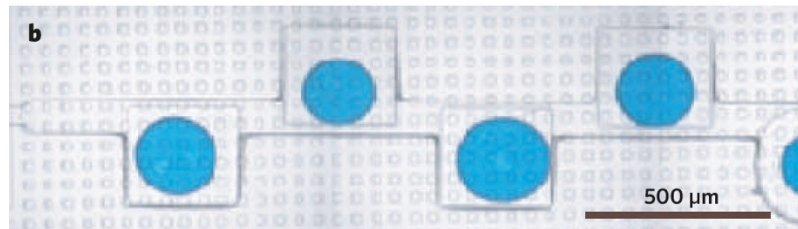


# Why Small?

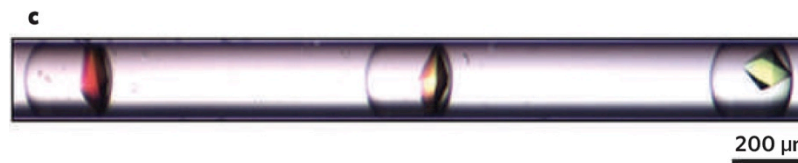
## Protein crystallization



Droplets containing proteins are trapped in microchannel wells.



Dyed droplets in the wells of the device.



Proteins crystallized within droplets in a glass capillary.

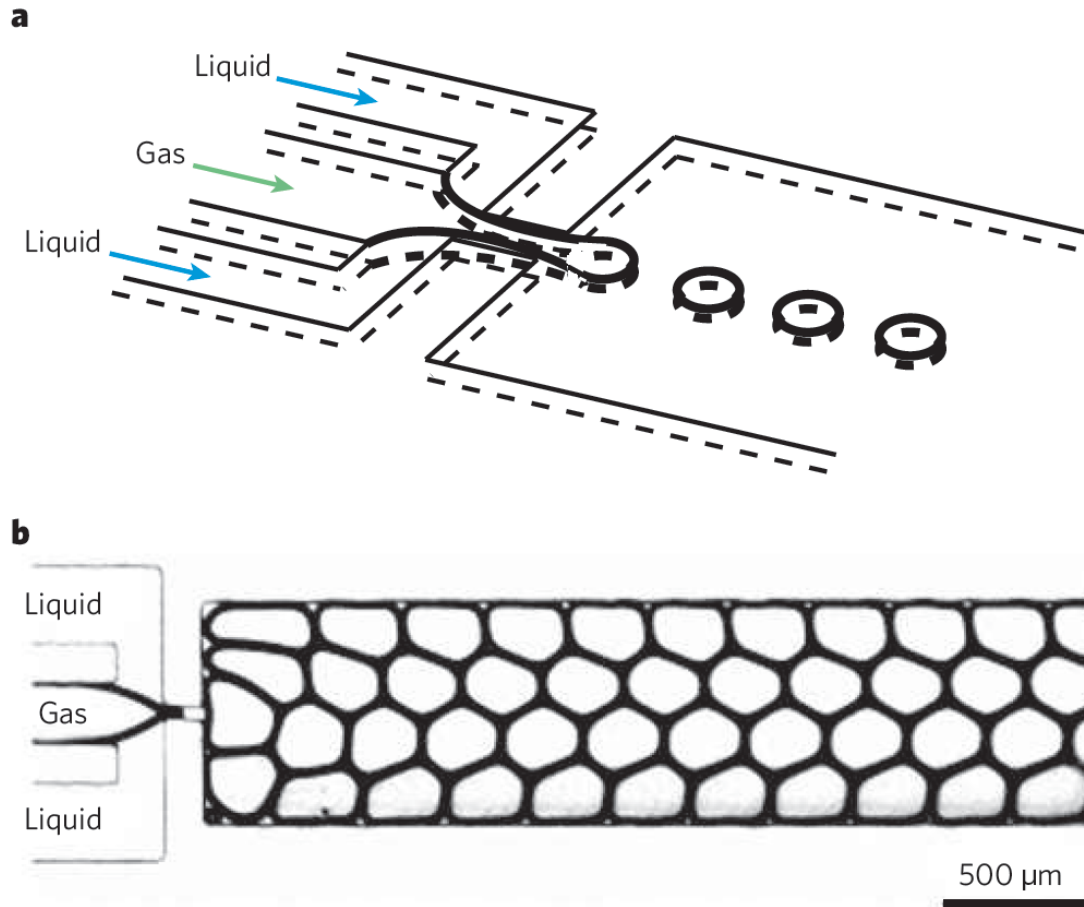
Device used to efficiently screen for optimal protein crystallization conditions

Zheng, Bo, Joshua D. Tice, L. Spencer Roach, and Rustem F. Ismagilov. "A Droplet-Based, Composite PDMS/Glass Capillary Microfluidic System for Evaluating Protein Crystallization Conditions by Microbatch and Vapor-Diffusion Methods with On-Chip X-Ray Diffraction." *Angewandte chemie international edition* 43, no. 19 (2004): 2508-2511.

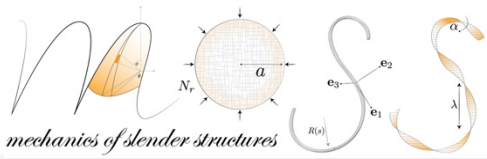
Whitesides, George M. "The origins and the future of microfluidics." *Nature* 442, no. 7101 (2006): 368-373.



## Bubbles and foams

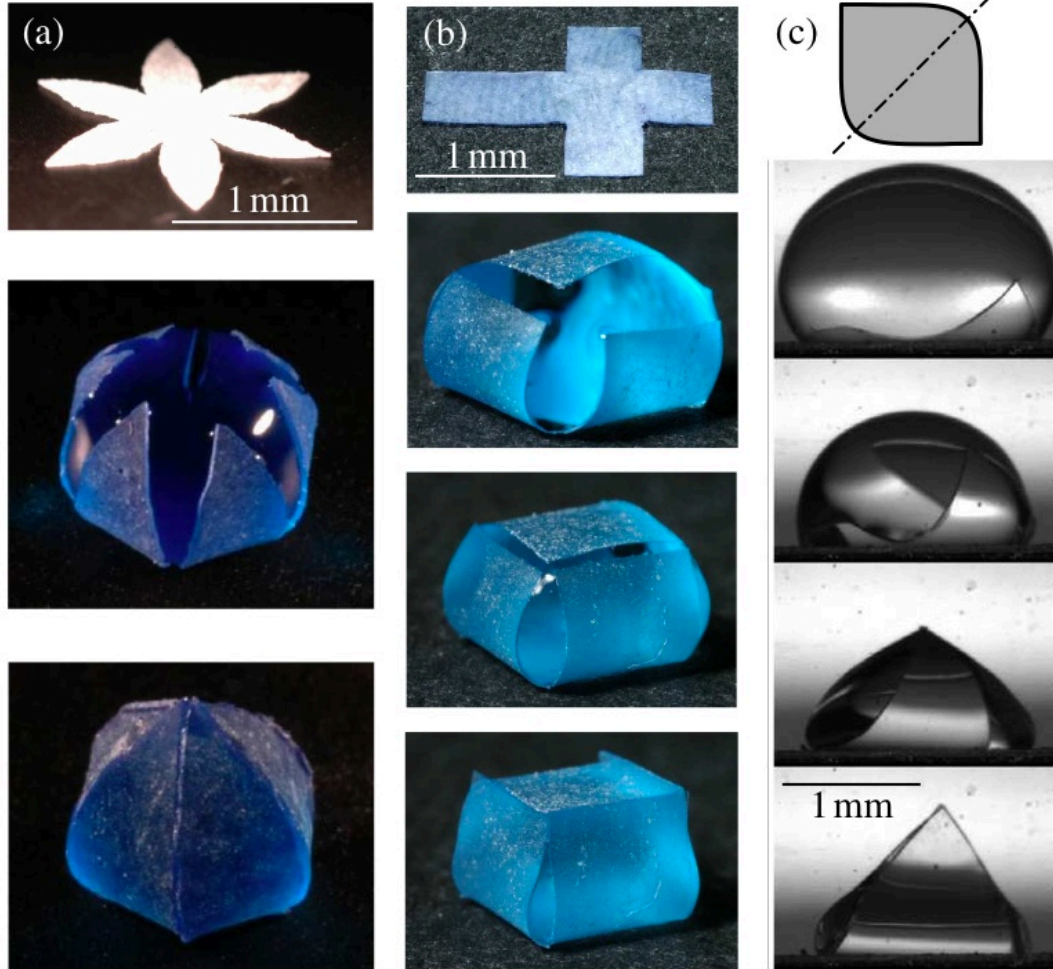


Bubble generation with tunable, monodisperse sizes

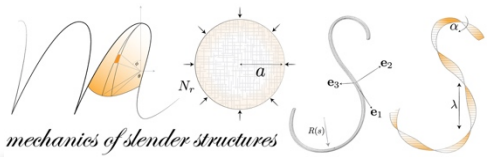


# Why Small?

## Capillary origami



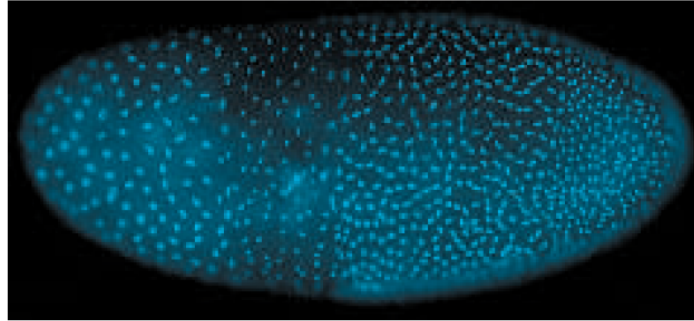
Using surface tension to deform thin structures



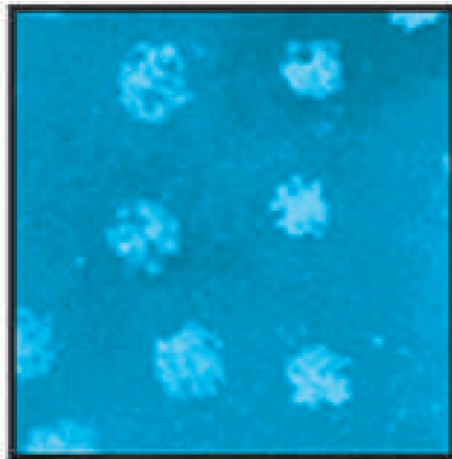
# Why Small?

## Cellular and developmental biology

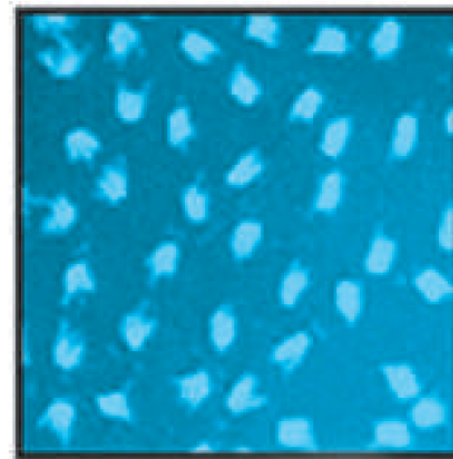
**a**



**b**



**c**



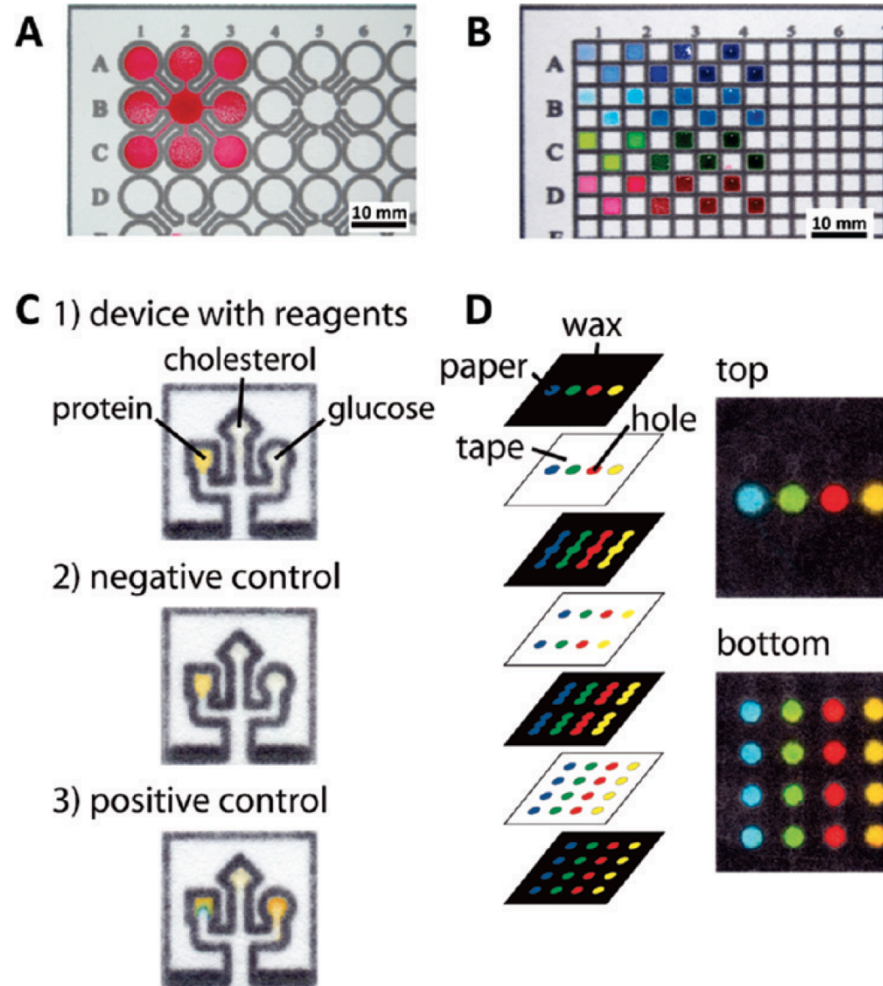
Study effect of temperature on the development of a fruitfly embryo. Immobilized embryo has water with cold (left) and warm (right) flow over it.





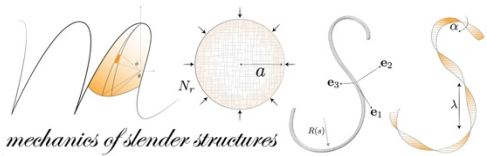
# Why Small?

## Inexpensive diagnostics

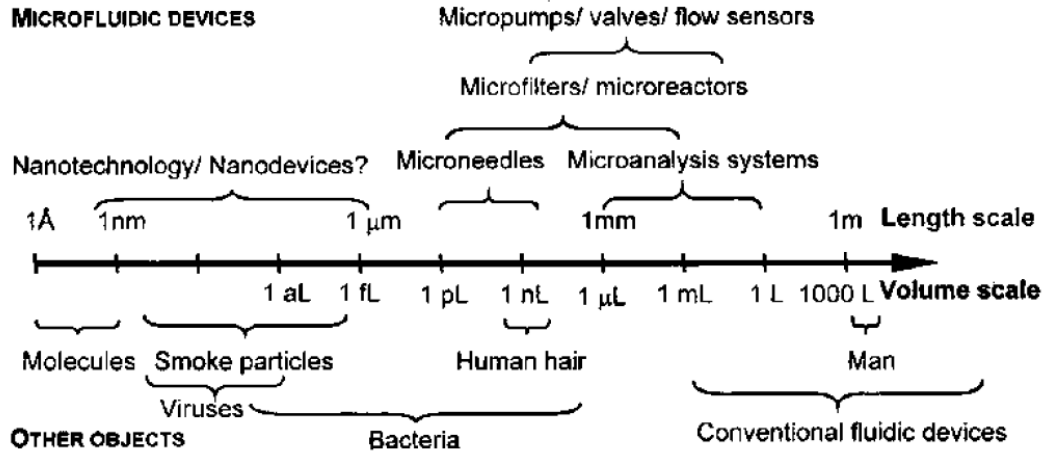


Costs as low as  
\$0.001 per device  
(€0.00089)

Low-cost, simple paper-based microfluidics for diagnostics



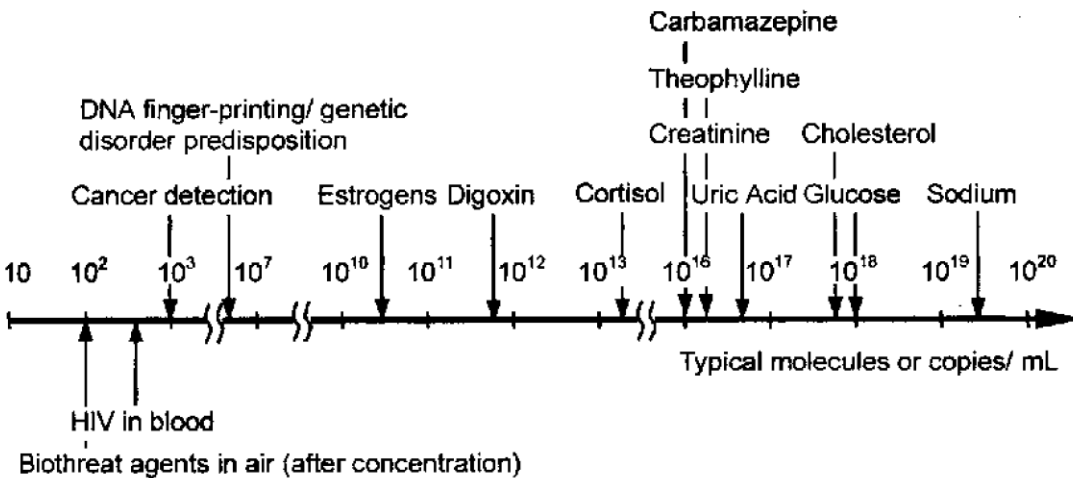
# Why Small?



Volume goes as  $L^3$

- Small decrease in size = large reduction of sample volume.

How much sample volume do we need?



## Detection of biomolecules:

**Digoxin** – heart stimulating drug

**Cortisol** – stress hormone from adrenal gland

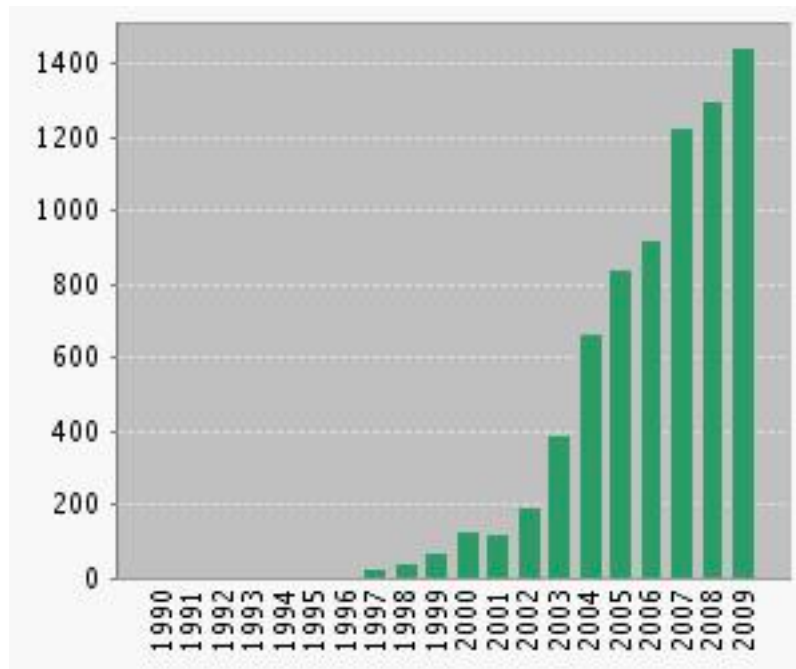
**Creatinine** – level in blood is measure of kidney function

**Theophylline** – drug used to treat respiratory diseases, e.g. asthma

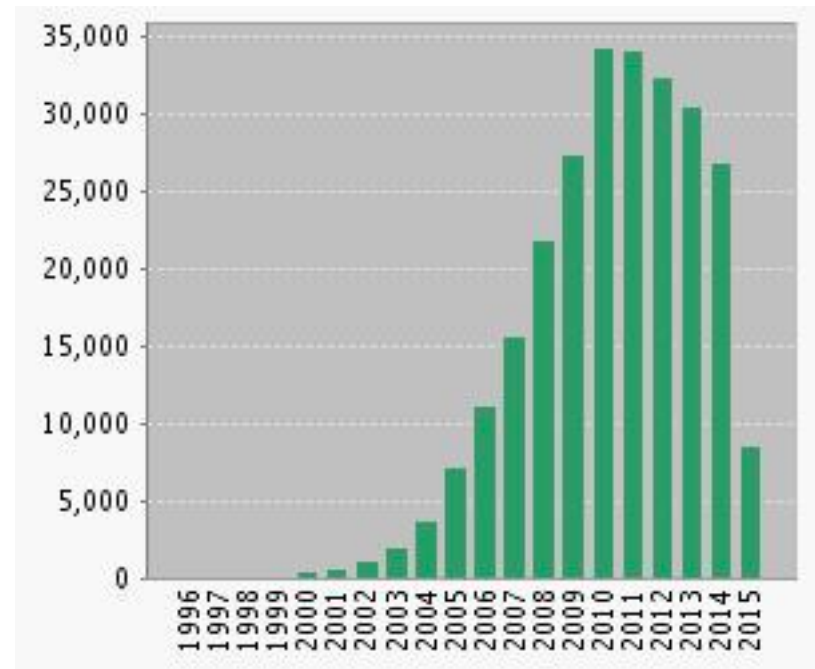


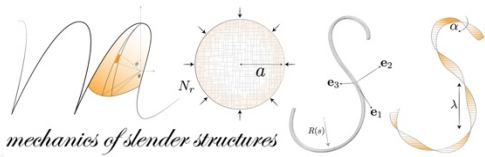
# Microfluidics

Published items in each year...



Citations in each year...





## Confined Fluid Flow: Microfluidics and Capillarity

**Reynolds** Number: Inertia vs. Viscous effects

- Review of characteristic flows...

**Péclet** Number: Transport phenomena in a continuum

- Diffusion, separation, and mixing...

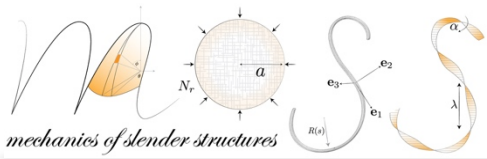
**Geometric** confinement: Controlling and manipulating fluid flow

- Microfluidic fabrication, valving, pumping...

**Capillary** Number: Viscosity vs. Surface tension

- Droplet formation, capillary rise, elasticity...





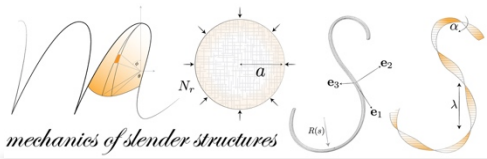
# Fluid Dynamics

## Navier-Stokes Equations (momentum conservation)

$$\underbrace{\rho \left( \underbrace{\frac{\partial \mathbf{u}}{\partial t}}_{\text{variation}} + \underbrace{\mathbf{u} \cdot \nabla \mathbf{u}}_{\text{convection}} \right)}_{\text{Inertial acceleration}} = \underbrace{\left( \underbrace{-\nabla p}_{\text{pressure}} + \underbrace{\mu \nabla^2 \mathbf{u}}_{\text{diffusion}} + \underbrace{\mathbf{f}}_{\text{body forces}} \right)}_{\text{Forces}}$$

## Continuity Equation (mass conservation)

$$\frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \mathbf{u}) = 0$$



# Fluid Behavior

## Inertial Forces



Trailing airplane vortices

## Viscous Forces



Coiling honey

**Reynolds Number:** inertial/viscous

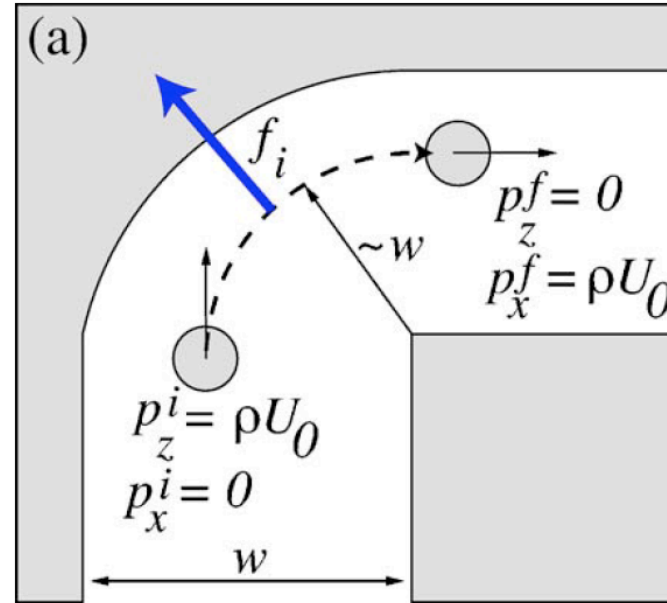
## Inertial Forces



## Viscous Forces



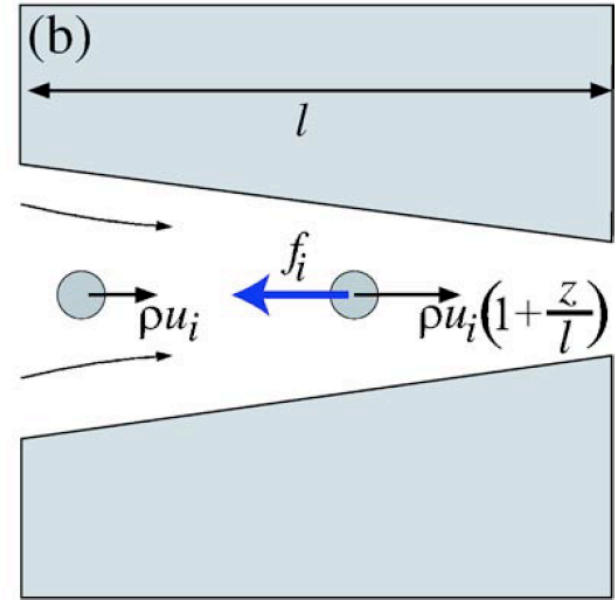
## Reynolds Number: inertial/viscous



Fluid element accelerating around curve.

- During a turn time:  
 $\tau_0 \sim w/U_0$
- Loss of momentum density:  
 $\rho U_0$
- By exerting an inertial centrifugal force density:

$$f_i \sim \rho U_0 / \tau_0 = \rho U_0^2 / w$$



Fluid element in a channel of contracting length.

- By mass conservation, velocity increases as:  
 $u \sim U_0(1 + z/l)$
- Gain momentum at a rate:

$$f_i \sim \rho \frac{du}{dt} = \rho U_0 \frac{du}{dz} \sim \frac{\rho U_0^2}{l}$$

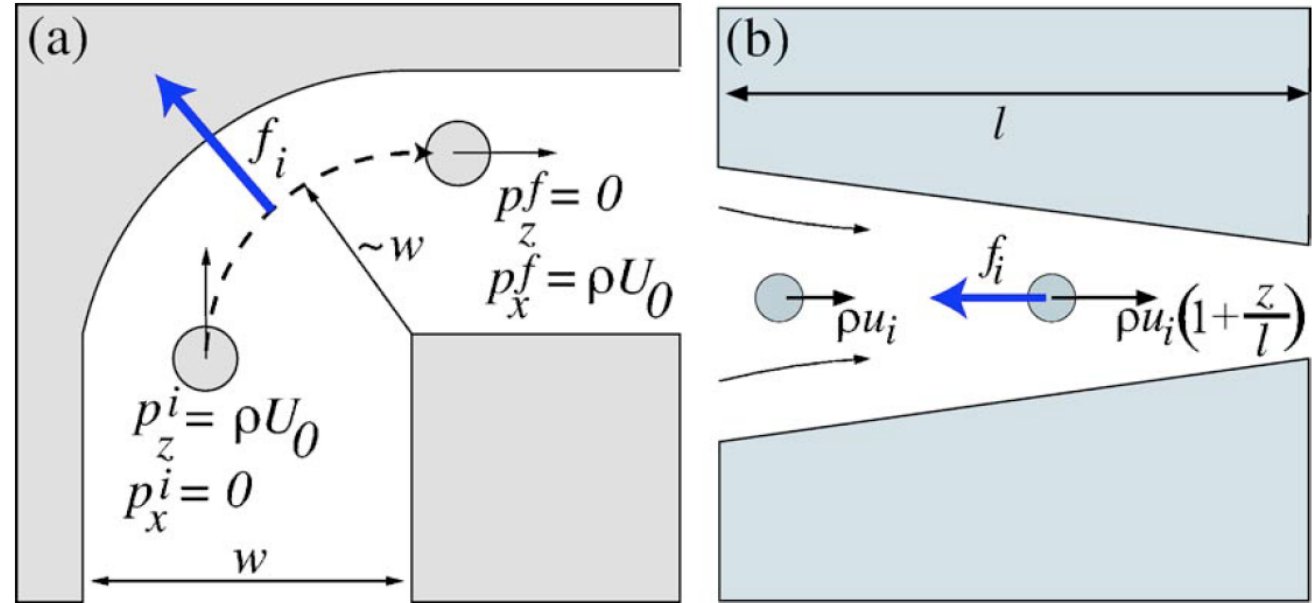
## Inertial Forces



## Viscous Forces



## Reynolds Number: inertial/viscous



Inertial Forces  $f_i \sim \rho U_0 / \tau_0 = \rho U_0^2 / w$

$$f_i \sim \rho \frac{du}{dt} = \rho U_0 \frac{du}{dz} \sim \frac{\rho U_0^2}{l}$$

## Viscous Forces

- Viscous force densities result from gradients in viscous stress:  $f_v \sim \mu U_0 / L_0^2$

Ratio of these two force densities is the **Reynolds number**:

$$\frac{f_i}{f_v} = \frac{\rho U_0 L_0}{\mu} \equiv \mathcal{R}$$





# Fluid Behavior

## Inertial Forces



## Viscous Forces



**Reynolds Number:** inertial/viscous  $\frac{f_i}{f_v} = \frac{\rho U_0 L_0}{\mu} \equiv \mathcal{R}$

Estimation of Reynolds numbers for common microfluidic devices.

- Typical fluid – water
  - Viscosity: 1.025 cP @ 25°C
  - Density: 1 g/mL
- Typical channel dimensions
  - Radius/height (smaller than width): 1 – 100  $\mu\text{m}$
- Typical velocities
  - Average velocity: 1  $\mu\text{m/s}$  – 1 cm/s

## Typical Reynolds number:

$$\mathcal{R} \sim \mathcal{O}(10^{-6}) - \mathcal{O}(10^1)$$

Low Reynolds number: viscous forces > inertial forces

- Flows are **linear**.
- Nonlinear terms in Navier-Stokes disappear
  - Linear, predictable **Stokes flow**



# Fluid Behavior

## Inertial Forces



## Reynolds Number: inertial/viscous

Inertial acceleration

$$\rho \underbrace{\left( \frac{\partial \mathbf{u}}{\partial t} + \mathbf{u} \cdot \nabla \mathbf{u} \right)}_{\text{variation}} = \underbrace{-\nabla p + \underbrace{\mu \nabla^2 \mathbf{u}}_{\text{diffusion}} + \mathbf{f}}_{\text{Forces}}$$

Reynolds number is focused on **steady inertial forcing** due to the convective derivative:

- Almost always unimportant in microfluidic flows.
- Time dependence arises from  $U_0$

$$\rho \mathbf{u} \cdot \nabla \mathbf{u}$$

## Viscous Forces



**Linear unsteady term** sets the inertial time scale to **establish steady flows**:

$$\rho \partial \mathbf{u} / \partial t$$

Time scale estimated by balancing unsteady **inertial force density** with **viscous force density**

$$f_u \sim \rho U_0 / \tau_i$$

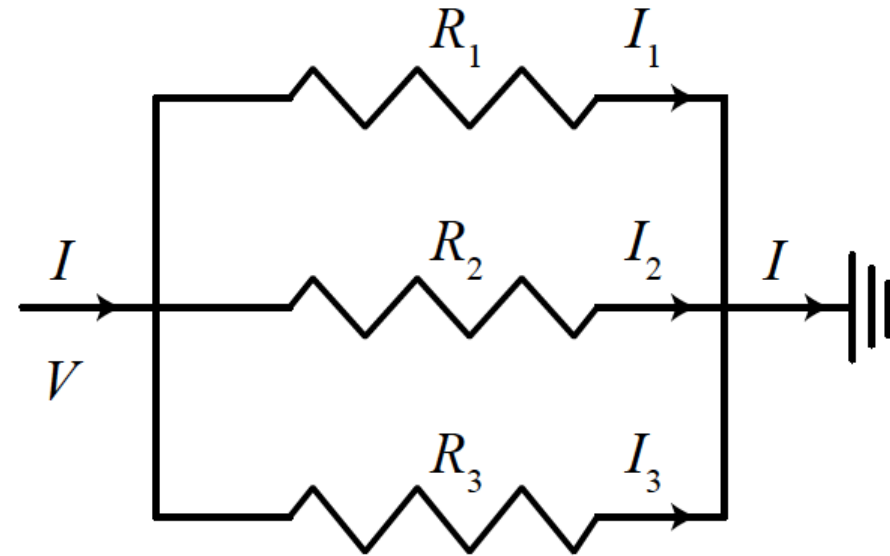
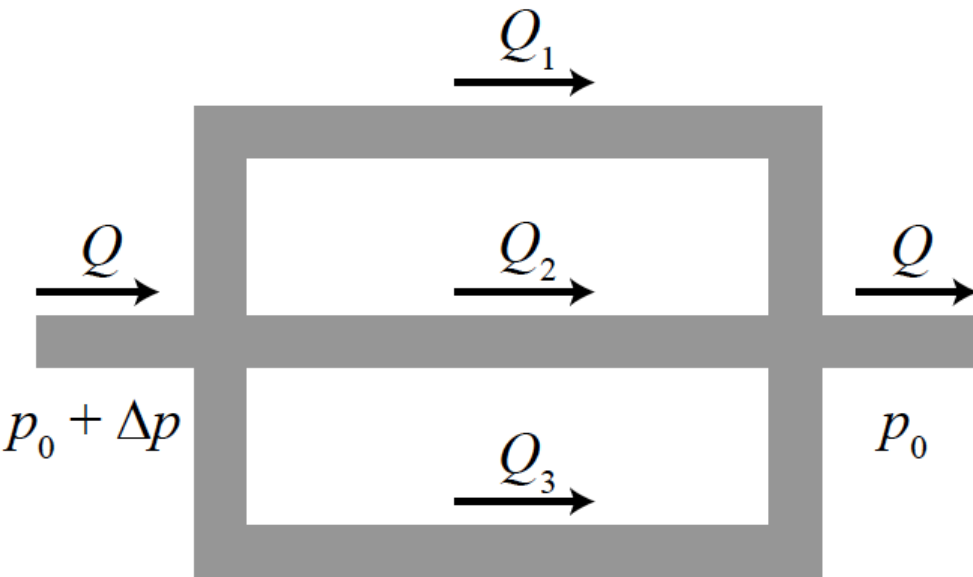
$$f_v \sim \mu U_0 / L_0^2$$

$$\tau_i \sim \frac{\rho L_0^2}{\mu}$$

Time required for a vorticity to diffuse a distance  $L_0$ , with a diffusivity  $\nu = \mu / \rho$ ,  $\tau_i \sim 10 \text{ms}$



# Fluids and Circuits



Pressure-driven flow in a network of parallel channels.

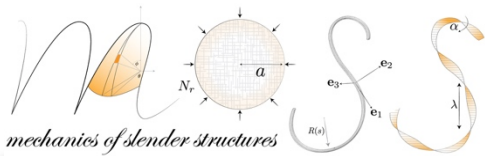
- Flow rates  $Q_i$  in three parallel channels.
- Total flow rate:  $Q = Q_1 + Q_2 + Q_3$
- Pressure drop and flow rate related by:

$$\Delta p = QR_H$$

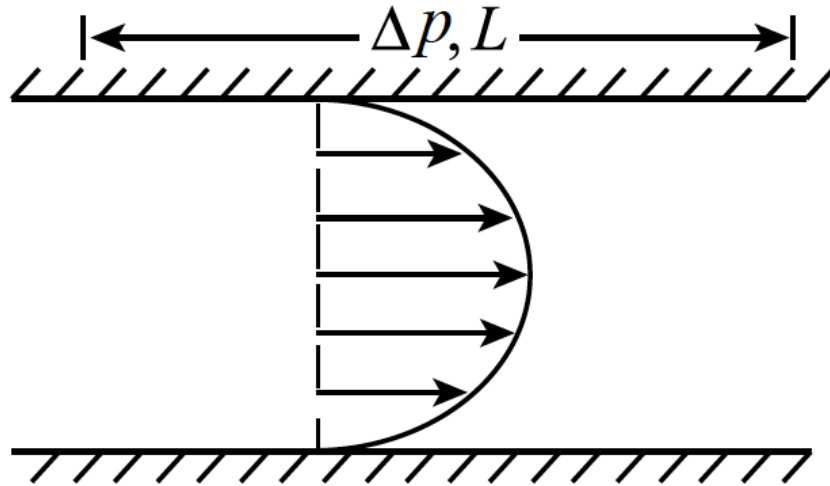
Electrical circuit with resistances in parallel.

- First elementary rule of circuit design is Ohm's law
- Relates electrical potential to current:

$$\Delta V = IR$$



# Flow: Cylindrical Tube



Poiseuille flow

Assumptions:

1. Neglect gravity:  $g_z = 0$
2. Steady state:  $\frac{\partial(\cdot)}{\partial t} = 0$
3. Axisymmetric:  $\frac{\partial(\cdot)}{\partial \theta} = 0$

Continuity Equation (cylindrical coords):

$$\frac{1}{r} \frac{\partial(ru_r)}{\partial r} + \frac{1}{r} \frac{\partial(ru_\theta)}{\partial \theta} + \frac{\partial u_z}{\partial z} = 0$$

Flow only in z direction:  $\frac{\partial u_z}{\partial z} = 0$

Navier-Stokes equations:

$$\underbrace{\rho \left( \frac{\partial \mathbf{u}}{\partial t} + \mathbf{u} \cdot \nabla \mathbf{u} \right)}_{\text{Inertial acceleration}} = \underbrace{-\nabla p + \mu \nabla^2 \mathbf{u} + \mathbf{f}}_{\text{Forces}}$$

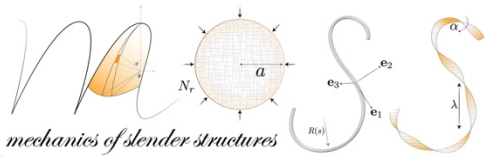
Reduces to the following:

$$0 = -\frac{\partial p}{\partial z} + \mu \frac{1}{r} \frac{d}{dr} \left( r \frac{du_z}{dr} \right)$$

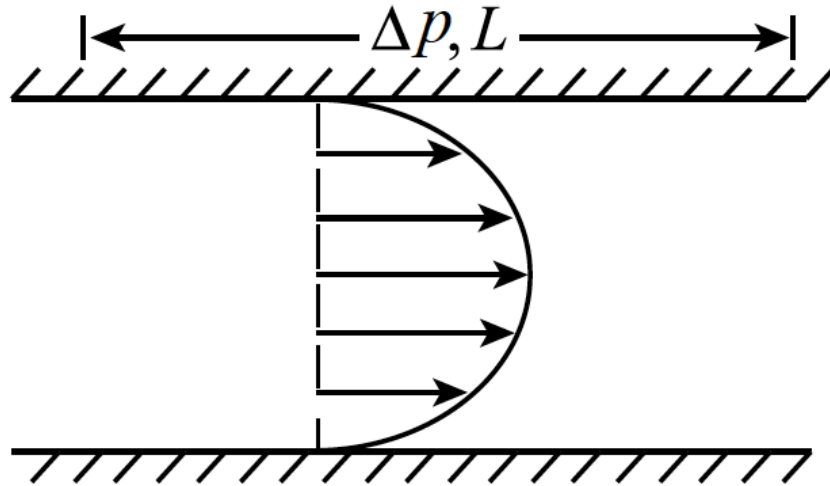
Pressure driven flow – linear change

$$\frac{\partial p}{\partial z} = \frac{p_2 - p_1}{L} = \frac{\Delta p}{L}$$





# Flow: Cylindrical Tube



Poiseuille flow

Boundary conditions:

1. Velocity is at a maximum at the center of the pipe:

$$\frac{du_z}{dr} = 0 \quad @ \quad r = 0$$

2. No-slip along the walls.

$$u_z = 0 \quad @ \quad r = R$$

Navier-Stokes equations - cylindrical pipe:

$$0 = -\frac{\partial p}{\partial z} + \mu \frac{1}{r} \frac{d}{dr} \left( r \frac{du_z}{dr} \right)$$

$$\int_0^r d \left( r \frac{du_z}{dr} \right) = \frac{1}{\mu} \frac{\Delta p}{L} \int_0^r r \, dr$$

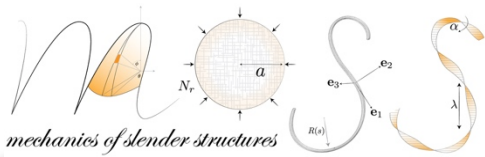
...integrate once...

$$r \frac{du_z}{dr} = \frac{1}{\mu} \frac{\Delta p}{L} \frac{r^2}{2} + C_1$$

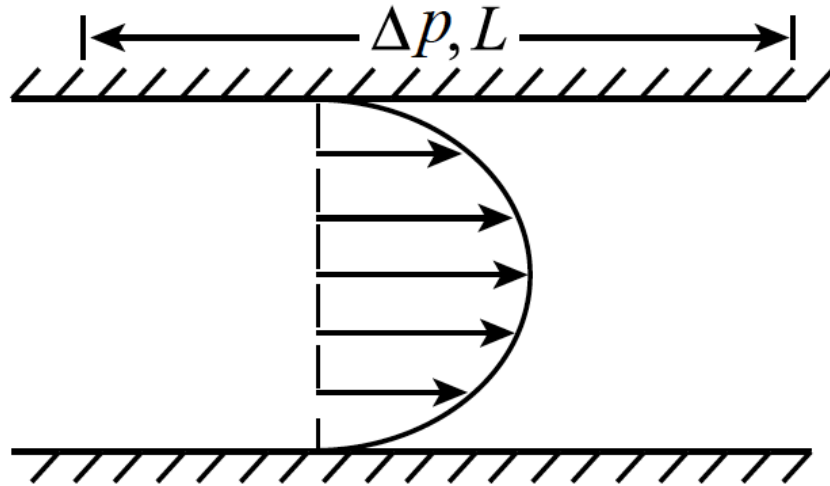
$$\int_0^r du_z = \int_0^r \left( \frac{1}{\mu} \frac{\Delta p}{L} \frac{r}{2} + \frac{C_1}{r} \right) dr$$

...integrate twice...

$$u_z = \frac{1}{\mu} \frac{\Delta p}{L} \frac{r^2}{4} + C_1 \ln(r) + C_2$$



# Flow: Cylindrical Tube



Poiseuille flow

Parabolic velocity distribution:

$$u(r) = \frac{1}{4\mu} \underbrace{\left| \frac{dp}{dz} \right|}_{\text{axial pressure gradient}} (a^2 - r^2)$$

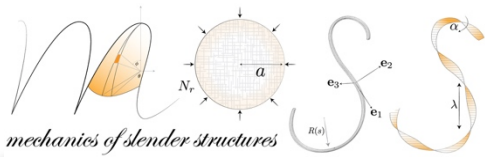
axial pressure gradient

Flow rate (volume/time)

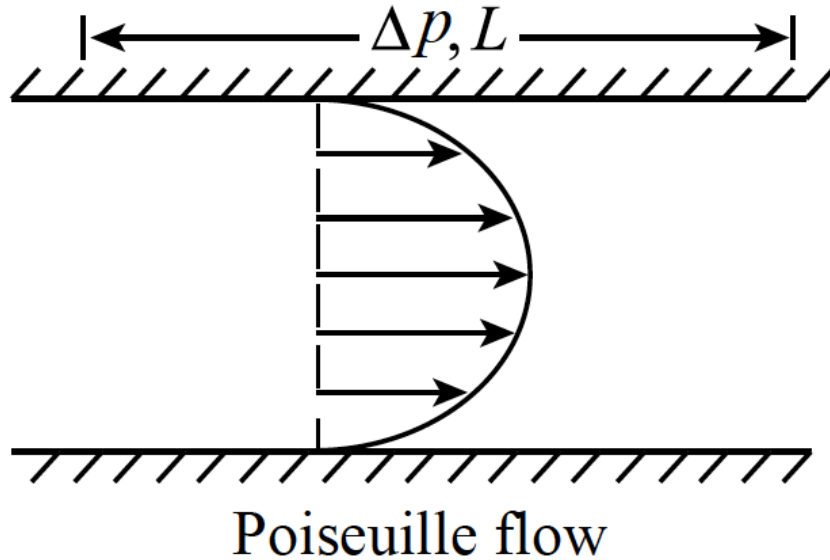
$$Q = 2\pi \int_0^a u(r) r \, dr = \frac{\pi a^4}{8\mu} \left| \frac{dp}{dz} \right|$$

The flow rate depends on the **fourth** power of the radius: important in small systems

- e.g. For the same pressure gradient, reducing the radius by a factor of **two** causes a **16-fold** reduction in flow rate.
- e.g. Consider a blood vessel – a **10% decrease in radius** produces more than a **40% decrease in the flow rate** of blood (PSA: Eat more kale!)



# Flow: Cylindrical Tube



Average velocity

$$U = \frac{Q}{\pi a^2} = \frac{a^2}{8\mu} \left| \frac{dp}{dz} \right|$$

Recall the Reynolds number:

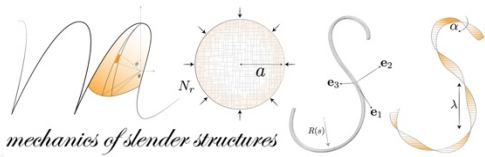
$$\mathcal{R} = \frac{\rho U a}{\mu}$$

For a fixed pressure gradient,  $\mathcal{R} \sim a^3$  and so a **factor of two** change in **radius** produces a **factor of eight** change in **Reynolds number**.

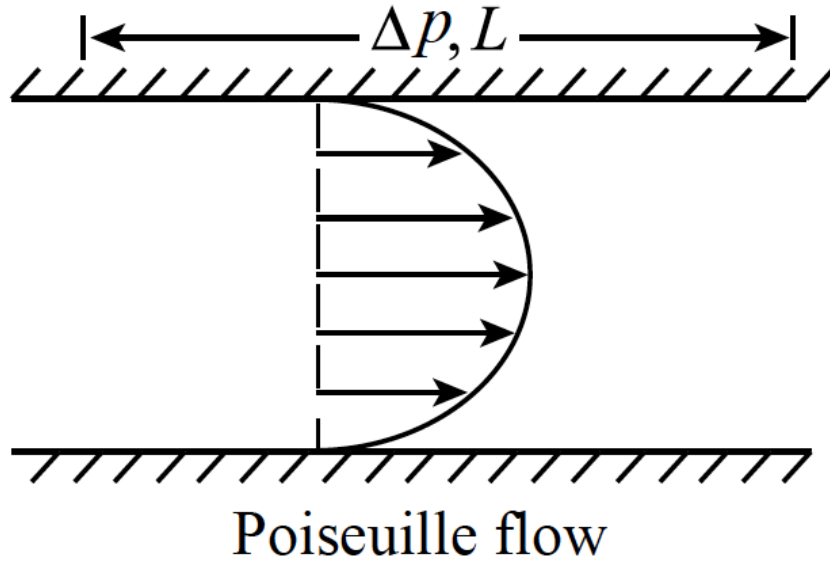
The average velocity is proportional to the pressure gradient, writing the flow field in vector form gives Darcy's law:

$$\frac{\mu U}{k} = -\nabla p$$

- $k$  is the **permeability** (dimensions length<sup>2</sup>)
- linear relation between pressure and velocity is due to **lack of inertia** effects
- order-of-magnitude of permeability is typically the **square of smallest dimension**.



# Flow: Cylindrical Tube



Hydrodynamic resistance

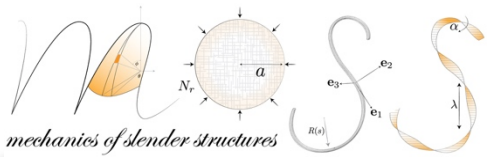
$$R_H = \frac{\Delta p}{Q} = \frac{8\mu L}{\pi a^4} \quad \text{recall: } -\frac{dp}{dz} = \frac{\Delta p}{L}$$

**Electrical analogy:** resistance in fluid channels depends on the **fourth** power of the radius rather than the **second** power in the electrical case.

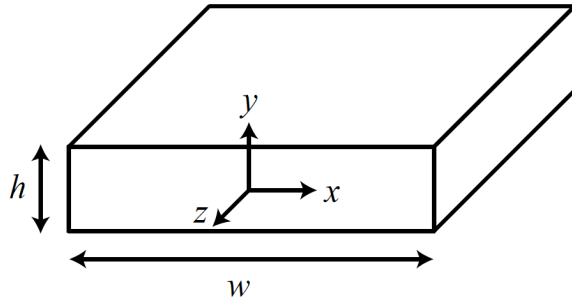
.....small matters!

## Scaling on physical & dimensional arguments...

- Can arrive at:  $u \sim a^2 \Delta p / (\mu L)$  and  $Q \sim ua^2 \sim a^4 \Delta p / (\mu L)$  from dimensional analysis.
- Fluid motion arises as a balance between the pressure drop driving motion and the frictional (viscous) resistance from the bounding walls. This balance applies **independent** of the **cross-sectional channel shape**.



# Flow: Rectangular



Parabolic velocity distribution:

$$u(y) = \frac{\Delta p}{2\mu L} \left[ \left( \frac{h}{2} \right)^2 - y^2 \right]$$

Approximate flow rate (channel of width w)

$$Q = w \int_0^h u(y) dy = \frac{wh^3 \Delta p}{12\mu L}$$

Average velocity:  $k = \frac{h^2}{12}$

Hydrodynamic Resistance:  $R_H = \frac{12\mu L}{wh^3}$

**Parabolic velocity distribution** (known analytically in terms of a Fourier series):

$$u(x, y) = \frac{\Delta p}{2\mu L} \left[ \left( \left[ \frac{h}{2} \right]^2 - y^2 \right) - \sum_{n=0}^{\infty} a_n \cos \left( \frac{\lambda_n y}{h/2} \right) \cosh \left( \frac{\lambda_n x}{h/2} \right) \right]$$

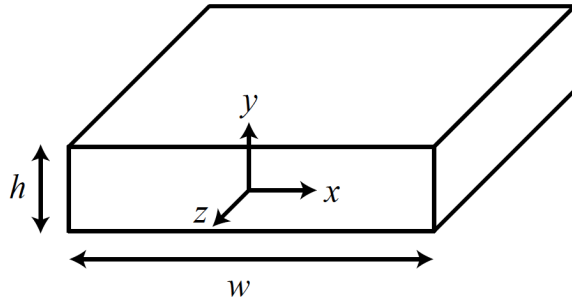
where:  $\lambda_n = \frac{(2n+1)\pi}{2}$

and, from the no-slip boundary conditions:  $a_n = \frac{h^2(-1)^n}{\lambda_n^3 \cosh(\lambda_n w/h)}$





# Flow: Rectangular



Parabolic velocity distribution:

$$u(y) = \frac{\Delta p}{2\mu L} \left[ \left( \frac{h}{2} \right)^2 - y^2 \right]$$

Approximate flow rate (channel of width w)

$$Q = w \int_0^h u(y) dy = \frac{wh^3 \Delta p}{12\mu L}$$

Average velocity:  $k = \frac{h^2}{12}$

Hydrodynamic Resistance:  $R_H = \frac{12\mu L}{wh^3}$

## Corresponding flow rate

$$Q = 4 \int_0^{w/2} \int_0^{h/2} u(x, y) dy dx$$

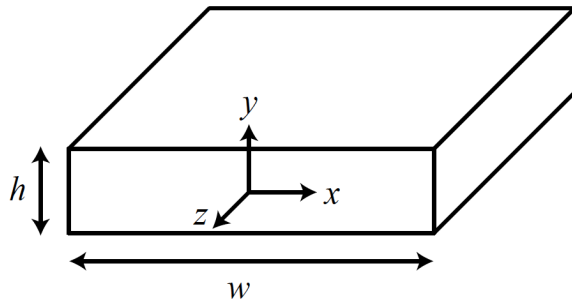
$$Q = \frac{wh^3 \Delta p}{12\mu L} \left[ 1 - 6 \left( \frac{h}{w} \right) \sum_{n=0}^{\infty} \lambda_n^{-5} \tanh \left( \frac{\lambda_n w}{h} \right) \right]$$

where:  $\lambda_n = \frac{(2n+1)\pi}{2}$

Flow rate is nearly linear in aspect ratio  $h/w$



# Flow: Rectangular

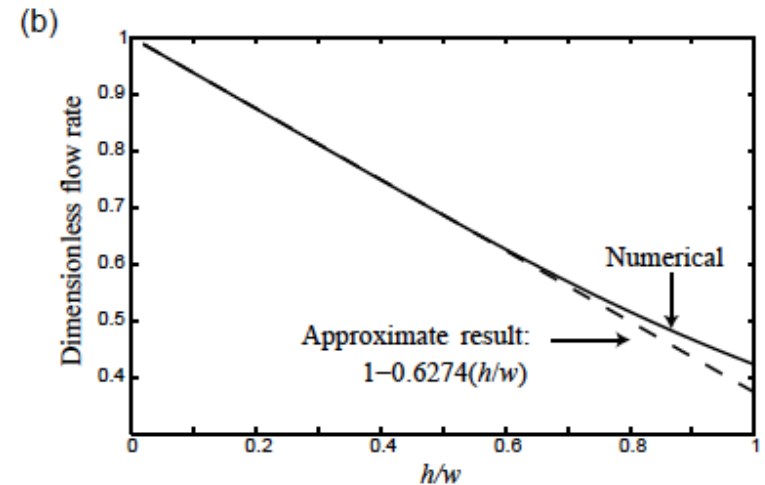
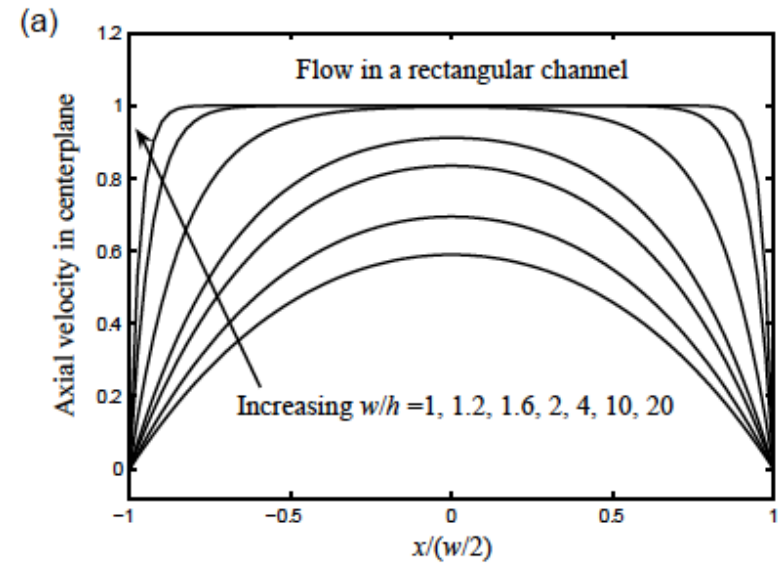


## Flow rate in a rectangular channel

$$Q = \frac{wh^3 \Delta p}{12\mu L} \left[ 1 - 6 \left( \frac{h}{w} \right) \sum_{n=0}^{\infty} \lambda_n^{-5} \tanh \left( \frac{\lambda_n w}{h} \right) \right]$$

## Approximate dimensionless flow rate

$$\underbrace{\frac{12\mu L Q}{wh^3 \Delta p}}_{\text{dimensionless flow rate}} \approx 1 - \frac{6(2^5)}{\pi^5} \underbrace{\frac{h}{w}}_{\text{aspect ratio}}$$





# Workout Problem

While developing a microfluidic chip for your research, you decide a scale model would help you gain intuition about flow characteristics at the microscale. While most scale models are smaller than the original, your model will be 100 times larger (so that you can see it without a microscope).

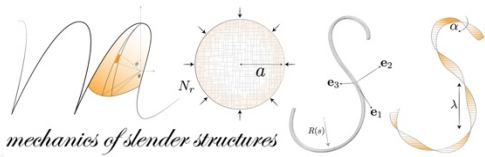
|           | <b>Viscosity</b> | <b>Density</b> |
|-----------|------------------|----------------|
| Honey     | 8750 cP          | 1.42 g/mL      |
| Olive oil | 81 cP            | 0.92 g/mL      |
| Water     | 1.025 cP @ 25°C  | 1 g/mL         |
| Molasses  | 50,000 cP        | 1.50 g/mL      |

For the microfluidic chip, the flow conditions are: fluid velocity 1 mm/s, channel diameter 100  $\mu\text{m}$ , and the liquid is water at  $T = 25^\circ\text{C}$ . The Reynolds number for both systems is to be duplicated, and the microchannel is hemicylindrical (use the hydraulic diameter). For the scale model, which of the condiments listed above from your kitchen would you choose as the flow liquid, if the velocity in the model is:

- a.) 1 mm/s
- b.) 60 mm/s
- c.) 340 mm/s

Recall: hydraulic diameter,  $D_H$ , relates flow in noncircular channels to those in a circular channel:

$$D_H = \frac{4A}{P}$$



# Workout Problem

**Hydraulic diameter of hemicylindrical channel:**

$$D_H = \frac{4A}{P} = \frac{2\pi r^2}{\pi r + 2r} = 61.1 \mu\text{m}$$

**Reynold's number of microfluidic chip:**

$$\mathcal{R} = \frac{\rho U D_H}{\mu} = (1000 \text{ kg/m}^3) (0.001 \text{ m/s}) (61.1 \times 10^{-6} \text{ m}) (1.025 \times 10^{-3} \text{ Pa} \cdot \text{s})^{-1}$$

$$\mathcal{R} = 0.0596$$

**a. Fluid flowing at 1 mm/s in scaled up model**

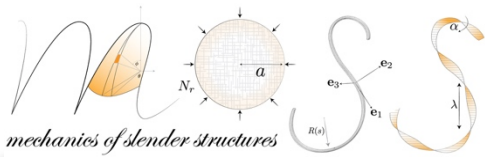
Use Olive Oil.  $\mathcal{R} = (920 \text{ kg/m}^3) (0.001 \text{ m/s}) \underbrace{(6.11 \times 10^{-3} \text{ m})}_{100 \text{ times larger}} (81 \times 10^{-3} \text{ Pa} \cdot \text{s})^{-1} = 0.0694$

**b. Fluid flowing at 60 mm/s in scaled up model**

Use Honey.  $\mathcal{R} = (1420 \text{ kg/m}^3) \underbrace{(0.06 \text{ m/s})}_{60 \text{ times larger}} (6.11 \times 10^{-3} \text{ m}) (8750 \times 10^{-3} \text{ Pa} \cdot \text{s})^{-1} = 0.0595$

**c. Fluid flowing at 340 mm/s in scaled up model**

Use Molasses.  $\mathcal{R} = (1500 \text{ kg/m}^3) \underbrace{(0.34 \text{ m/s})}_{340 \text{ times larger}} (6.11 \times 10^{-3} \text{ m}) (50 \text{ Pa} \cdot \text{s})^{-1} = 0.0620$



## Confined Fluid Flow: Microfluidics and Capillarity

**Reynolds** Number: Inertia vs. Viscous effects

- Review of characteristic flows...

**Péclet** Number: Transport phenomena in a continuum

- Diffusion, separation, and mixing...

**Geometric** confinement: Controlling and manipulating fluid flow

- Microfluidic fabrication, valving, pumping...

**Capillary** Number: Viscosity vs. Surface tension

- Droplet formation, capillary rise, elasticity...

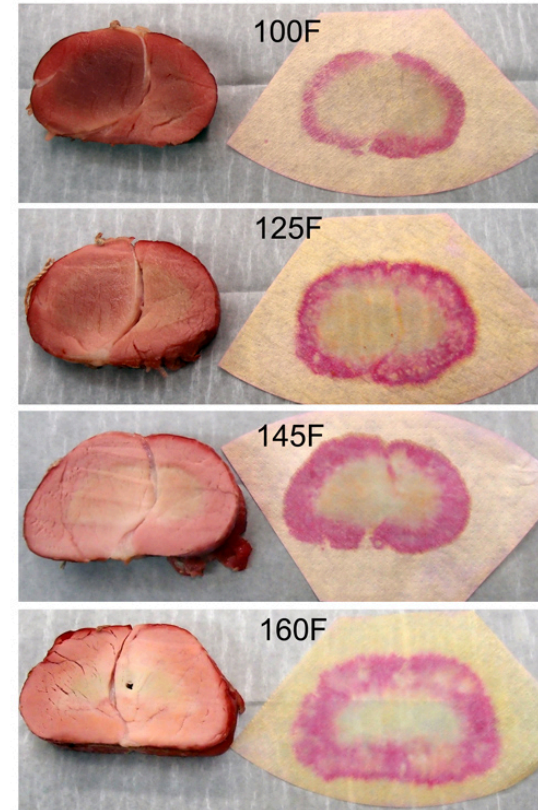


## Convection



Convection current as hot water mixes with cold water

## Diffusion



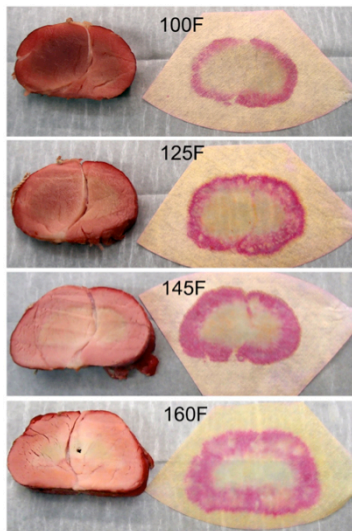
Diffusion of salt ions while curing pork

**Péclet Number:** convection/diffusion

## Convection



## Diffusion



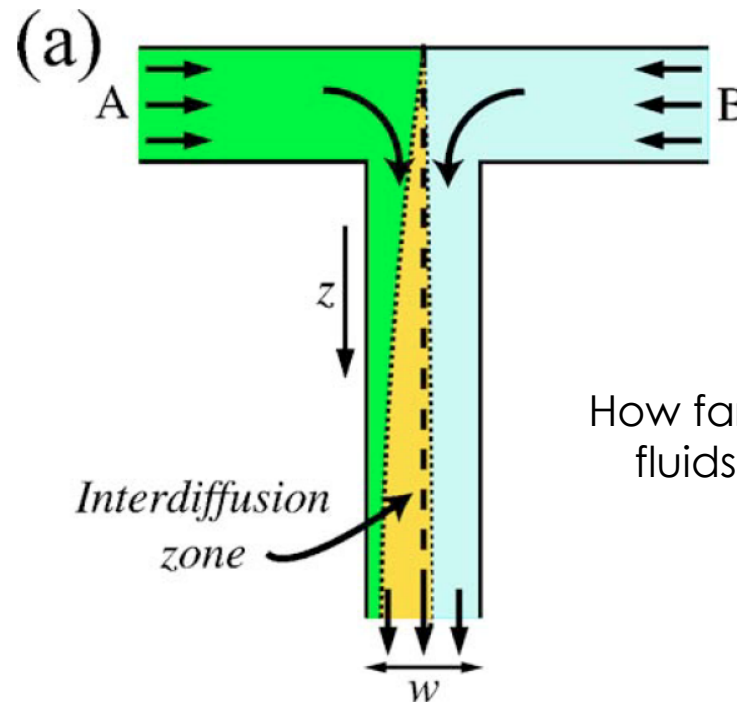
## Péclet Number: convection/diffusion

High Reynolds: eddies chaotically stretch and fold fluid elements.

- Turbulent mixing & thermal convection move fluids.

Low Reynolds: mixing occurs by diffusion only

- Diffusion is remarkably slow – long mixing times.



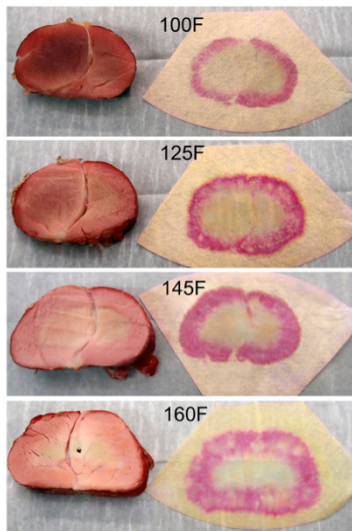
**T-Junction** in which two fluids are injected and flow alongside each other

How far down the channel must the fluids flow before the channel is homogenized?

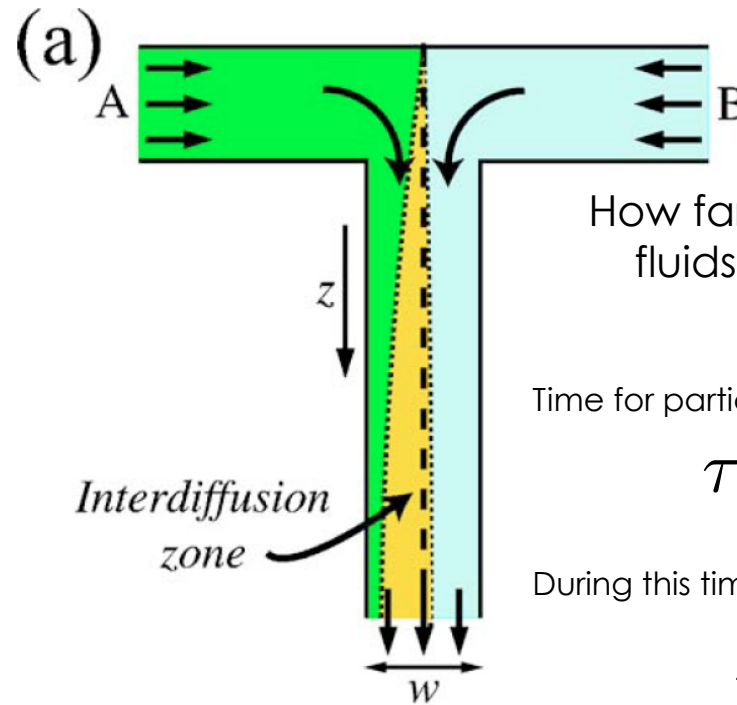
## Convection



## Diffusion



## Péclet Number: convection/diffusion



How far down the channel must the fluids flow before the channel is homogenized?

Time for particles/molecules to diffuse across channel:

$$\tau_D \sim w^2 / D$$

During this time, the stripe moves down the channel:

$$Z \sim U_0 w^2 / D$$

Number of channel widths required for complete mixing:

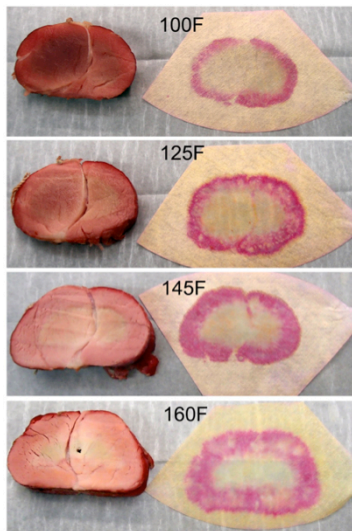
$$\frac{Z}{w} \sim \frac{U_0 w}{D} \equiv \mathcal{P}$$



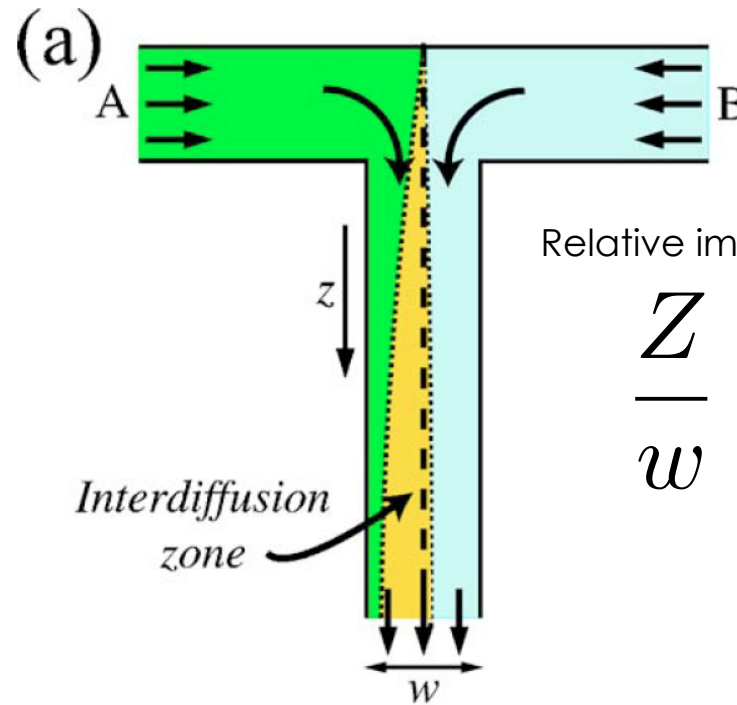
## Convection



## Diffusion



## Péclet Number: convection/diffusion



Relative importance: **convection** to **diffusion**:

$$\frac{Z}{w} \sim \frac{U_0 w}{D} \equiv \mathcal{P}$$

Consider a **small protein** flowing with liquid:

- Typical size: 5 nm
- Diffusion constant:  $40 \mu\text{m}^2/\text{s}$
- e.g. Channel width:  $100 \mu\text{m}$
- e.g. Flow velocity:  $100 \mu\text{m}/\text{s}$

$$\mathcal{P} \sim 250 \text{ channel widths}$$

$$L_0 \sim 2.5 \text{ cm}$$

$$\tau_D \sim 4 \text{ min}$$

## Parallel Laminar Flows

Pressure-driven laminar flow of two adjacent miscible streams.

- Solution on left contains **calcium**.
- Solution on right contains calcium-dependent **fluorophore**, Fluo-3.
- In water, Fluo-3 and calcium form a fluorescent complex at a **diffusion limited rate**.

Convective-diffusion equation:

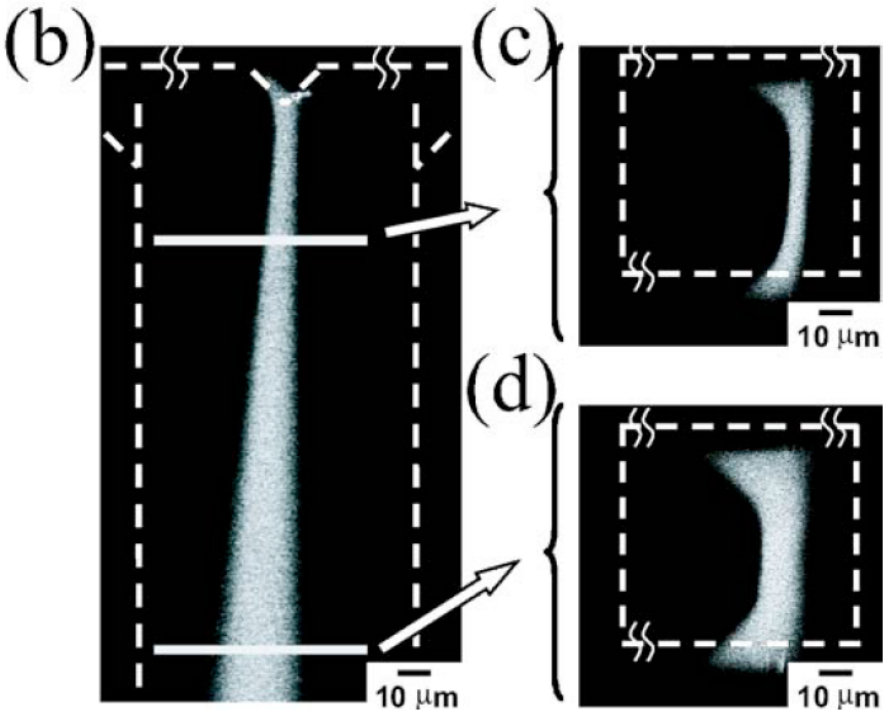
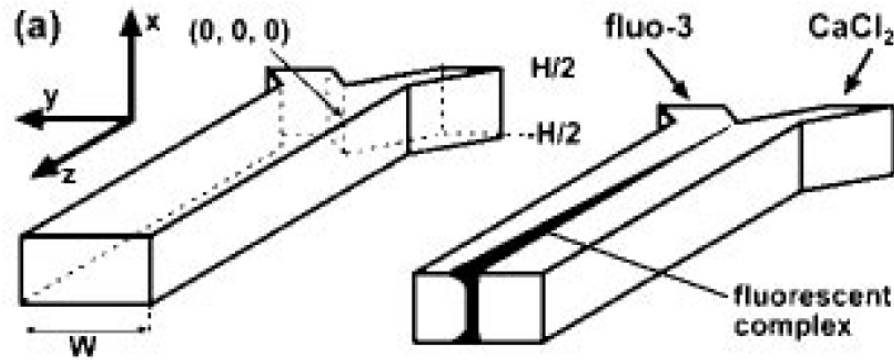
$$\mathcal{P} \mathbf{u} \cdot \nabla c = \nabla^2 c \cong \left( \frac{\partial}{\partial x} + \frac{\partial}{\partial y} \right) c$$

For high Péclet number flows,  $\mathcal{P} \gg z/H \gg 1$

- Diffusive broadening down the channel depends on both  $y$  &  $x$

Away from boundaries, the interfacial region:

$$\underbrace{U_m}_{\text{Maximum velocity}} \frac{\partial c}{\partial z} = D \nabla^2 c \longrightarrow \delta_y(z) \sim \left( \frac{Dz}{\underbrace{U_a}_{\text{Average velocity}}} \right)^{1/2}$$





## Parallel Laminar Flows

Pressure-driven laminar flow of two adjacent miscible streams.

- Solution on left contains **calcium**.
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- In water, Fluo-3 and calcium form a fluorescent complex at a **diffusion limited rate**.

Away from boundaries, the interfacial region:

$$\delta_y \sim t^{1/2} \sim z^{1/2}$$

**Lévy Problem:** Near from boundaries, velocity varies **linearly** with distance into the channel.

Diffusion across a linear flow field

$$\delta_x \sim (D_m t)^{1/2}$$

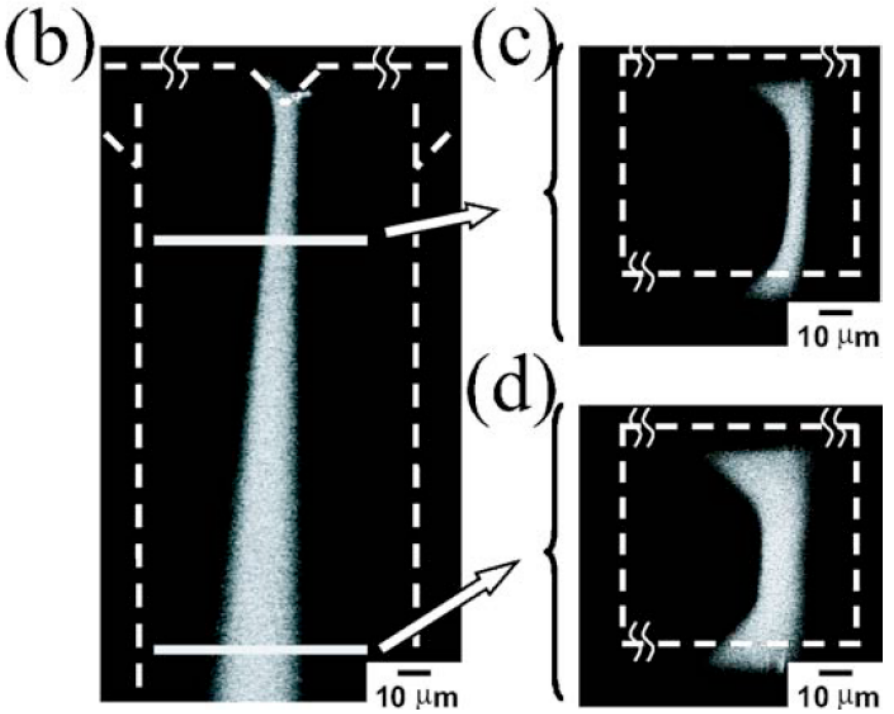
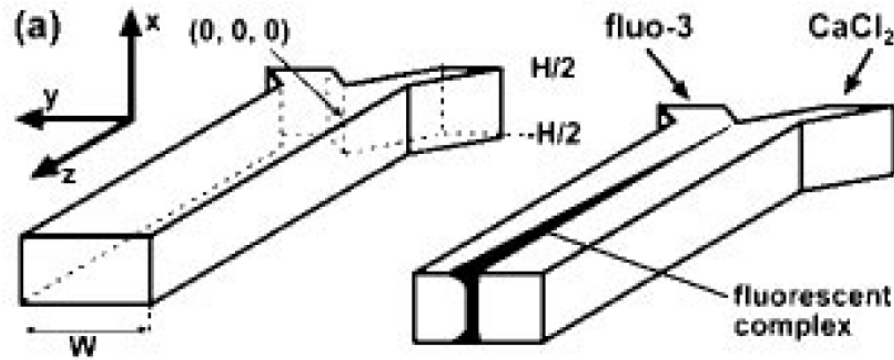
Thickness of diffusion boundary layer

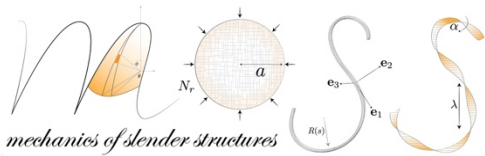
$$t(z) \sim z / (G \delta_x)$$

$$\text{Shear rate: } G = \frac{\partial u_z}{\partial x}$$

Diffusion is the only time scale in both x & y:

$$\delta_y \sim \delta_x \sim \left( \frac{z D_m}{G} \right)^{1/3}$$





# Sensing & Filtering

## Parallel Laminar Flows

Pressure-driven laminar flow of two adjacent miscible streams.

- Solution on left contains **calcium**.
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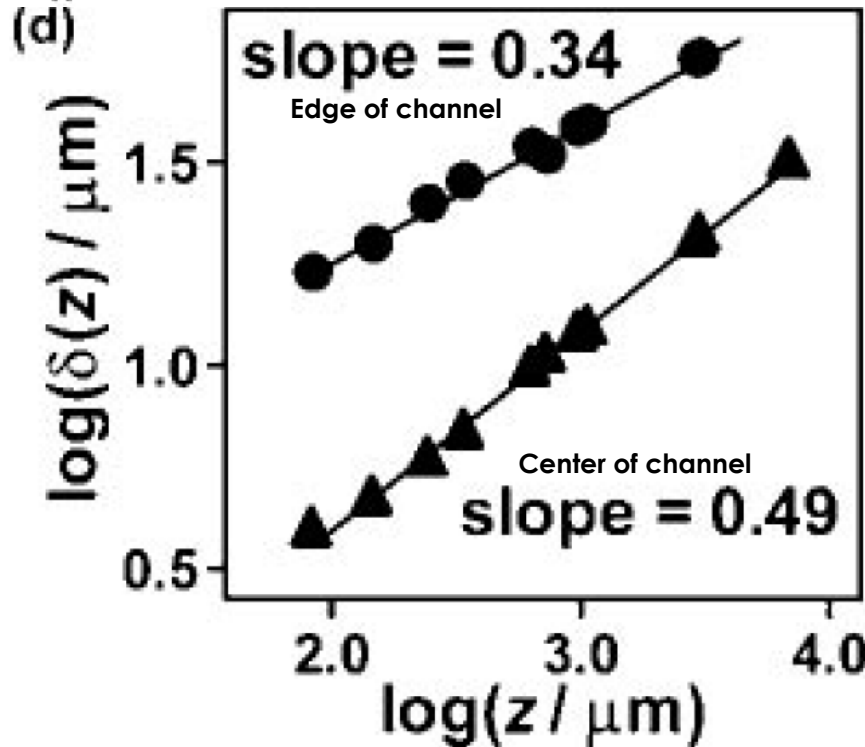
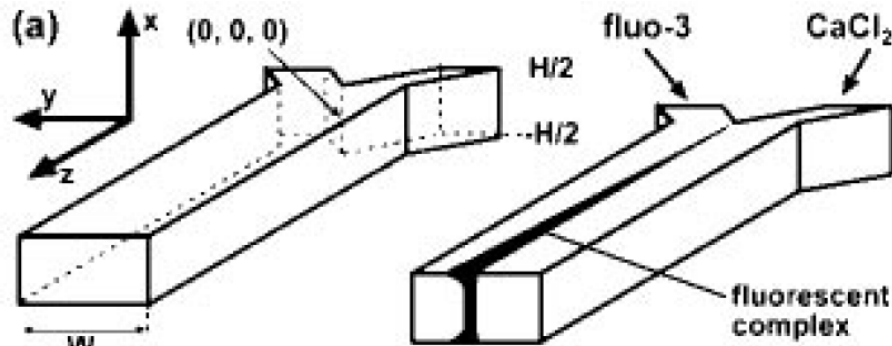
Thickness of diffusion boundary layer

$$t(z) \sim z / (G \delta_x)$$

$$\text{Shear rate: } G = \frac{\partial u_z}{\partial x}$$

Diffusion is the only time scale in both x & y:

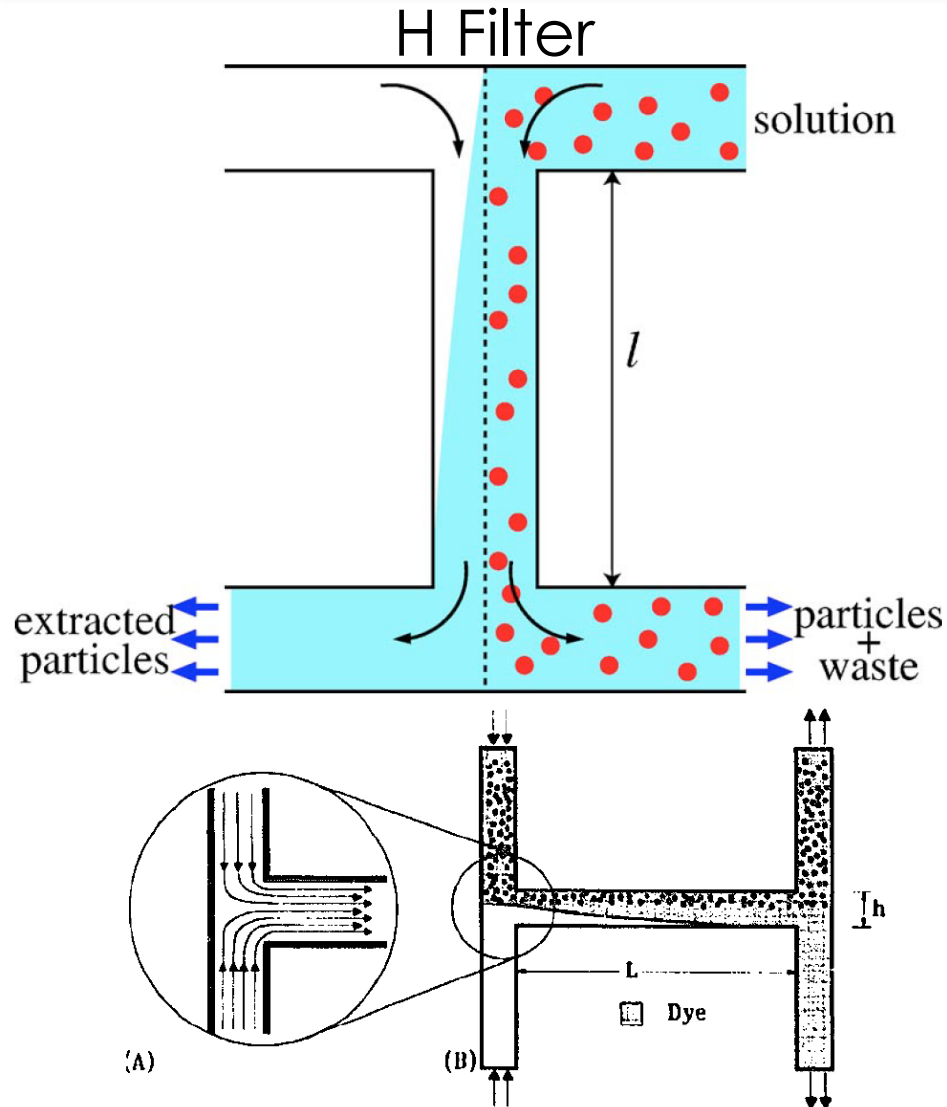
$$\delta_y \sim \delta_x \sim \left( \frac{z D_m}{G} \right)^{1/3}$$





# Sensing & Filtering

## Parallel Laminar Flows



Filter particles by size without a membrane.

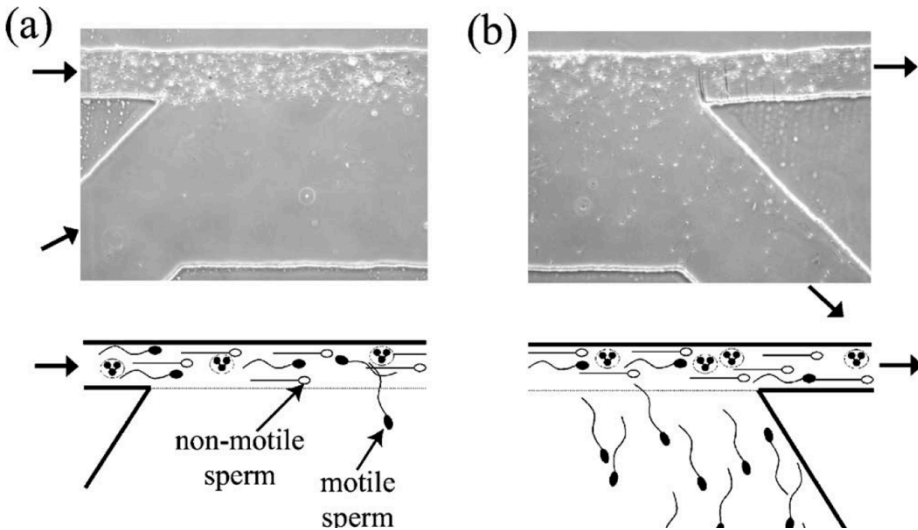
- Pressure-driven laminar flow of two adjacent miscible streams.
- One stream is a dilute solution of different sized particles.
- Each particle has its own **diffusivity** and **Péclet number**.

**Péclet number** determines the channel length required for each component to **diffuse across the channel width**.

**H filter** works best when one Péclet number is large and the other is small.

- e.g. small particles diffuse across channel, large ones do not.

## H Filter - Variant

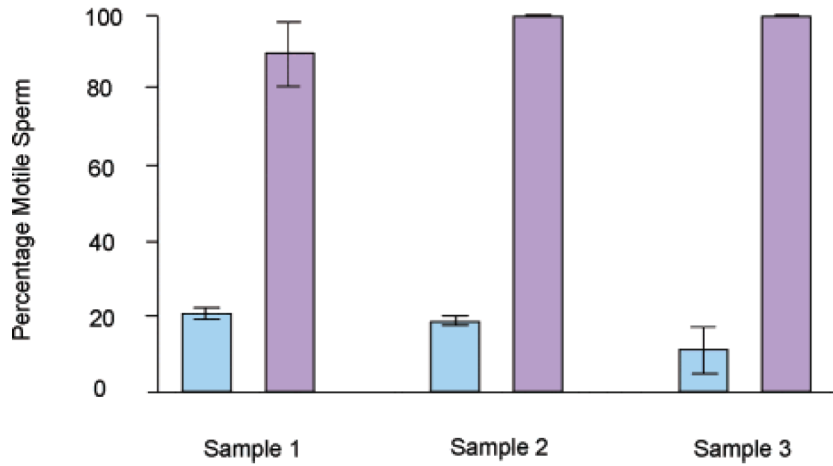


### Separation by Péclet number

- Components to be separated spread across channel at different rates.
- e.g. Separation of motile vs. nonmotile sperm.

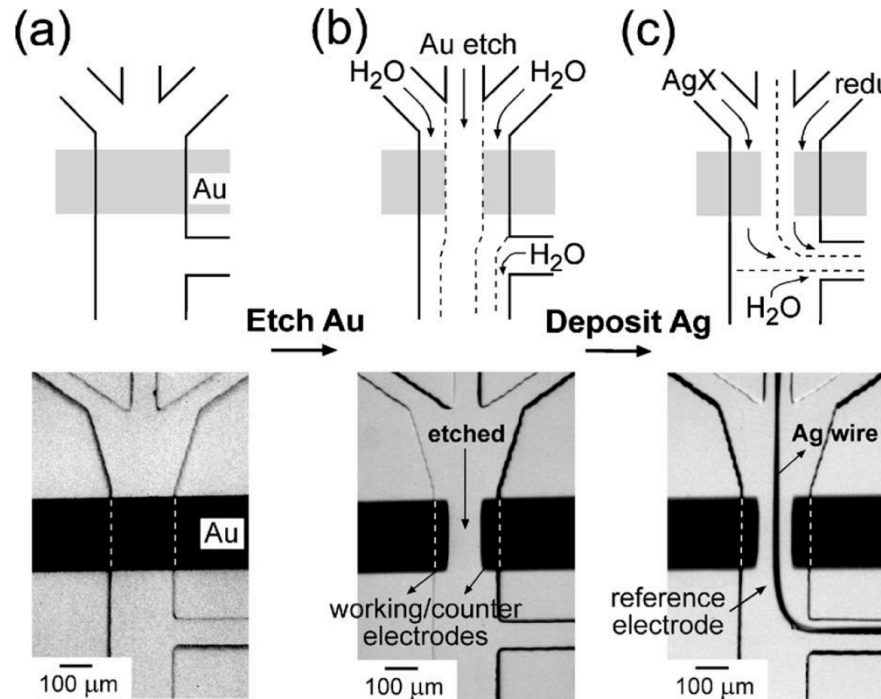
**Motile** sperm **swim rapidly** and **randomly** to fill the channel, as compared to nonmotile sperm.

- Nonmotile sperm spread by diffusion alone.



Motile sperm purity at inlet (blue) vs. motile sperm purity at outlet (purple)

## Fabrication using multiple laminar streams ...beating diffusion...



### Fabrication of a three electrode system

- Multiple streams & large Péclet number.
- Minimal mixing over large distances.

Nonmixing, **high Péclet flows** can fabricate/etch structures within microchannels.

e.g. Parallel streams **selectively etch** a gold electrode

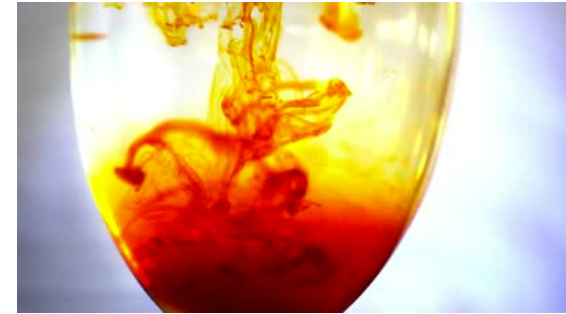
- A silver wire is formed by a precipitation reaction between the two streams.
- Result: three-electrode system.



Rapid mixing means doing better than diffusion.

Why mix?

- Study chemical reaction kinetics...
- Probe protein folding...



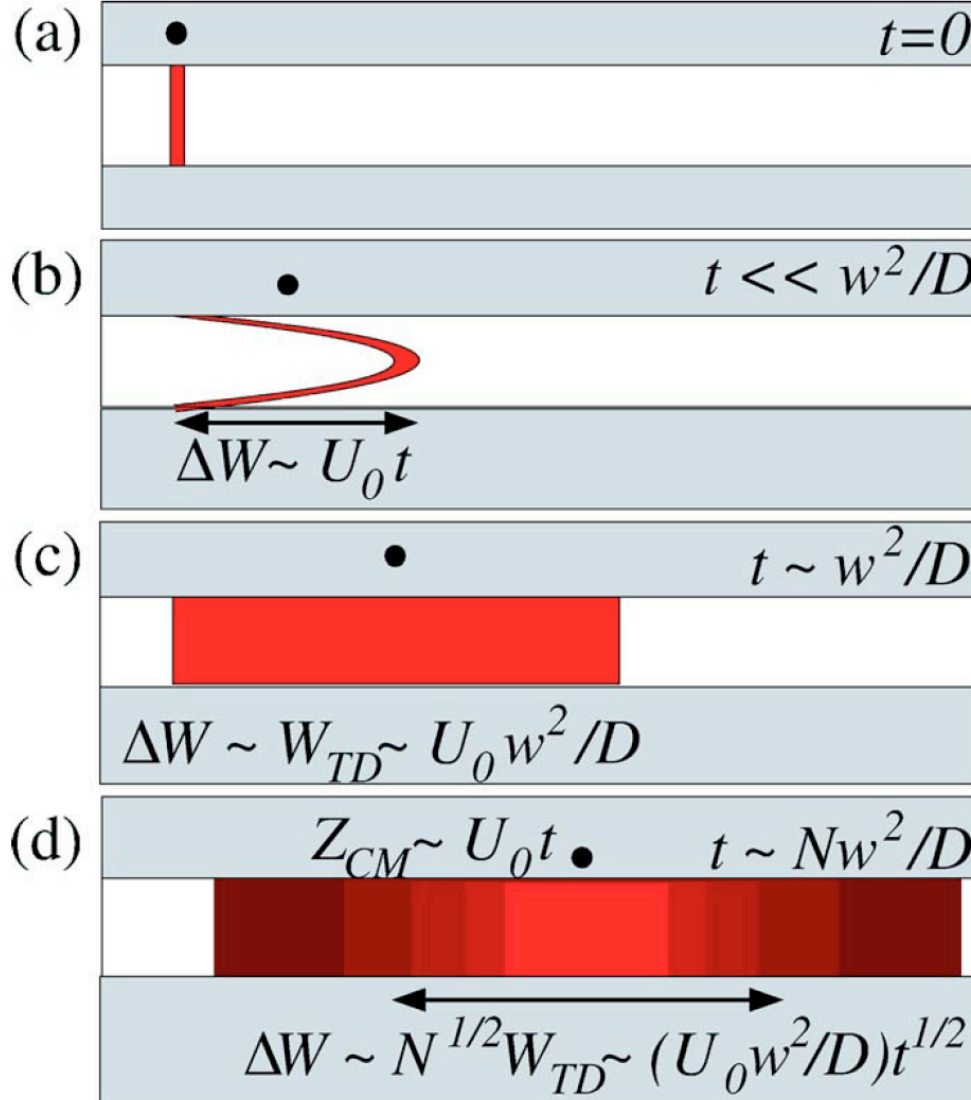
Fluid **stirring** will **stretch** and **fold** inhomogenous fluid elements until **mixing** occurs.

- Mixing: diffusive migration across streamlines.
- Stirring motions reduce distance over which mixing must occur.

General design principles for mixing:

**Dispersion of tracers** occurs first by **convective stretching** with the fluid, followed by **diffusive homogenization**.

Tracers: an object transported by and diffuses into the fluid,  
e.g. dyes, analyte molecules, proteins, cells, salt, or heat.



## Taylor Dispersion

Thin stripe of tracers spans circular channel

- Channel radius:  $w$

Pressure-driven (Poiseuille) flow stretches strip into a parabola with profile:

$$u_z = U_0 \left( 1 - \frac{r^2}{w^2} \right)$$

Molecular diffusion across the channel smears the parabola into a plug of width:

$$\Delta W \sim W_{TD} \sim U_0 \frac{w^2}{D}$$

Each stripe is **convectively stretched** then **diffusively smeared**

After  $N$  time steps of  $t \sim Nw^2/D$ , the initial stripe grows into a Gaussian with width:

$$\langle W^2 \rangle^{1/2} \sim N^{1/2} W_{TD} \sim \left( \frac{U_0^2 w^2}{D} t \right)^{1/2}$$

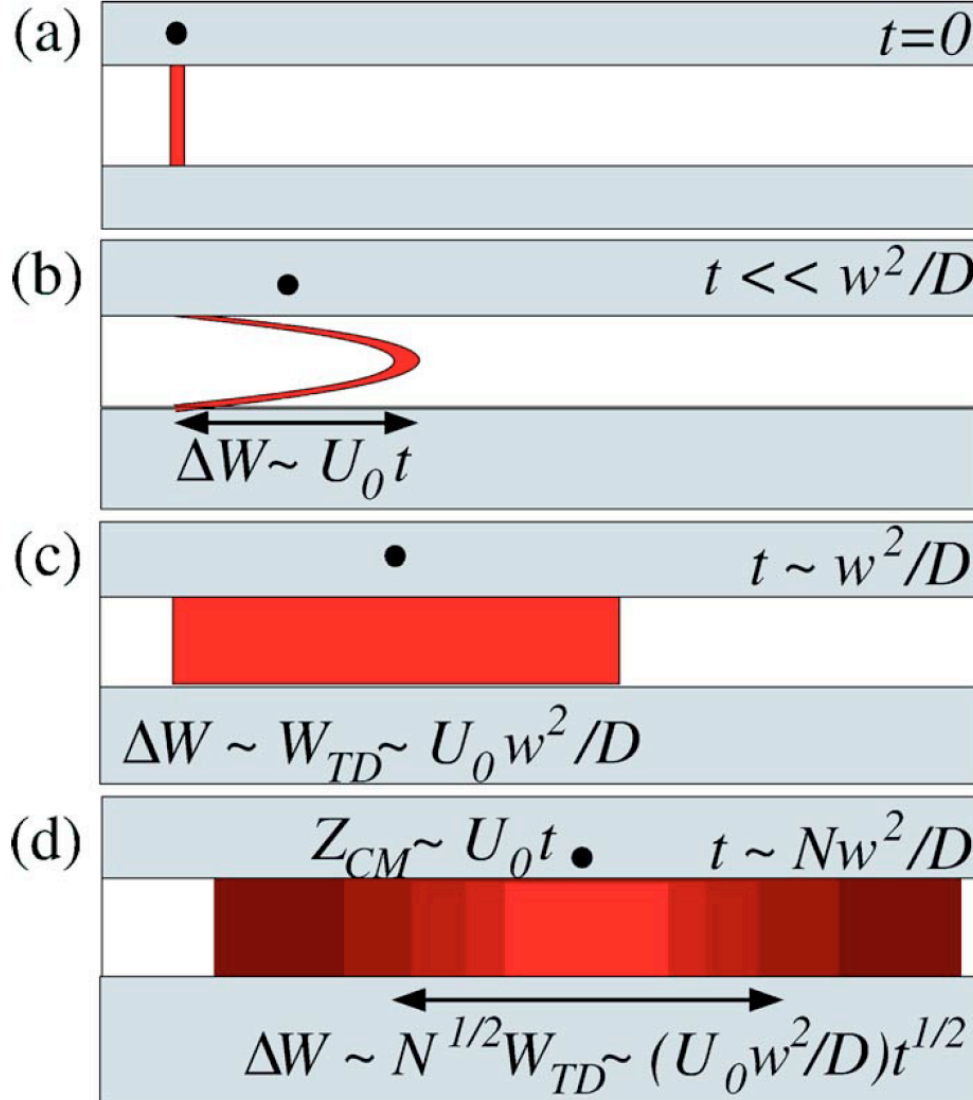
Taylor, Geoffrey. "Dispersion of soluble matter in solvent flowing slowly through a tube." In Proceedings of the Royal Society of London A: Mathematical, Physical and Engineering Sciences, vol. 219, no. 1137, pp. 186-203. The Royal Society, 1953.

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Squires, Todd M., and Stephen R. Quake. "Microfluidics: Fluid physics at the nanoliter scale." Reviews of modern physics 77.3 (2005): 977.



# Mixing



## Taylor Dispersion

Initial stripe grows to a Gaussian with width:

$$\langle W^2 \rangle^{1/2} \sim N^{1/2} W_{TD} \sim \left( \frac{U_0^2 w^2}{D} t \right)^{1/2}$$

The **tracer distribution grows** diffusively with an effective long-time **axial diffusivity**:

$$D_z \sim \frac{U_0^2 w^2}{D} \sim \mathcal{P}^2 D$$

Axial diffusivity occurs in addition to molecular diffusivity.

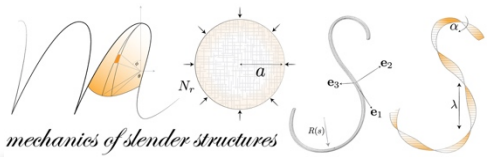
Relevant examples for specific geometries:

$$D_z = \frac{U_0^2 w^2}{48D} \quad D_z = \frac{U_0^2 h^2}{210D} \quad D_z = \frac{2U_0^2 h^2}{15D}$$

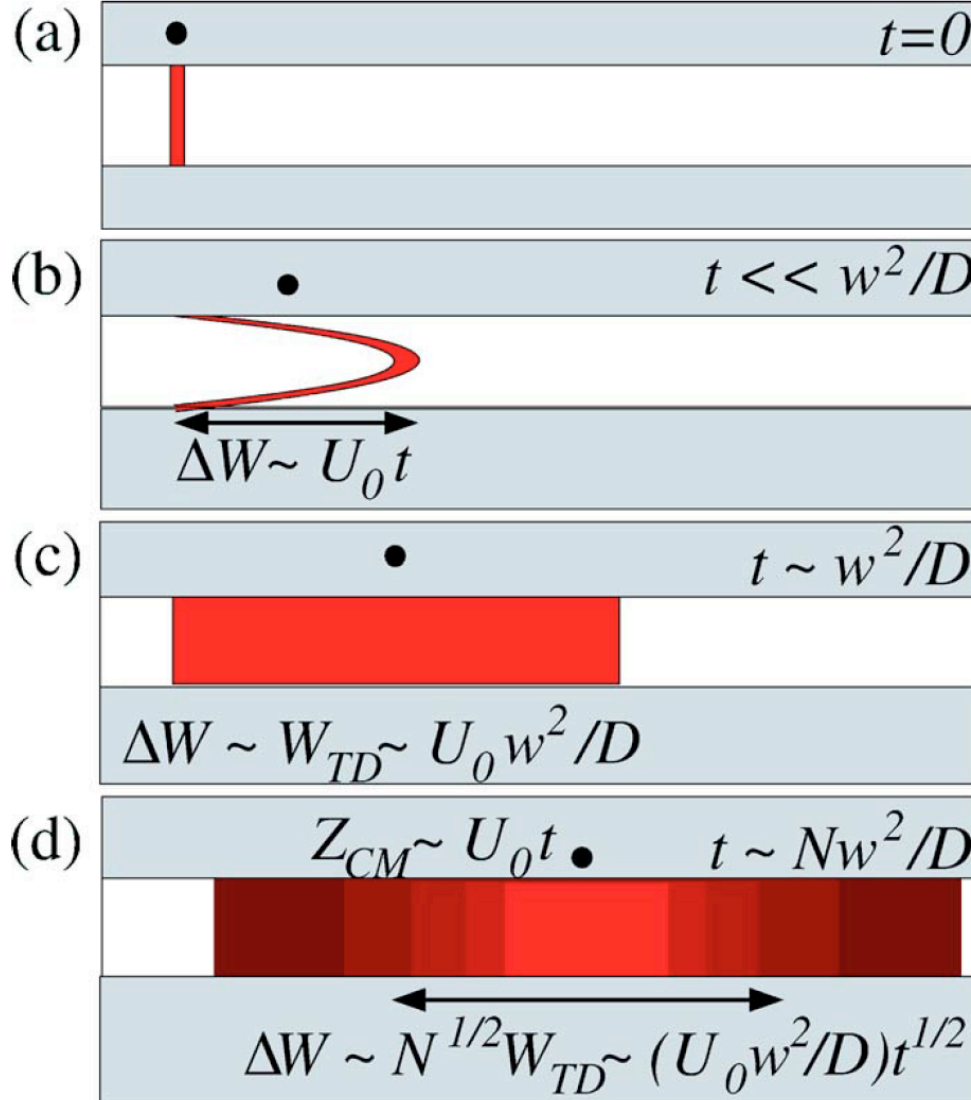
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# Mixing



## Taylor Dispersion

$$D_z \sim \frac{U_0^2 w^2}{D} \sim \mathcal{P}^2 D$$

Convective stretching enhances axial dispersivity

Taylor dispersivity is only valid at

- Long time scales:  $t \gg w^2/D$
- Downstream lengths:  $L \gg \mathcal{P}w$

Taylor dispersion acts in direction of flow.

- Not observed in T sensors or H filters

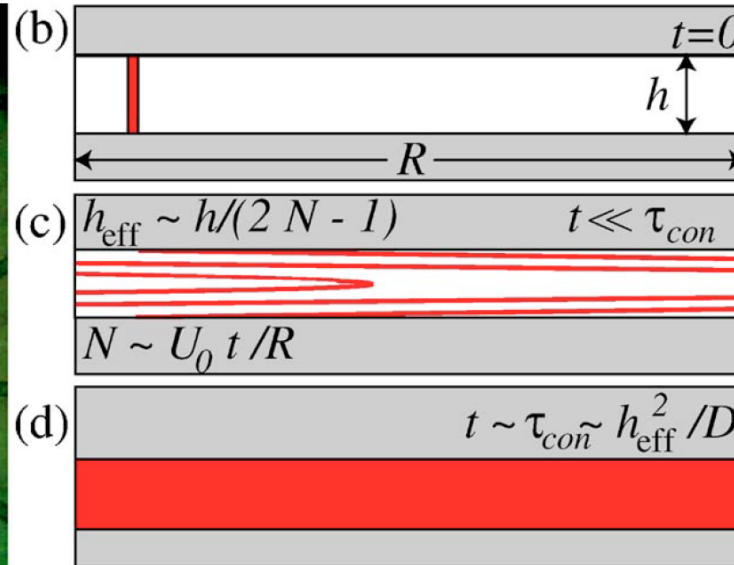
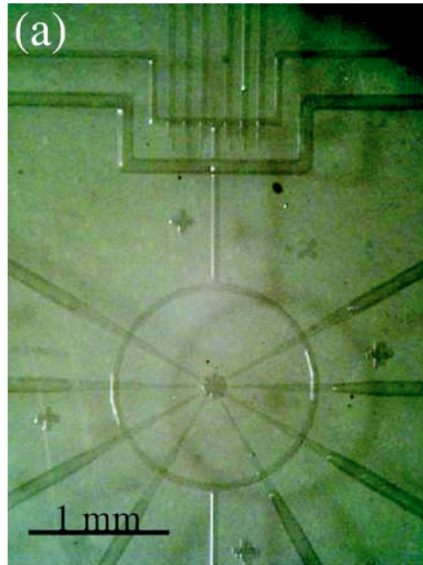
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## Rotary Mixer



Fluid pumped around a circular channel:

- radius  $R$
- height  $h$
- average velocity  $U_0$ .

(i.) Diffusion-dominated

$$\mathcal{P} = U_0 h / D \ll 1$$

- Mixing occurs when tracers diffuse around the circumference of the ring, at time:

$$\tau_R \sim \frac{(2\pi R^2)}{D} = \left(\frac{2\pi R}{h}\right)^2 \tau_D$$

- Independent of Péclet number.

(ii.) Taylor dispersion-mediated

$$1 \ll \mathcal{P} \ll 2\pi R / h$$

- Axial spreading increases diffusivity with Taylor dispersivity:

$$\tau_{TD} \sim \frac{(2\pi R^2)}{D_z} = \frac{D(2\pi R)^2}{U_0^2 h^2} \sim \frac{\tau_R}{\mathcal{P}^2}$$

- Dominant when molecules diffuse over  $h$  before convection.

(iii.) Convectively Stirred

$$\mathcal{P} \gg 2\pi R / h$$

- At high flow rates, tracer stripes **fold** into themselves before molecules diffuse across channel.

$$\tau_{\text{con}} \sim \mathcal{P}^{-2/3} \left(\frac{\pi R}{h}\right)^{2/3} \tau_D$$

$$\sim \mathcal{P}^{-2/3} \left(\frac{h}{\pi R}\right)^{4/3} \tau_R$$

## Chaotic advection

Even simple **Stokes flows** can have **chaotic streamlines** that exponentially **stretch and fold**.

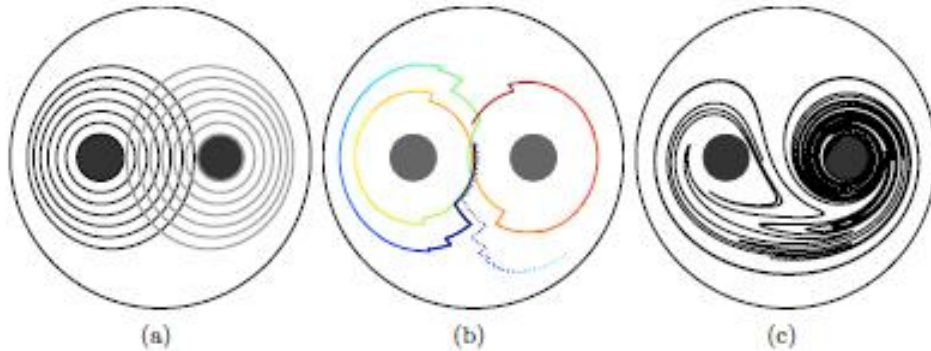
Steady, incompressible **two-dimensional flows** are integrable and **cannot exhibit chaotic trajectories**.

Steady **three-dimensional flows** can have chaotic streamlines (as can unsteady two dimensional flows).

Can occur in droplets by superposing two simple flow fields

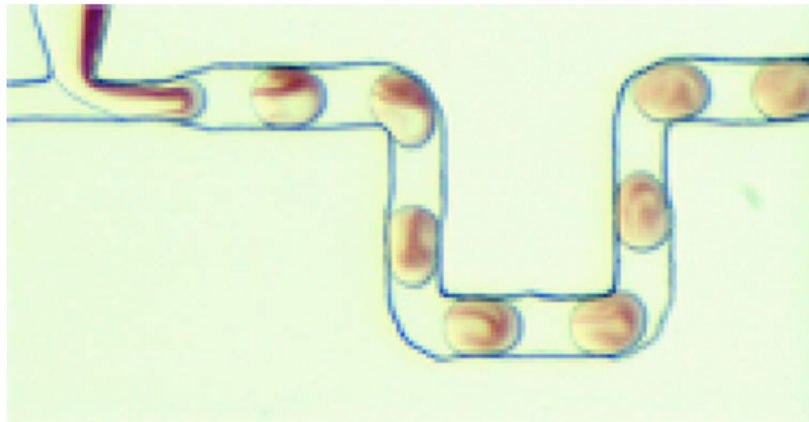
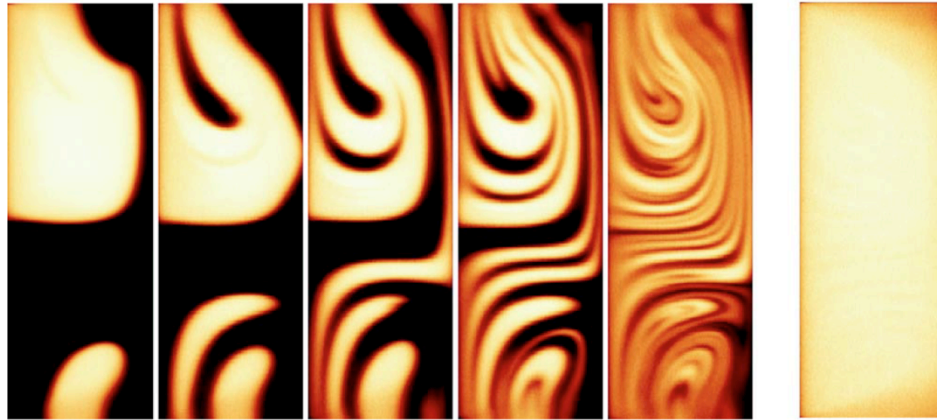
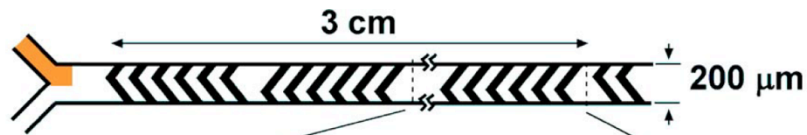
- (e.g. sedimenting drop in a shear flow)

Blinking vortex flow





## Continuous-flow staggered herringbone mixer



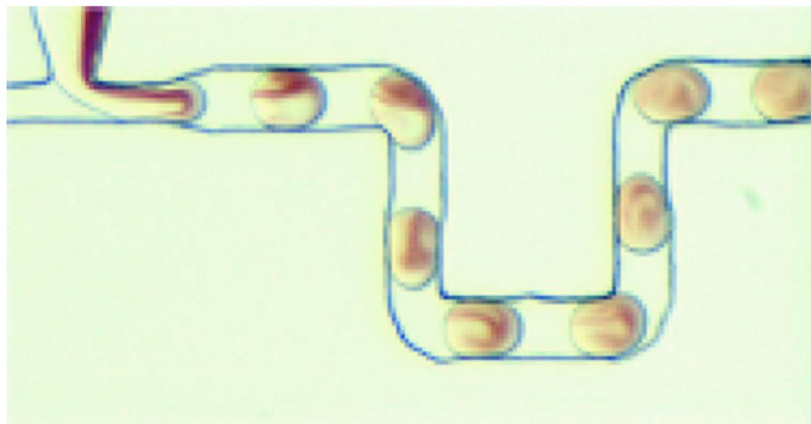
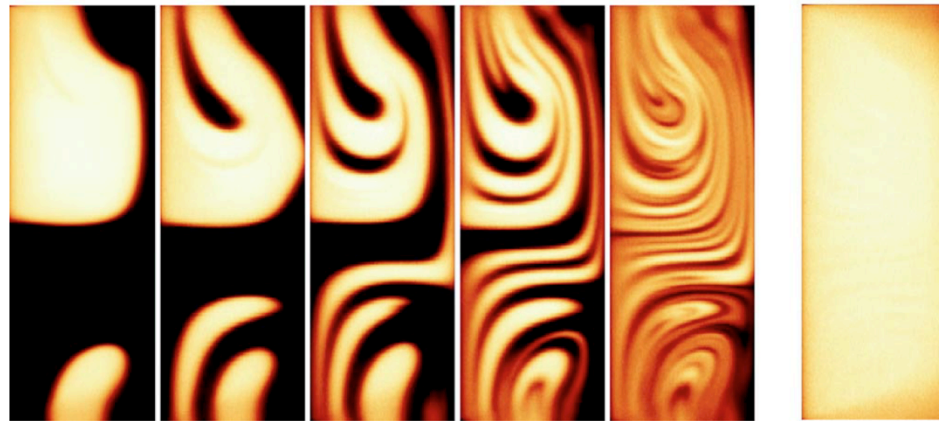
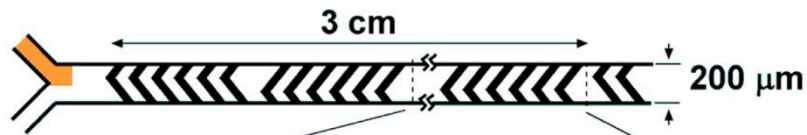
## Chaotic advection

Even simple **Stokes flows** can have **chaotic streamlines** that exponentially **stretch and fold**.

The **staggered herringbone mixer** is a chaotic mixer for continuous flow systems – independent of inertia.

- **Asymmetric grooves** in channel walls induce axially modulated **secondary flow**.
- Counter-rotating fluid rolls.
- Asymmetry is periodically reversed so that the distance between stripes halves with each cycle – **exponential stretching/folding**.

Continuous-flow staggered herringbone mixer



## Chaotic advection

Asymmetry pattern on channel causes exponential stretching/folding.

After  $N$  cycles of time:  $\tau_{cyc} \sim N L_{cyc} / U$

Stripes separated by distance:  $h_{eff} \sim h / 2^N$

Time to diffuse between stripes:  $\tau_D \sim h_{eff}^2 / D$

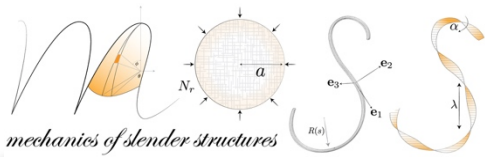
**Mixing:**  $\tau_{cyc} \sim \tau_D$

Therefore, the number of cycles:

$$N_{chaotic} \sim \ln \mathcal{P}$$

Time for chaotic mixing:

$$\tau_{chaotic} \sim \frac{L_{cyc}}{h} \frac{\ln \mathcal{P}}{\mathcal{P}} \tau_D$$



## Confined Fluid Flow: Microfluidics and Capillarity

**Reynolds** Number: Inertia vs. Viscous effects

- Review of characteristic flows...

**Péclet** Number: Transport phenomena in a continuum

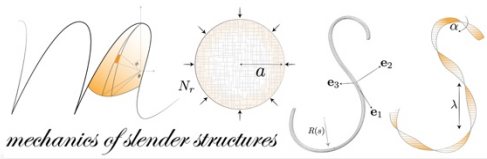
- Diffusion, separation, and mixing...

**Geometric** confinement: Controlling and manipulating fluid flow

- Microfluidic fabrication, valving, pumping...

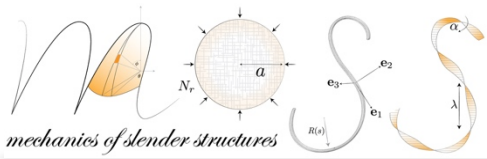
**Capillary** Number: Viscosity vs. Surface tension

- Droplet formation, capillary rise, elasticity...



# Geometric Confinement: Control & Manipulation of Fluid Flow



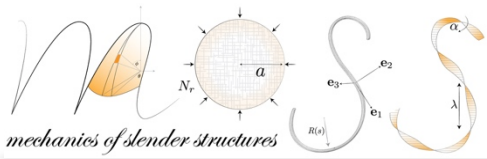


# A Giraffe's Jugular

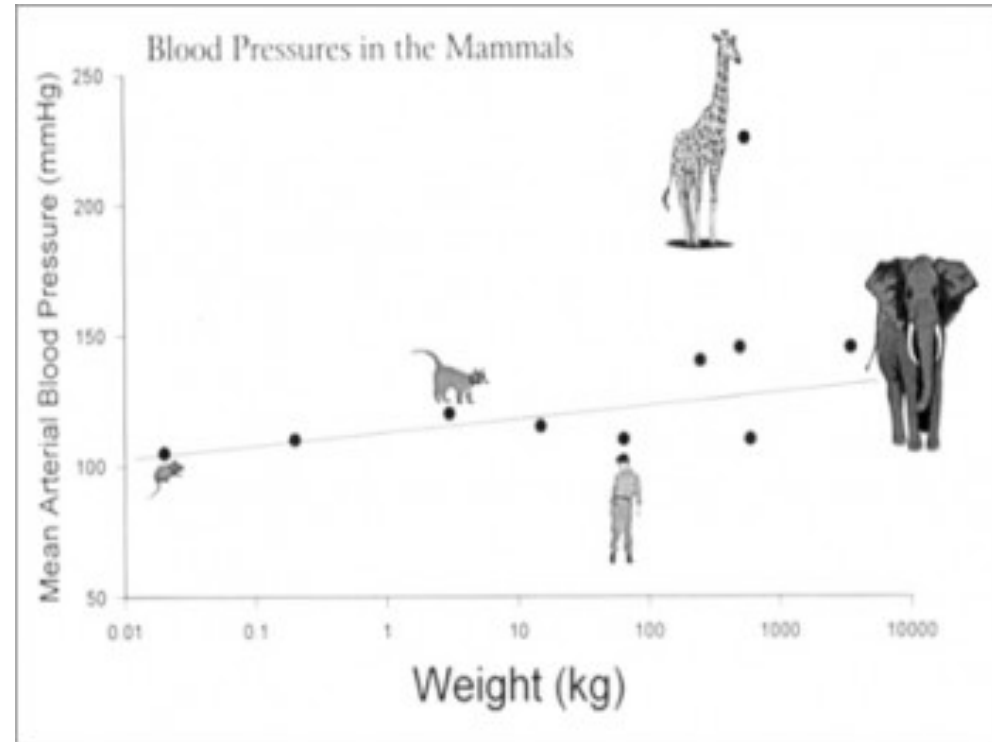
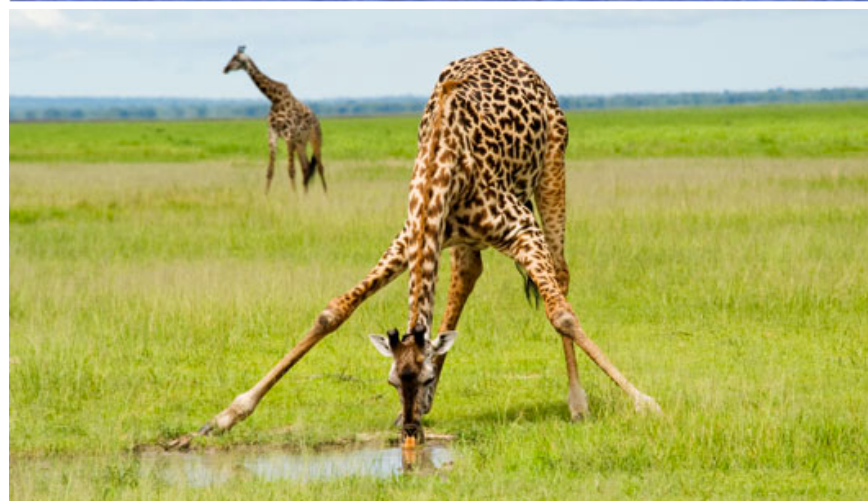


[http://upload.wikimedia.org/wikipedia/commons/e/e6/Giraffes\\_at\\_west\\_midlands\\_safari\\_park.jpg](http://upload.wikimedia.org/wikipedia/commons/e/e6/Giraffes_at_west_midlands_safari_park.jpg)





# A Giraffe's Jugular

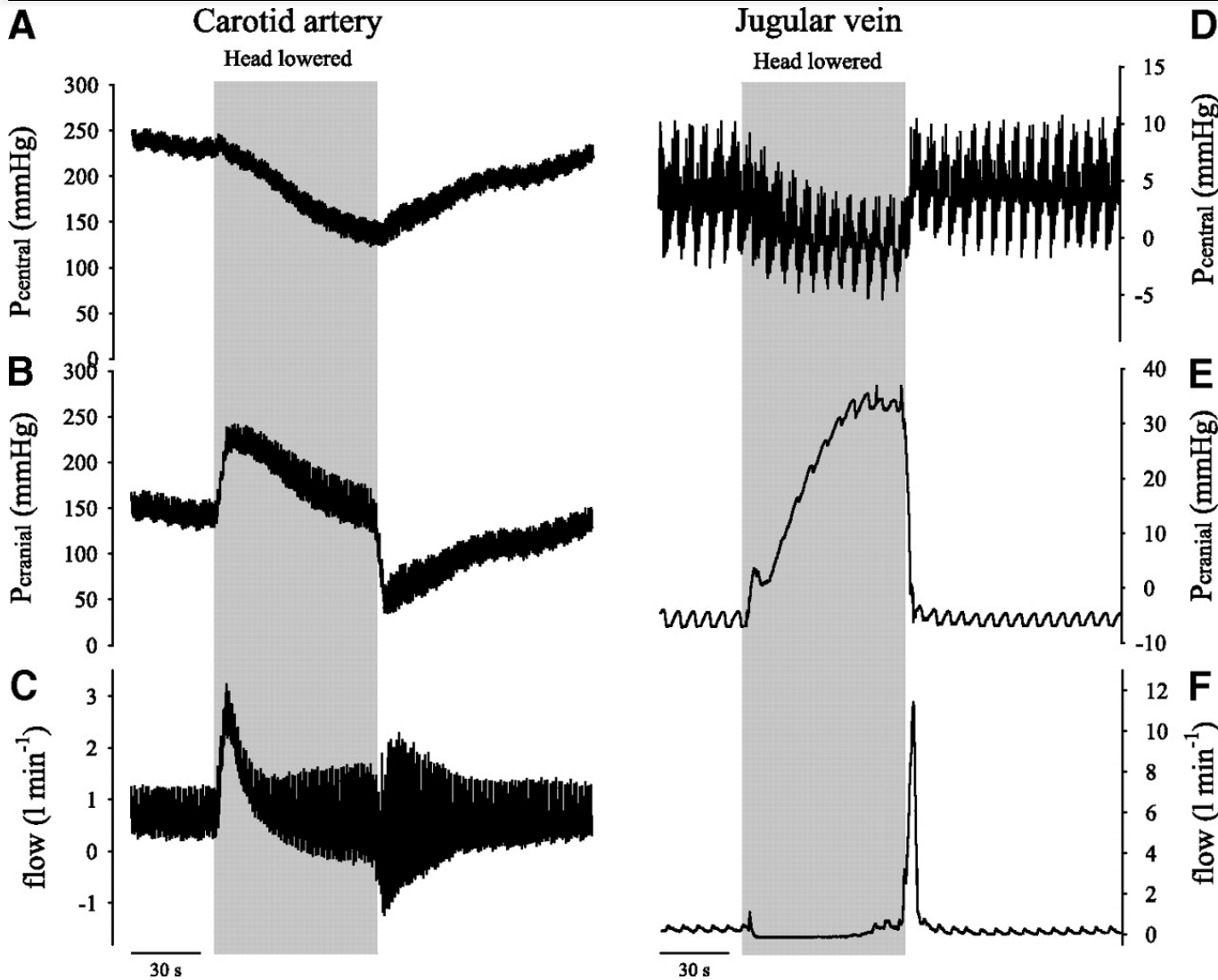


Top giraffe: <http://cdn.asiancorrespondent.com/wp-content/uploads/2011/11/Giraf2-349x248.jpg>  
 Bottom giraffe: [http://www.onekind.org/uploads/a-z/az\\_giraffe1.jpg](http://www.onekind.org/uploads/a-z/az_giraffe1.jpg)





# A Giraffe's Jugular



Arterial and venous pressure decrease upon lowering the head.

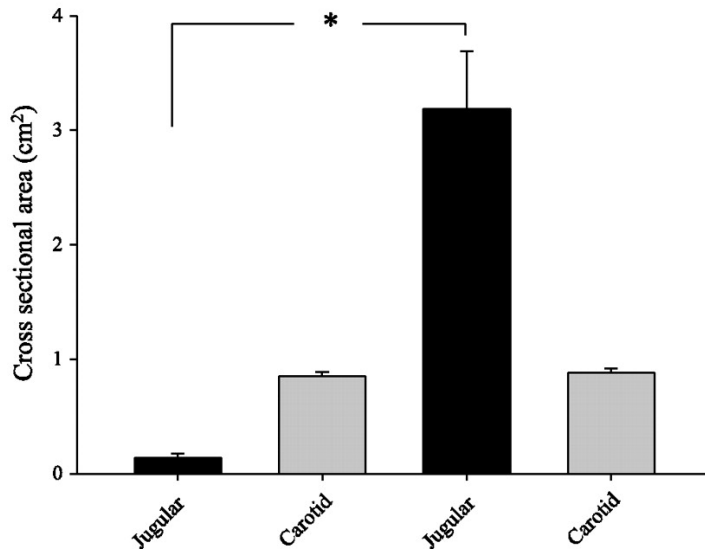
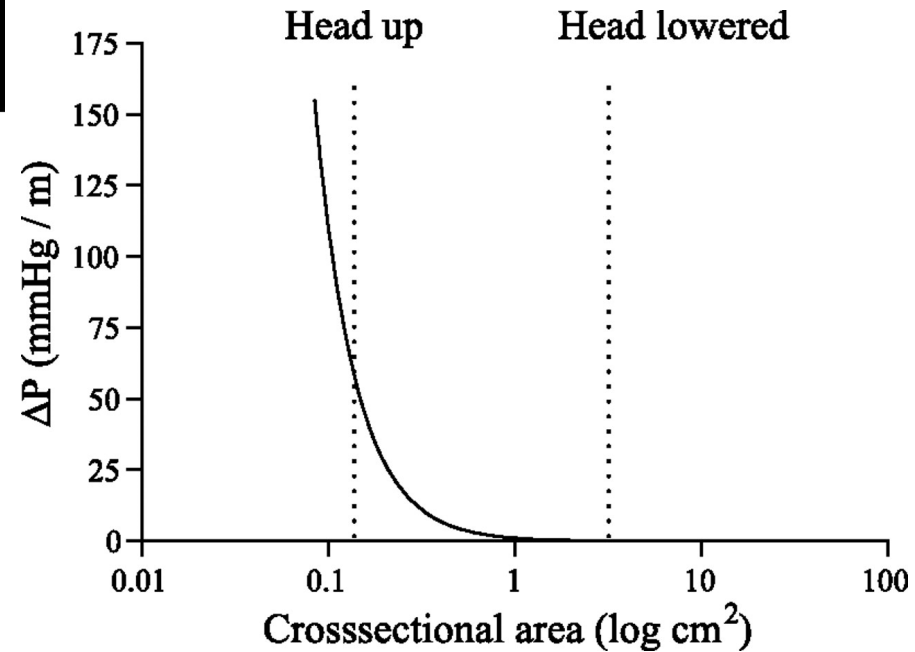
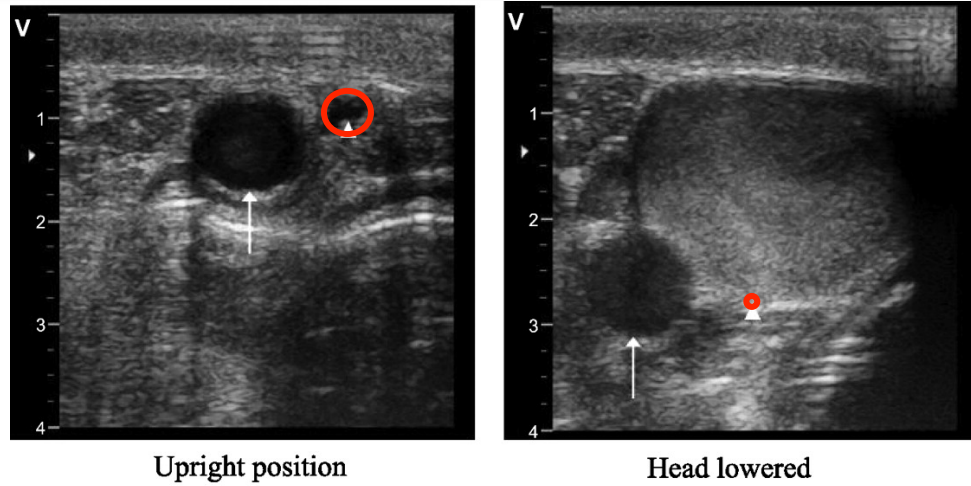
Rise in cranial jugular pressure coincides with cessation of jugular flow.

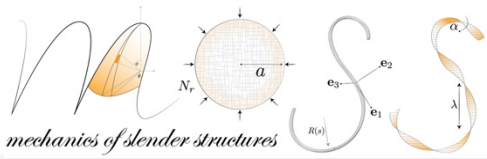
Jugular venous flow ceased for ~30s when lowered.



# A Giraffe's Jugular

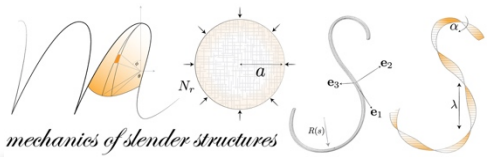
Jugular vein's cross sectional area changes upon lifting/lowering of the Giraffe's head.





# A Giraffe's Jugular

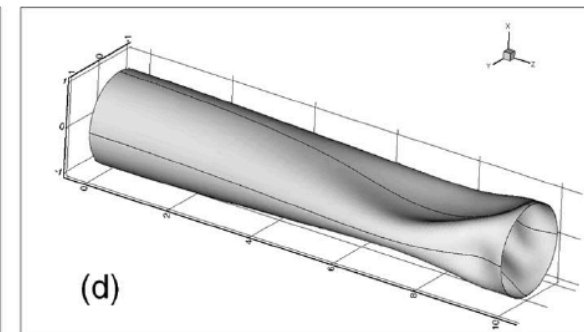
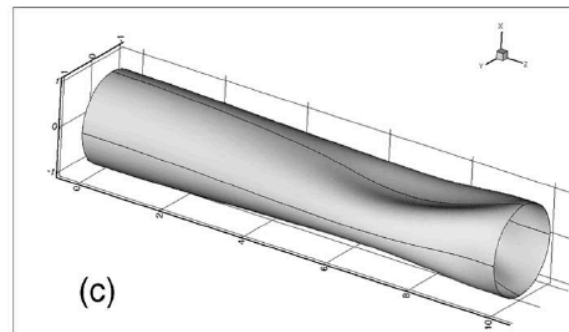
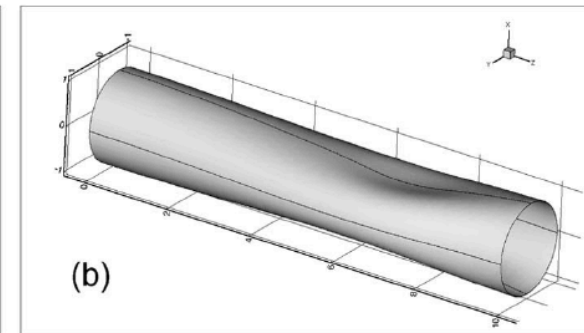
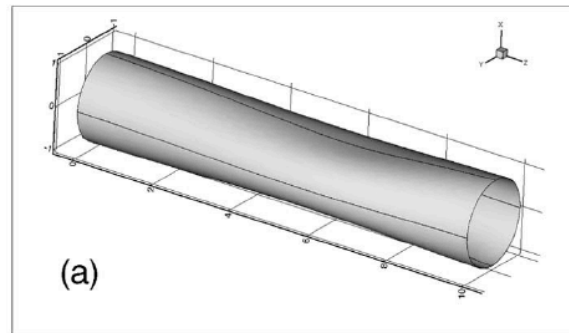
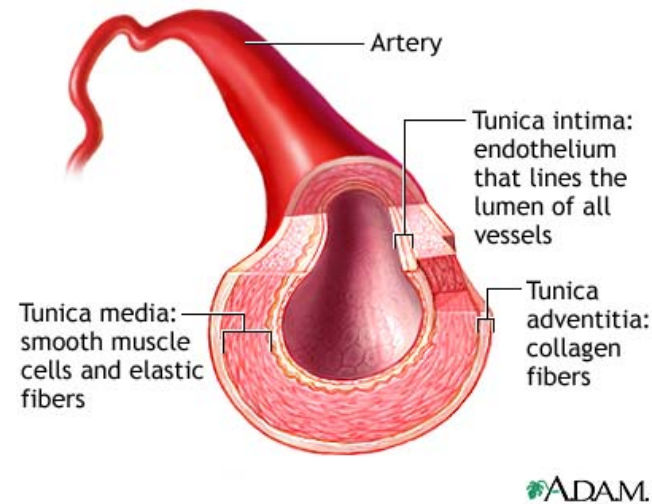




# Flexible Tubes

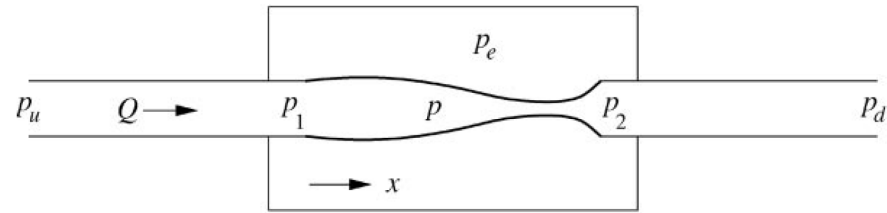
Almost all **vessels** carrying **fluids** with the body are **flexible**.

Fluid-structure interactions between **internal flow** and tube **deformation** often **dictate** a vessel's **biological function** or dysfunction.

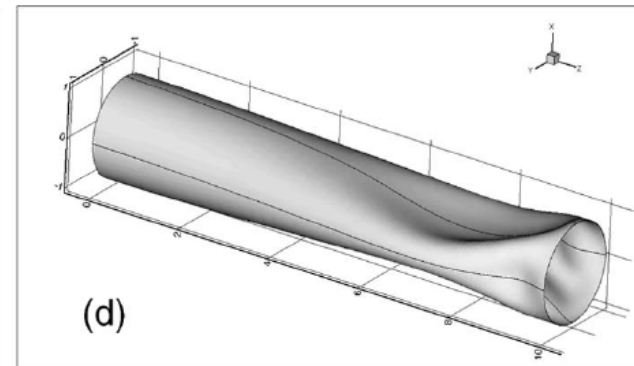
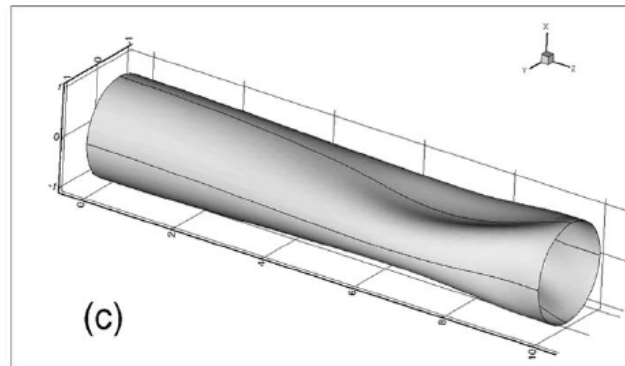
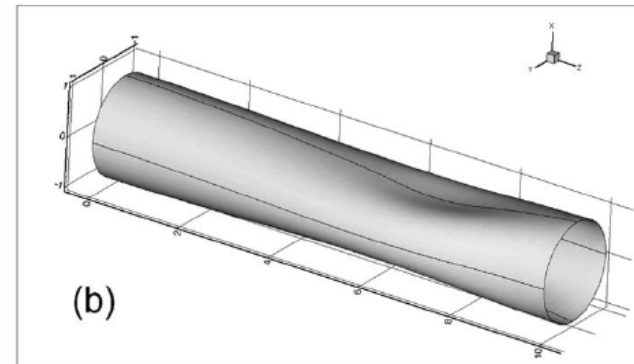
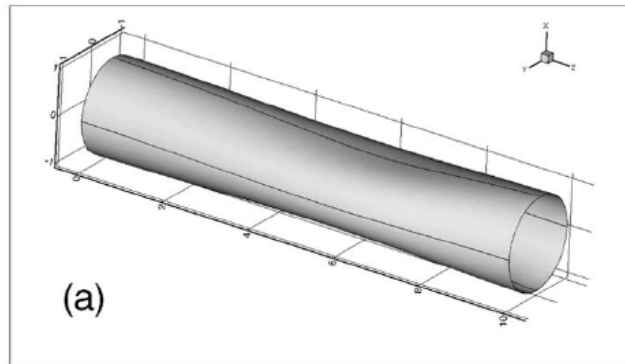
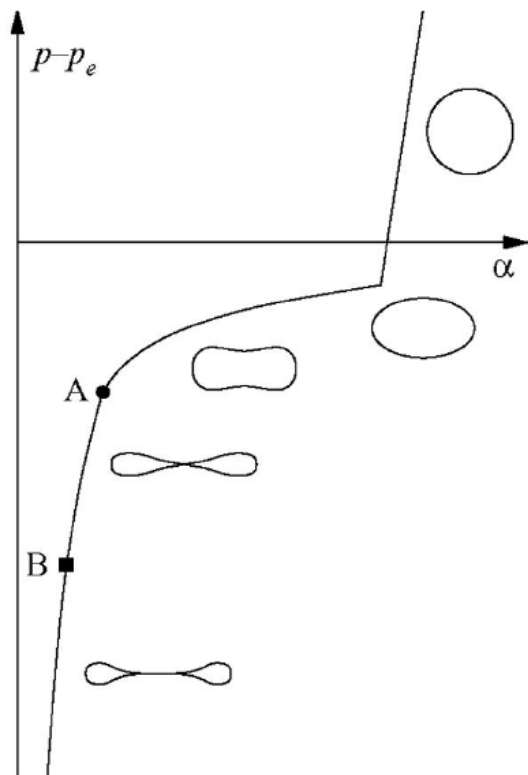




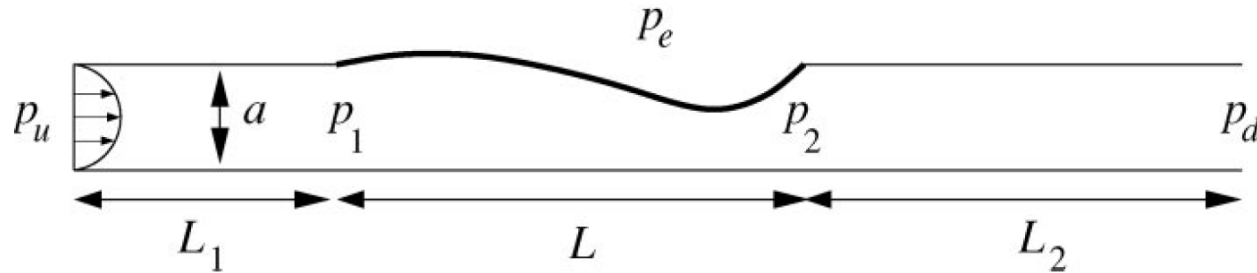
## Starling Resistor



**Figure 1** A Starling Resistor: a collapsible tube is mounted between two rigid tubes and is enclosed in a chamber held at pressure  $p_e$ . Flow with volume flux  $Q$  is driven by the imposed pressure drop  $p_u - p_d$ .



## Starling Resistor (1D Model)



Mass conservation:

$$\frac{\partial \alpha}{\partial t} + \frac{\partial (u\alpha)}{\partial x} = 0$$

$\alpha =$  tube cross sectional area

Momentum conservation:

$$\rho \left( \frac{\partial u}{\partial t} + u \frac{\partial u}{\partial x} \right) \approx \underbrace{-\frac{\partial p}{\partial x}}_{\text{pressure}} - \underbrace{R(u, \alpha)}_{\text{viscous resistance per unit length}}$$

Pressure-area relation:

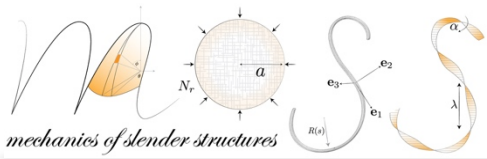
$$p - p_e = \underbrace{\mathcal{P}(\alpha)}_{\text{compliance}} - \underbrace{T \frac{\partial^2 \alpha}{\partial x^2}}_{\text{longitudinal tension}} + \underbrace{D \frac{\partial \alpha}{\partial t}}_{\text{viscous damping}} + \underbrace{M \frac{\partial^2 \alpha}{\partial t^2}}_{\text{inertia}}$$

(Very) Reduced model

- “Bench-top” model for a deformable airway
- Consider airways as single, compliant tube
- Prone to instabilities – best with low Reynolds

Compliance of tube gives rise to  
**fluid-structure interactions**

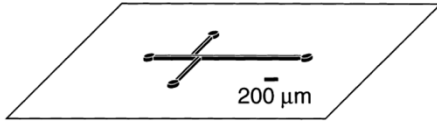




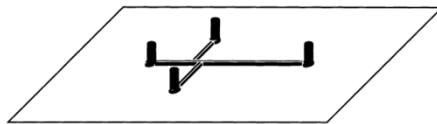
How do we translate these ideas  
to microfluidic devices?

# Fabrication

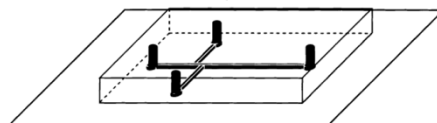
**A** Fabricate master by rapid prototyping



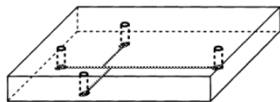
**B** Place posts to define reservoirs



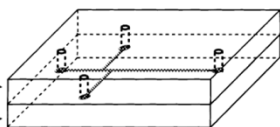
**C** Cast prepolymer and cure



**D** Remove PDMS replica from master

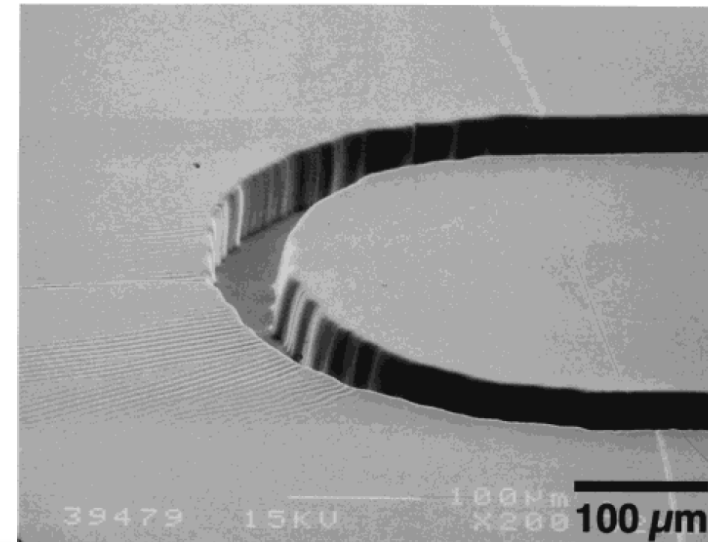
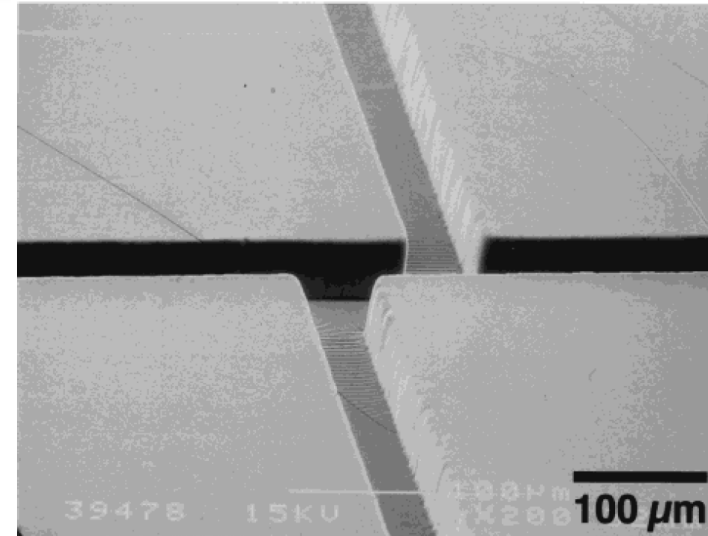
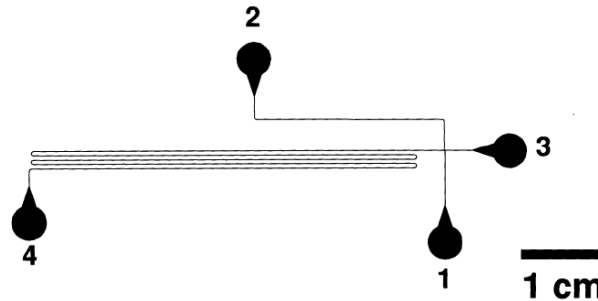


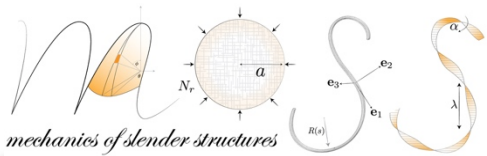
**E** Oxidize PDMS replica and flat in plasma and seal



replica

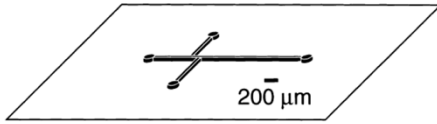
flat



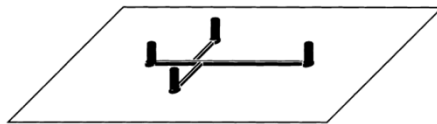


# Fabrication

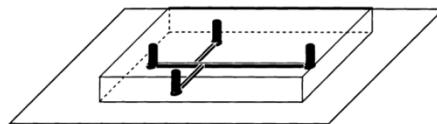
## A Fabricate master by rapid prototyping



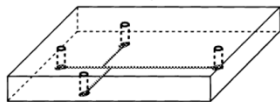
## B Place posts to define reservoirs



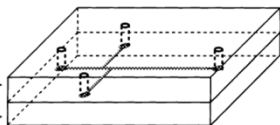
## C Cast prepolymer and cure



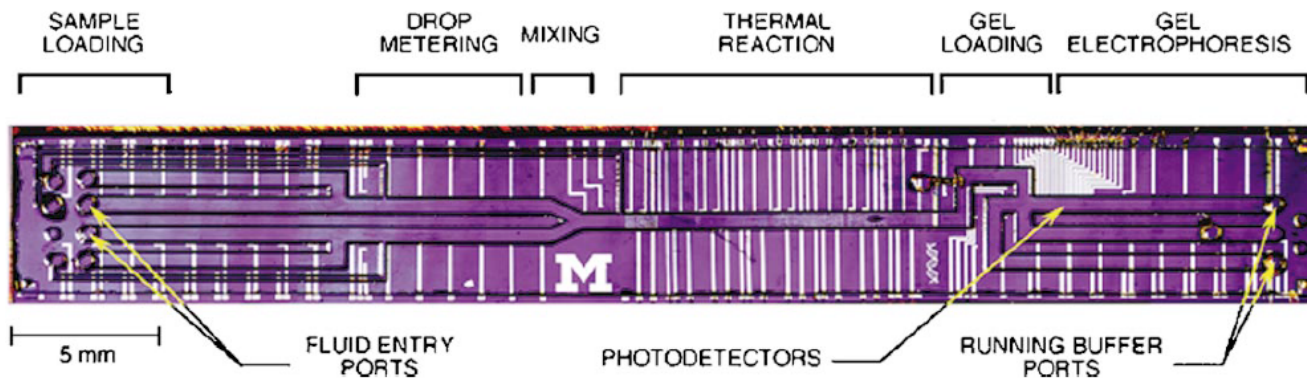
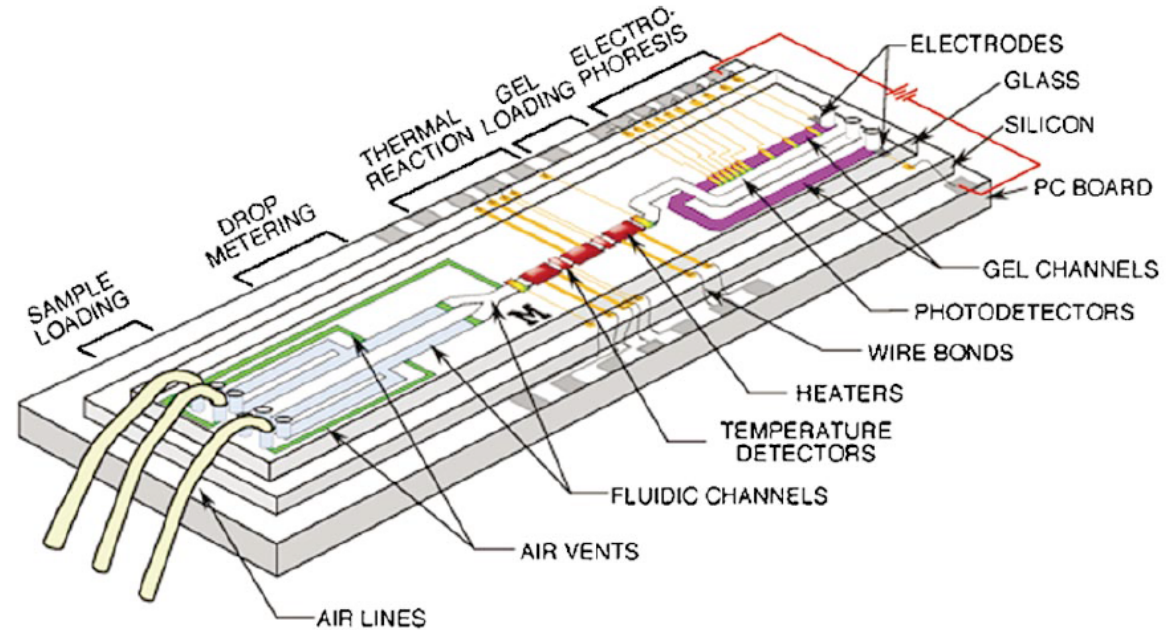
## D Remove PDMS replica from master



## E Oxidize PDMS replica and flat in plasma and seal



replica  
flat



Duffy, David C., et al. "Rapid prototyping of microfluidic systems in poly(dimethylsiloxane)." *Analytical chemistry* 70.23 (1998): 4974-4984.

M.A. Burns, C.H. Mastrangelo, T.S. Sammarco, F.P. Man, J.R. Webster, et al., "Microfabricated structures for integrated DNA analysis." *Proc. Natl. Acad. Sci. USA*, 93:5556-5561, (1996).

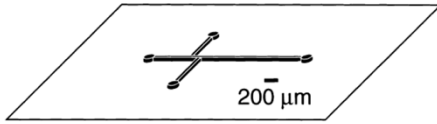




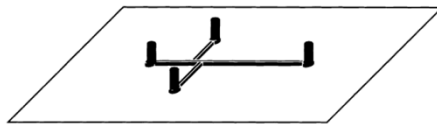


# Fabrication

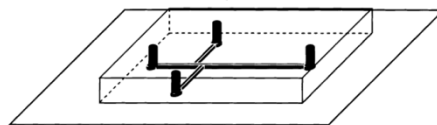
## A Fabricate master by rapid prototyping



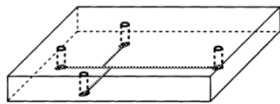
## B Place posts to define reservoirs



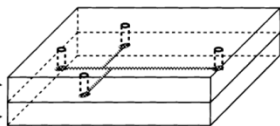
## C Cast prepolymer and cure



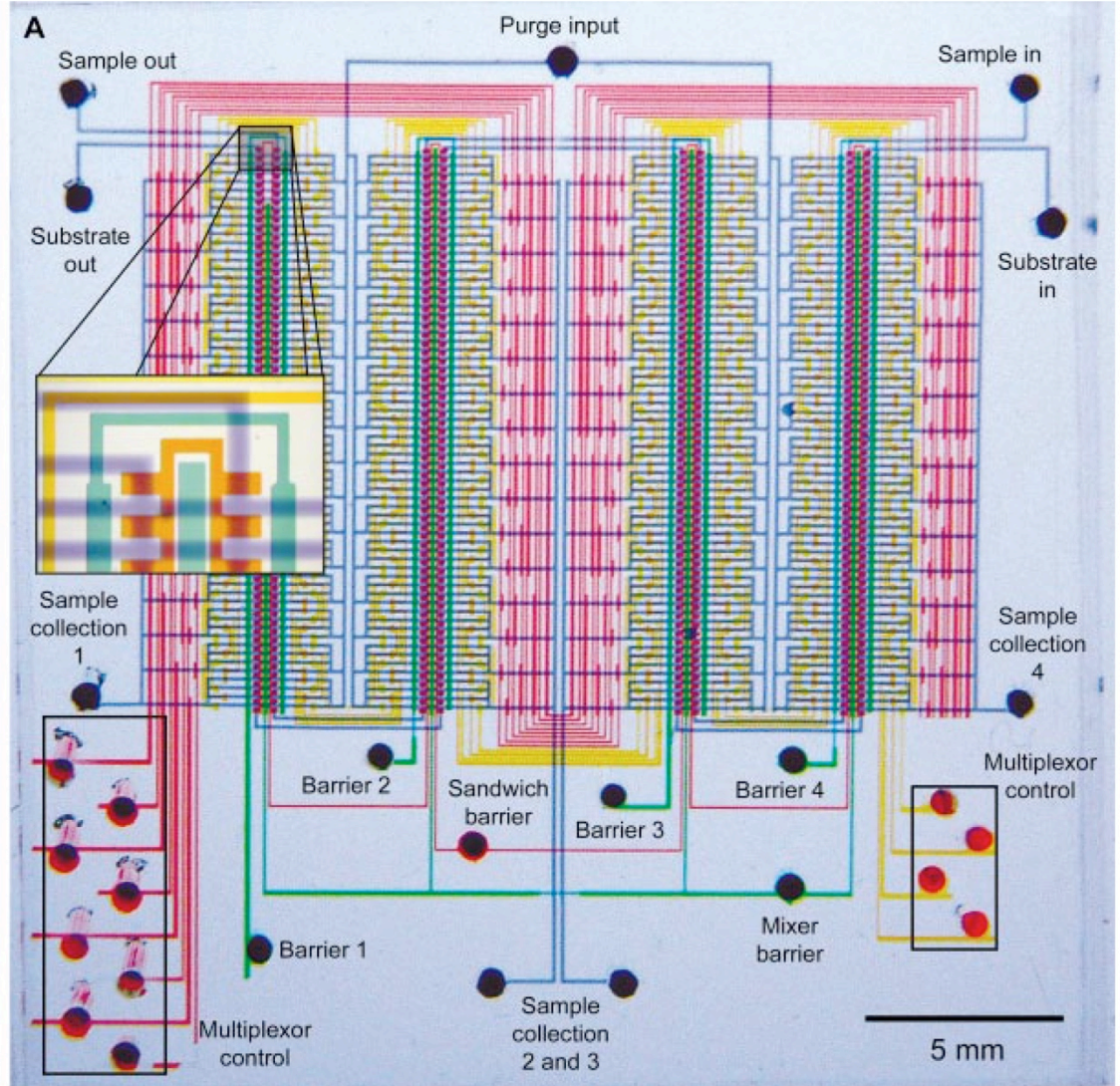
## D Remove PDMS replica from master



## E Oxidize PDMS replica in plasma and seal

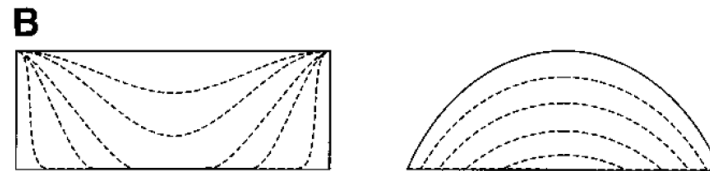
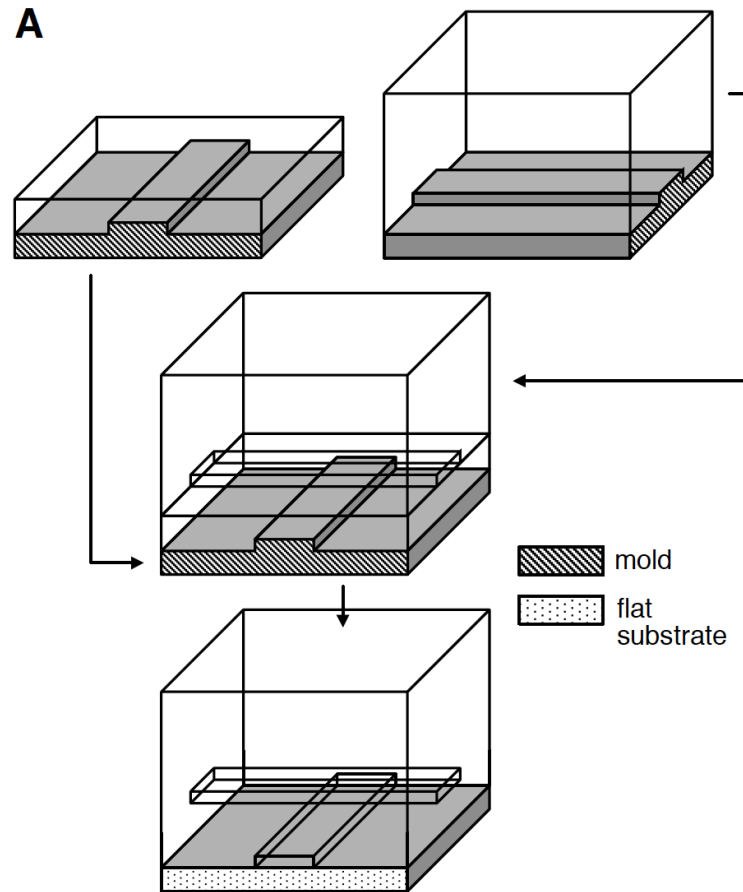


replica  
flat



## “Quake” valve

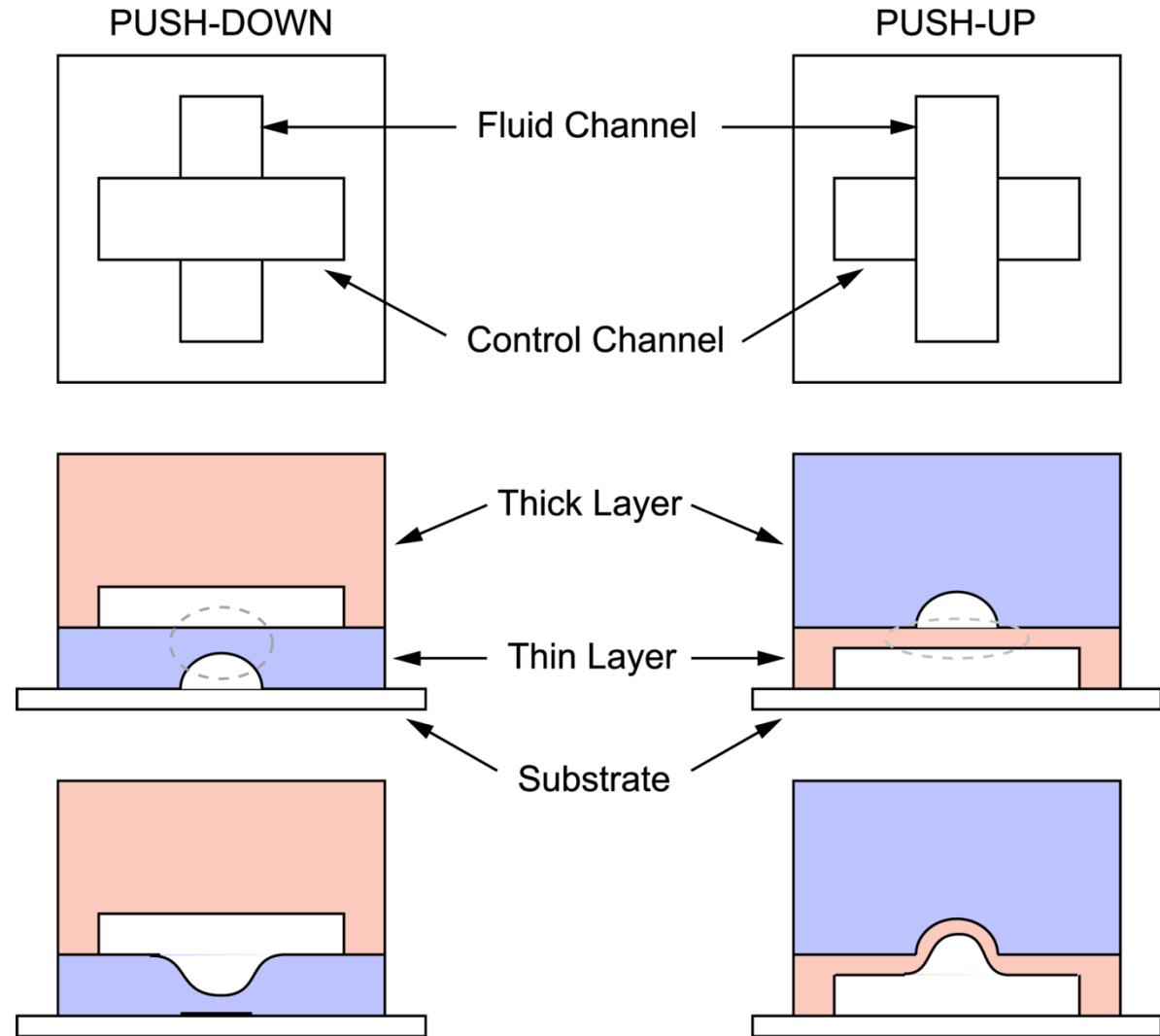
- Bilayer microfluidic chip.
- Thin film separating two flow channels.
  - One channel: Fluid
  - One channel: Air (controlling)
- Pressurized air deflects the thin film and closes the fluid channel.



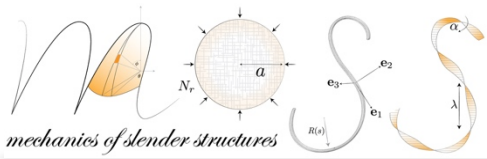
# Pneumatic Valves

## “Quake” valve

- Bilayer microfluidic chip.
- Thin film separating two flow channels.
  - One channel: Fluid
  - One channel: Air (controlling)
- Pressurized air deflects the thin film and closes the fluid channel.

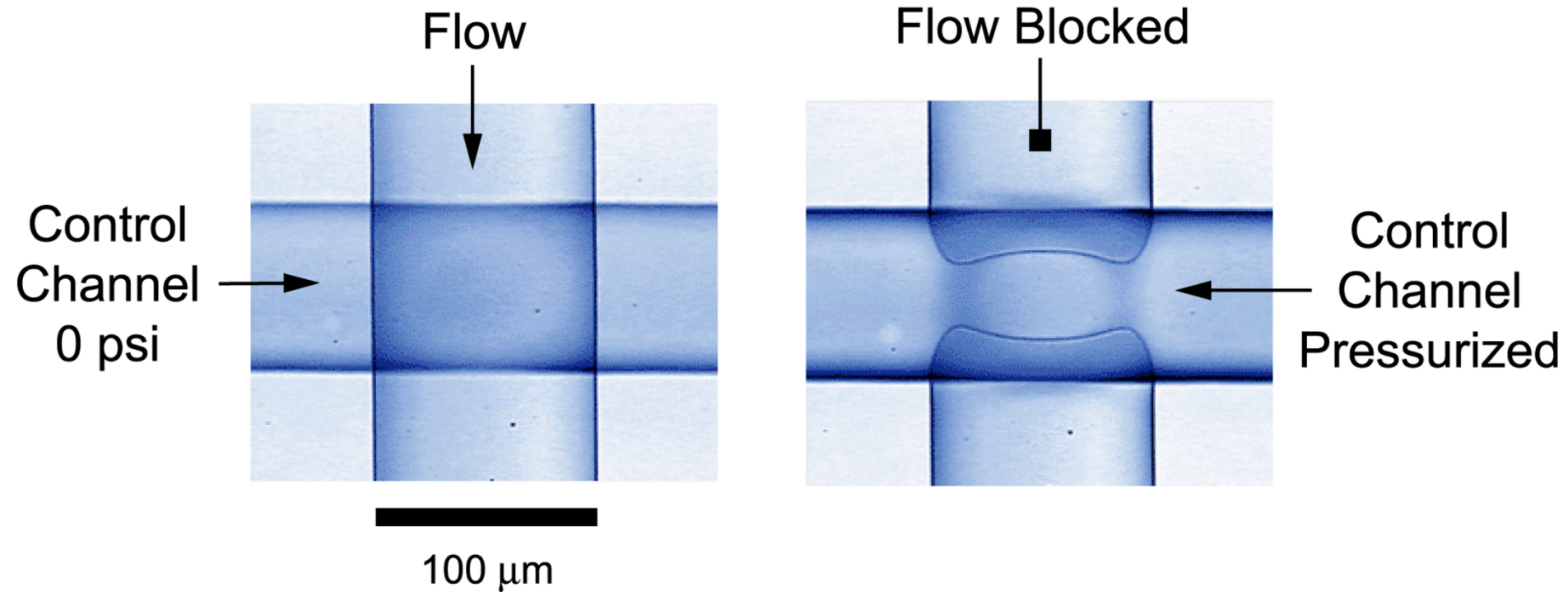






# Pneumatic Valves

## “Quake” valve



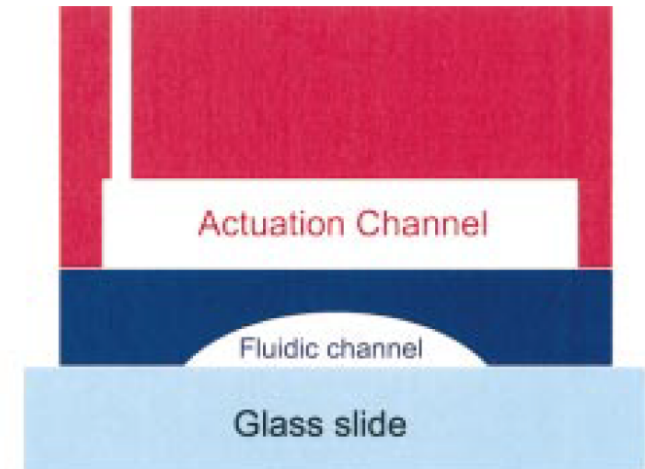


# Pneumatic Valves

## “Quake” valve

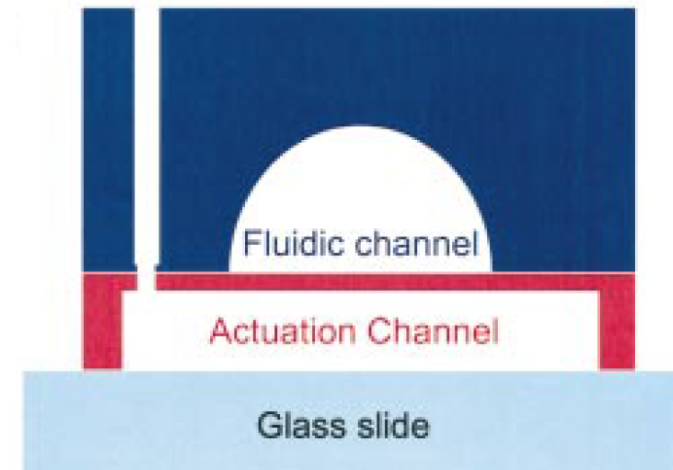
### Push-down Valves

- Control lines above flow channels.
- Pressure flattens membrane valve **down** to seal.
- Suitable for low aspect ratio (1:10) & shallow (~10um) channels.
  - Flow geometry: 100um x 13um
  - Control geometry: 100um x 10-25um
- Applications: where fluid flow must be in contact with substrate (spotting DNA, patterned substrate, etc.)



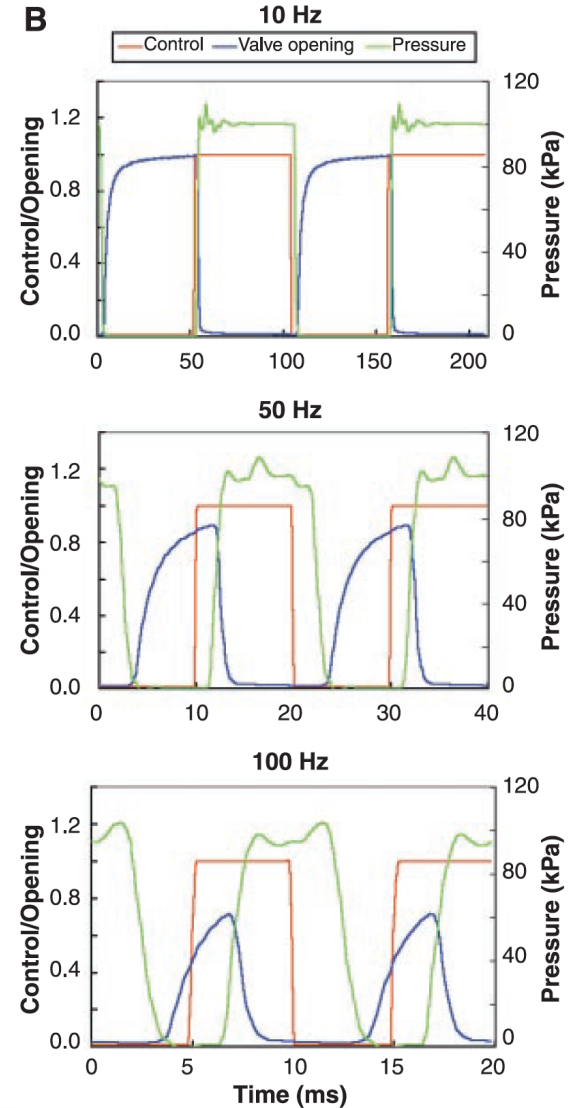
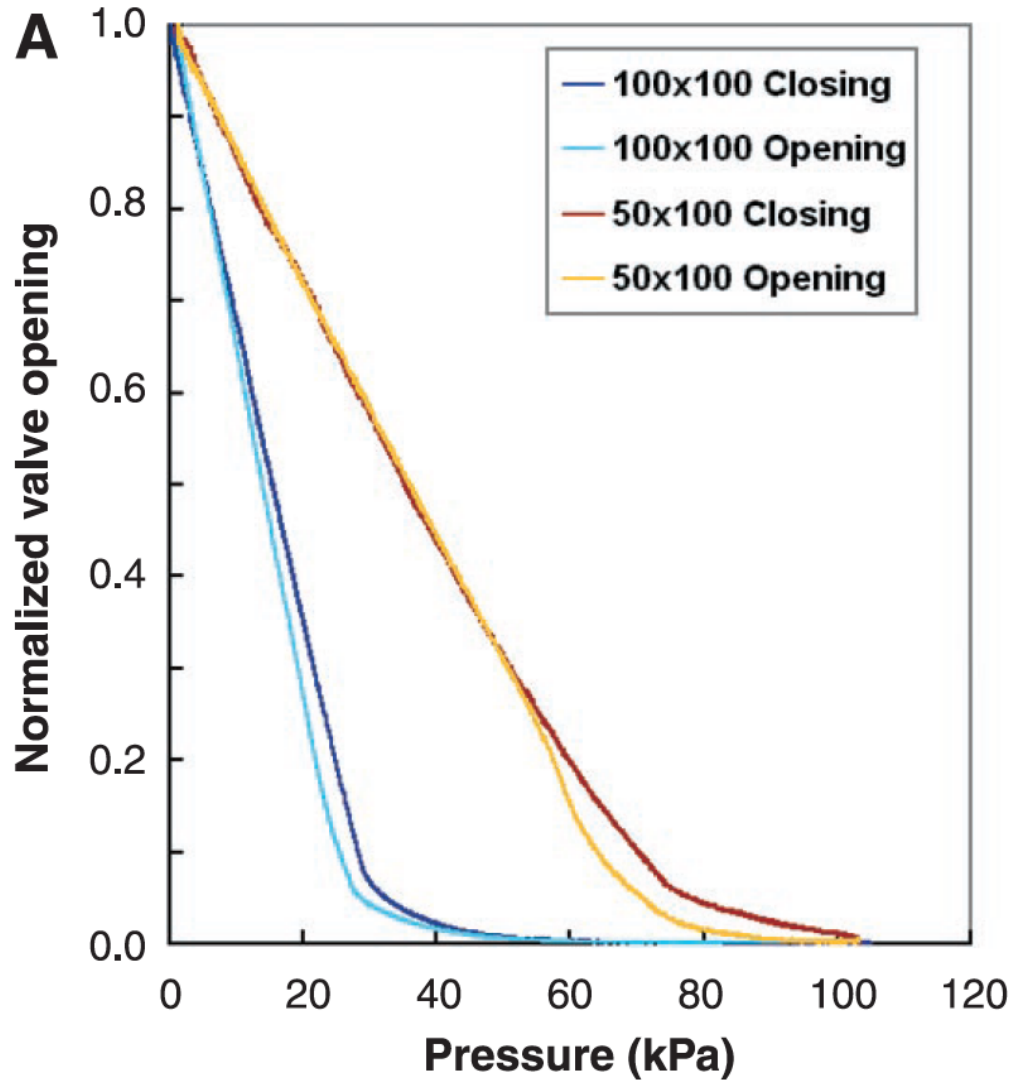
### Push-up Valves

- Control lines below flow channels.
- Pressure deflect membrane valve **up** to seal.
  - Flow geometry: 100um x 13um-50um
  - Control geometry: 100um x 10-25um
- Applications: suspension of large particles (eukaryotic cells, large beads, etc.)

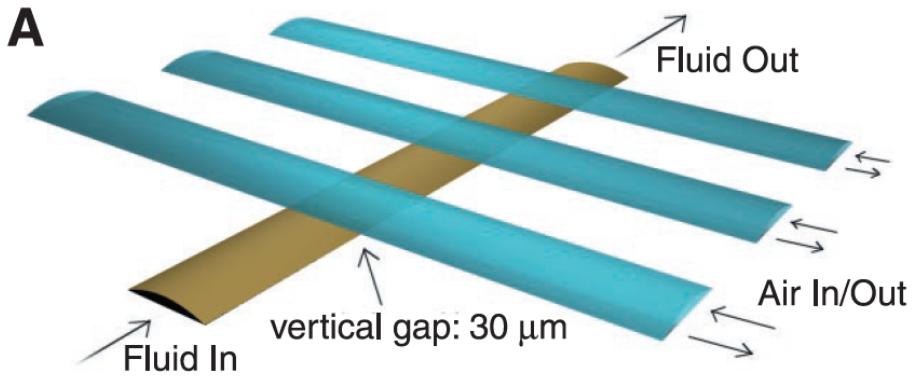




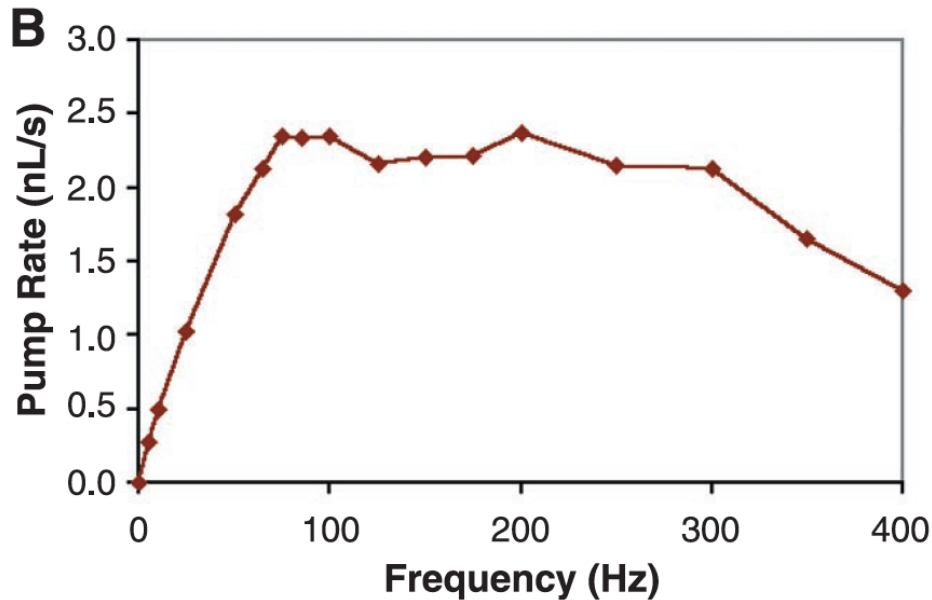
# Pneumatic Valves



# Pneumatic Pumps

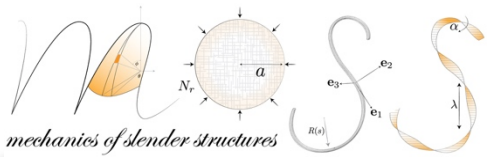


Multiple control channels – alternating pressure in/out.



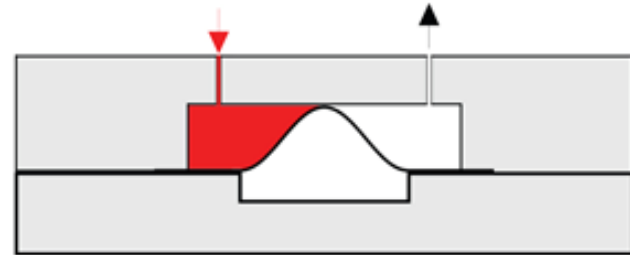
Peristaltic pumping:

- Frequency dependent flow rate

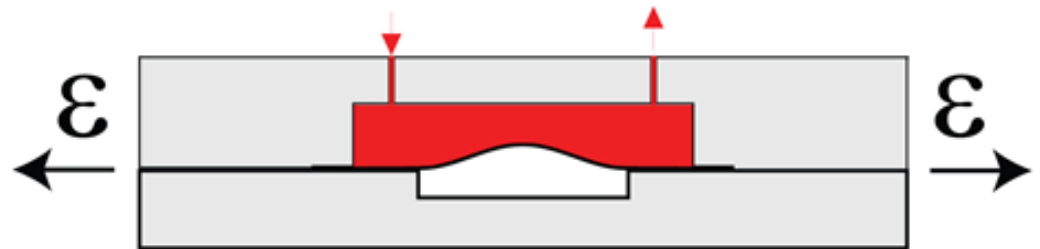


# Mechanical Valves

Flexible microfluidic device with single deformable arch.

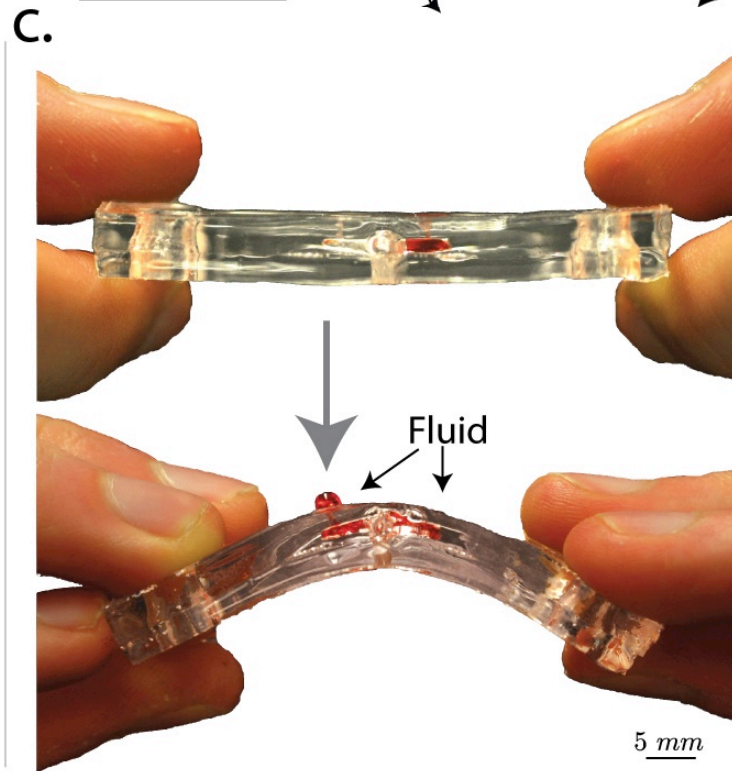
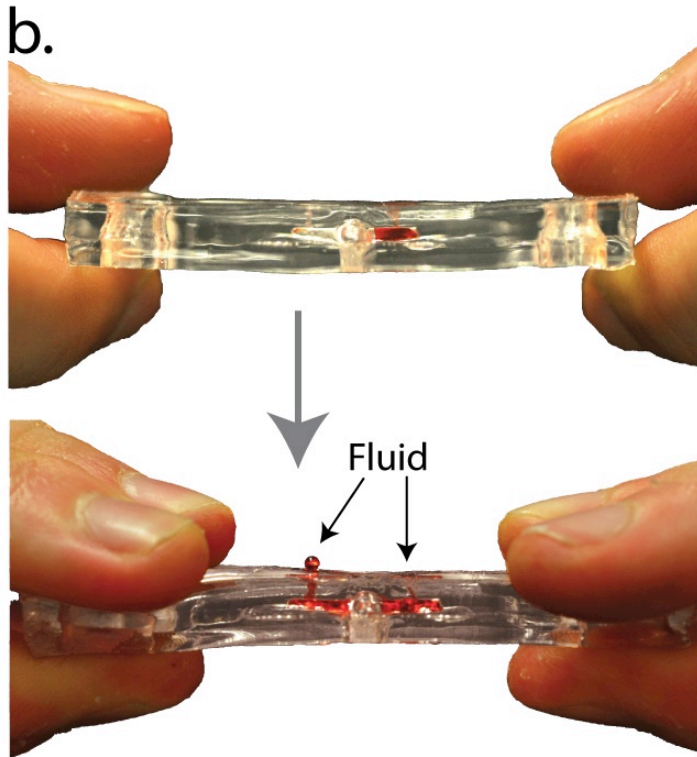
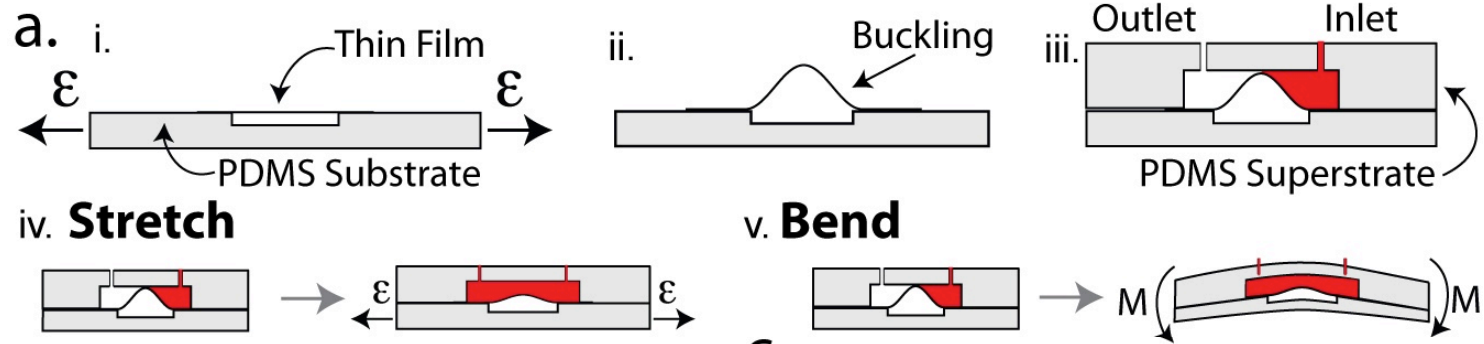


Stretching /bending the device reduces the arch height, partially opens the channel.



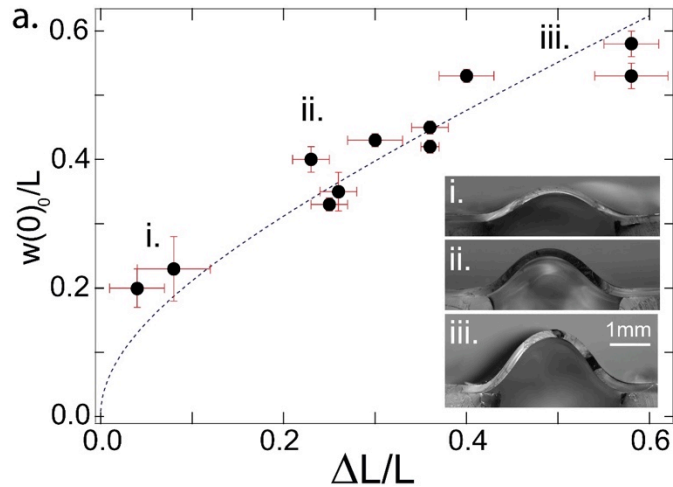


# Mechanical Valves





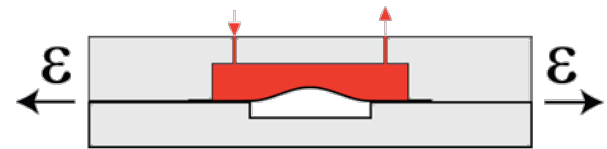
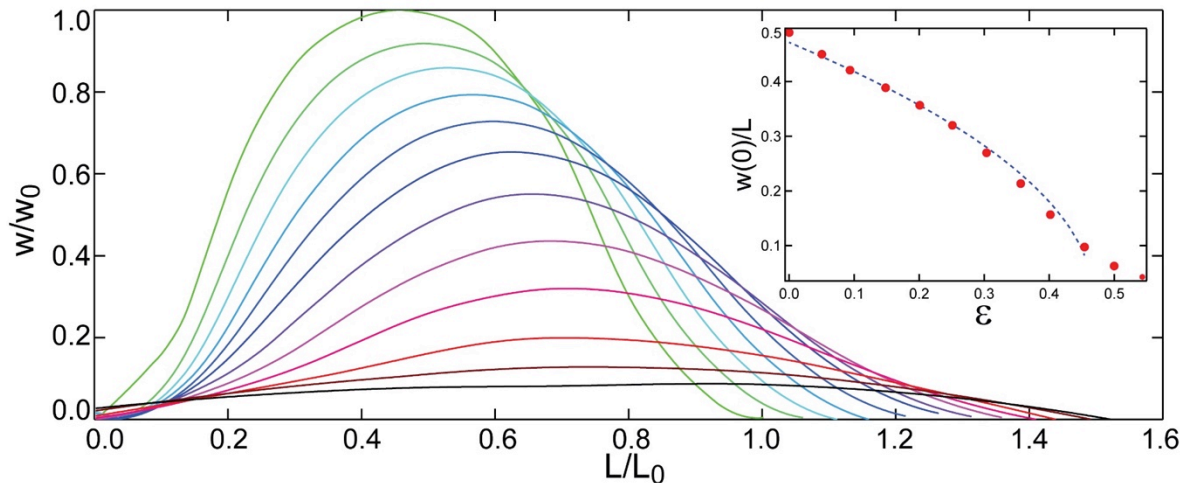
# Mechanical Valves



Buckled arch in microfluidic chamber.

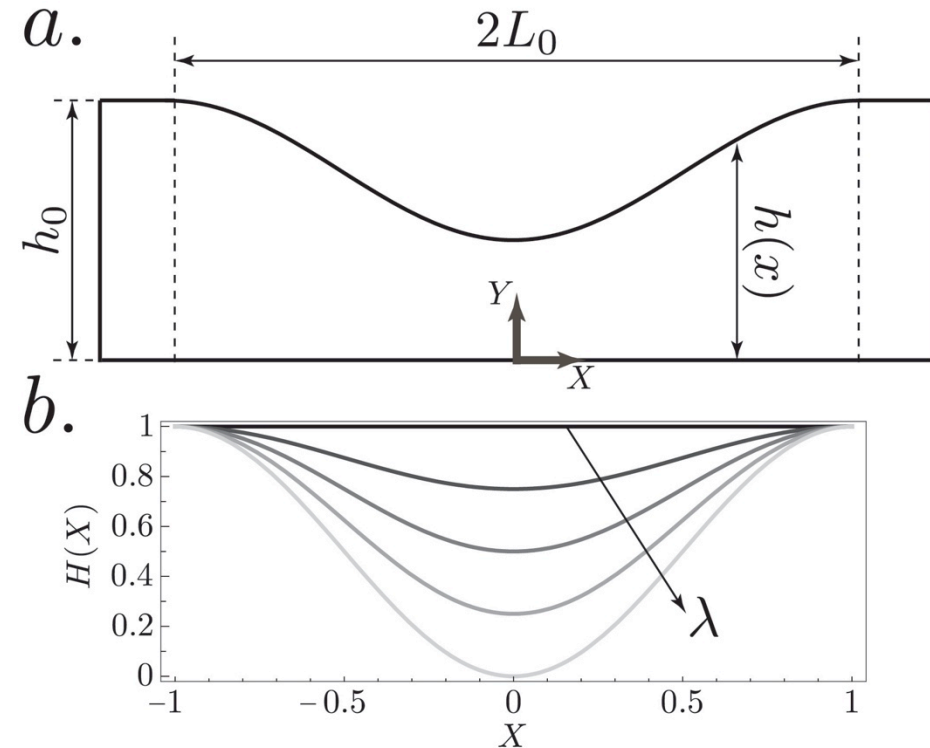
Arch shape obtained from buckling analysis of elastica. Apex:

$$w(0) = \frac{2}{\pi} \sqrt{\Delta L_0 (L_0 + \Delta L_0)}$$





# Mechanical Valves



## Navier-Stokes Equations:

...conservation of momentum...

$$\overbrace{\rho \left( \frac{\partial \mathbf{u}}{\partial t} + \mathbf{u} \cdot \nabla \mathbf{u} \right)}^{\text{Inertial acceleration}} = \overbrace{-\nabla p + \mu \nabla^2 \mathbf{u} + \mathbf{f}}^{\text{Forces}}$$

## Continuity Equation:

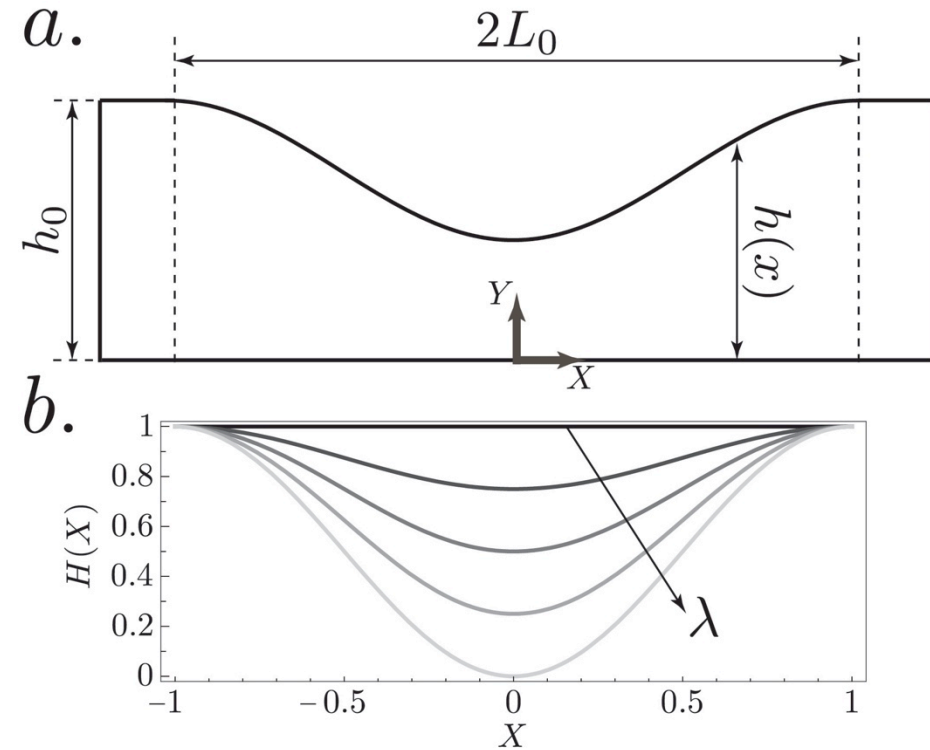
...conservation of mass...

$$\frac{\partial \rho}{\partial t} + \nabla \cdot (\rho \mathbf{u}) = 0$$

## Stokes Equations:

$$\mu \nabla^2 \mathbf{u} = \nabla p \quad \text{Neglect inertia \& body forces}$$

$$\nabla \cdot \mathbf{u} = 0 \quad \text{Incompressible: } \delta \rho / \rho \ll 1$$



Shape of the impingement:

$$H(X) = 1 - \frac{\lambda}{2} [1 + \cos(\pi X)]$$

Stokes Equations:

$$\mu \nabla^2 \mathbf{u} = \nabla p \quad \text{Neglect inertia \& body forces}$$

$$\nabla \cdot \mathbf{u} = 0 \quad \text{Incompressible: } \delta\rho/\rho \ll 1$$

Dimensionless Parameters:

Lengths:  $X = \frac{x}{L_0}, Y = \frac{y}{h_0}$

Velocities:  $U = \frac{u}{q_0/h_0}, V = \frac{v}{q_0/L_0}$

Pressure:  $P = \frac{p}{\Delta p} = \frac{p}{\mu q_0 L_0 / h_0^3}$

Dimensionless Equations:

$$\frac{\partial U}{\partial X} + \frac{\partial V}{\partial Y} = 0$$

$$\delta^2 \frac{\partial^2 U}{\partial X^2} + \frac{\partial^2 U}{\partial Y^2} = \frac{\partial P}{\partial X}$$

$$\delta^4 \frac{\partial^2 V}{\partial X^2} + \delta^2 \frac{\partial^2 V}{\partial Y^2} = \frac{\partial P}{\partial Y}$$

Boundary Conditions:

$$U = 0, V = 0 \text{ at } Y = 0 \text{ and } H(X) \quad \text{No slip}$$

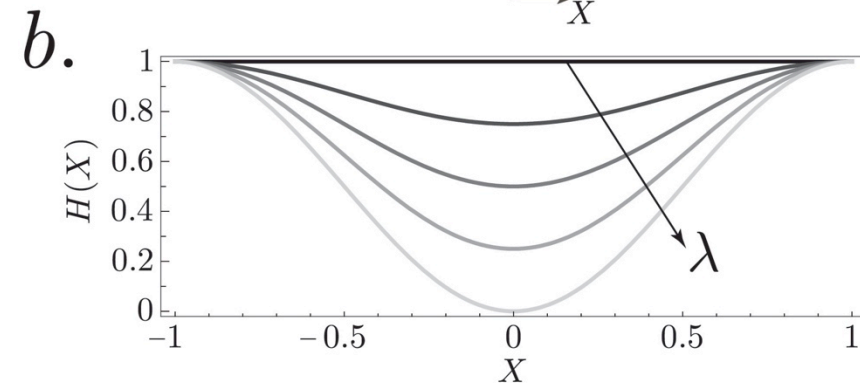
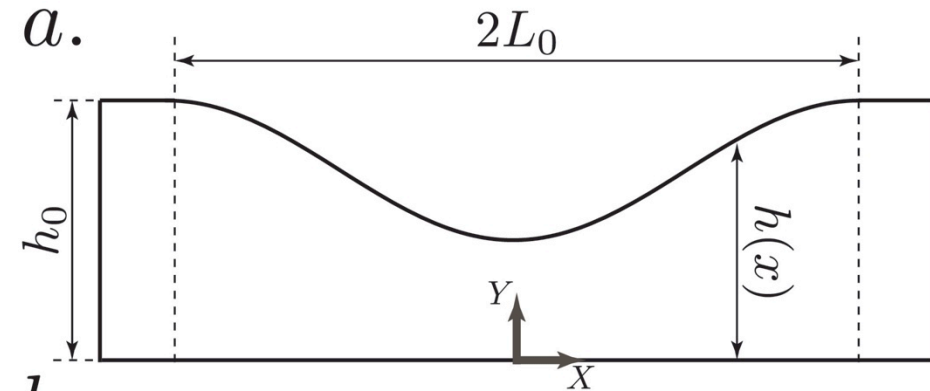
$$\int_0^{H(X)} U(X, Y) dY = 1 \quad \text{Total flow rate is prescribed}$$

Perturbation Expansion:

$$U(X, Y; \delta) = U_0(X, Y) + \delta^2 U_2(X, Y) + \delta^4 U_4(X, Y) + \dots$$

$$V(X, Y; \delta) = V_0(X, Y) + \delta^2 V_2(X, Y) + \delta^4 V_4(X, Y) + \dots$$

$$P(X, Y; \delta) = P_0(X, Y) + \delta^2 P_2(X, Y) + \delta^4 P_4(X, Y) + \dots$$



Shape of the impingement:

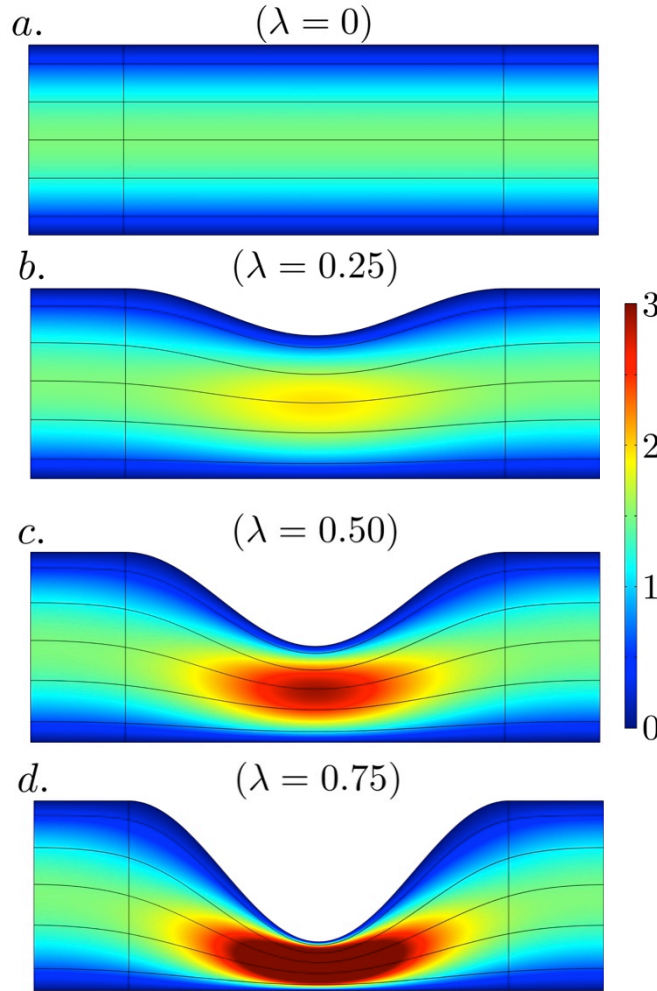
$$H(X) = 1 - \frac{\lambda}{2} [1 + \cos(\pi X)]$$





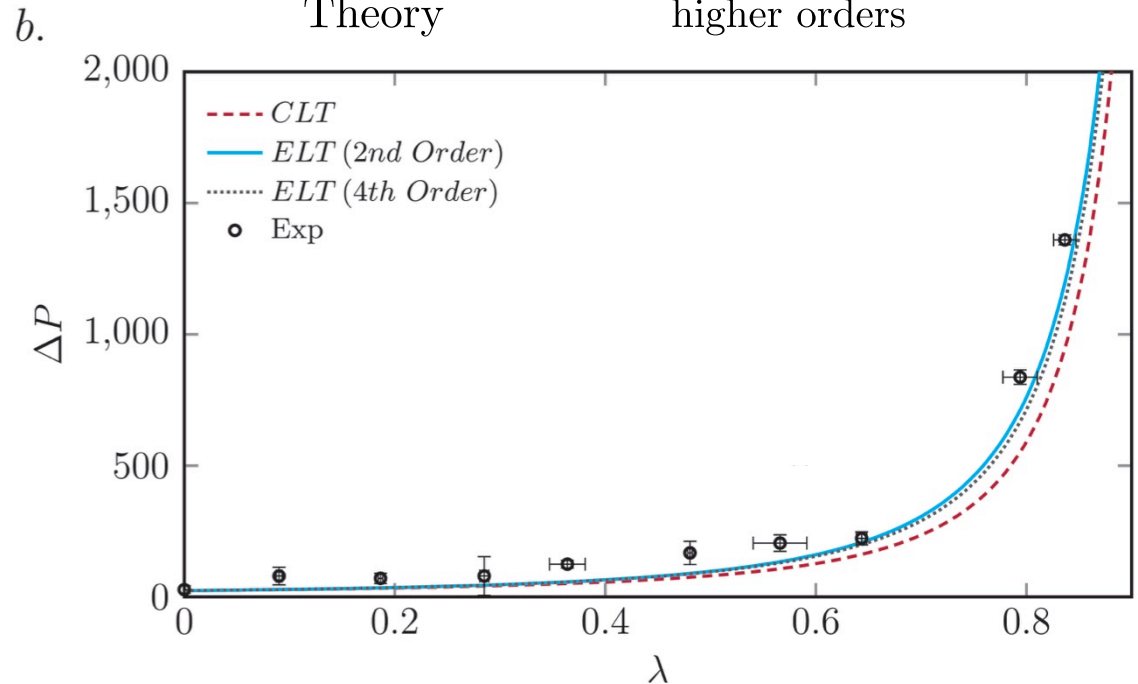
# Extended Lubrication

## Fluid flow through channels with variable geometry



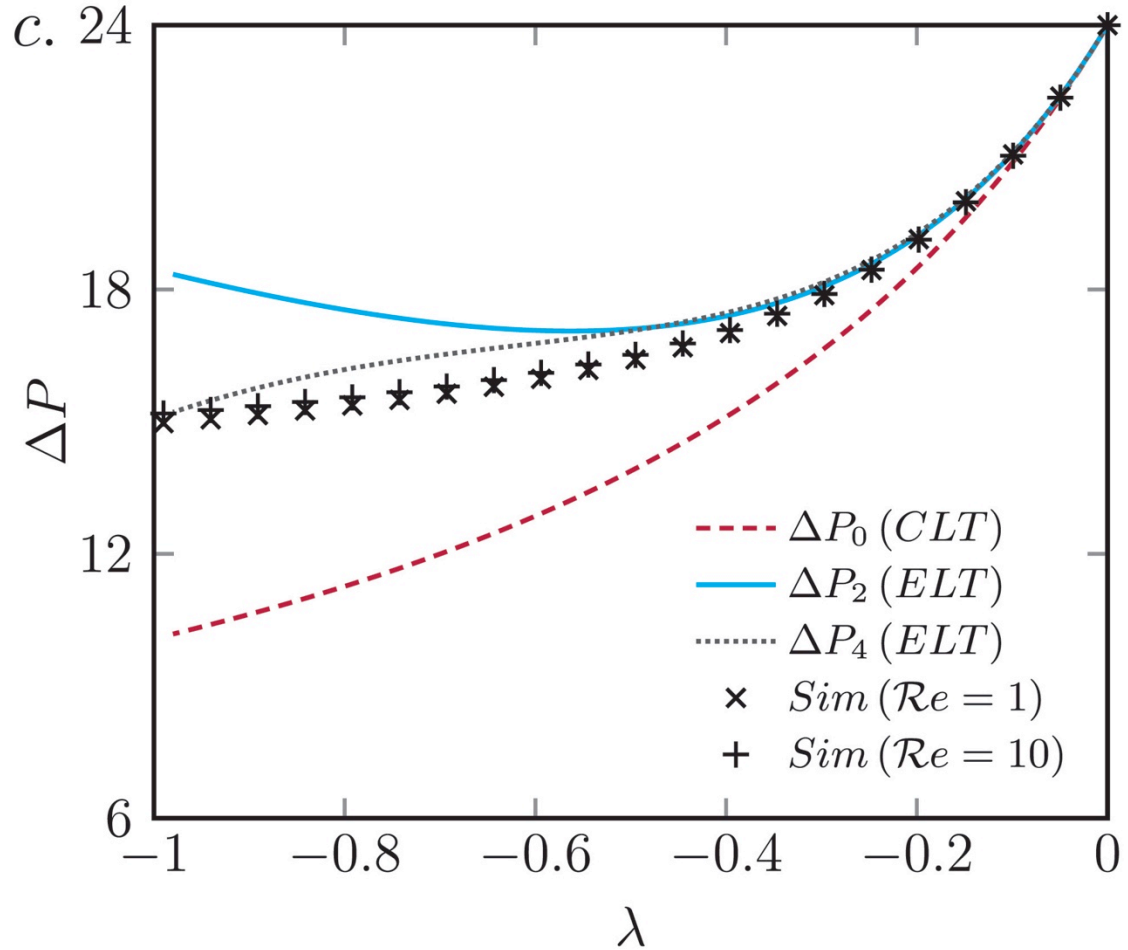
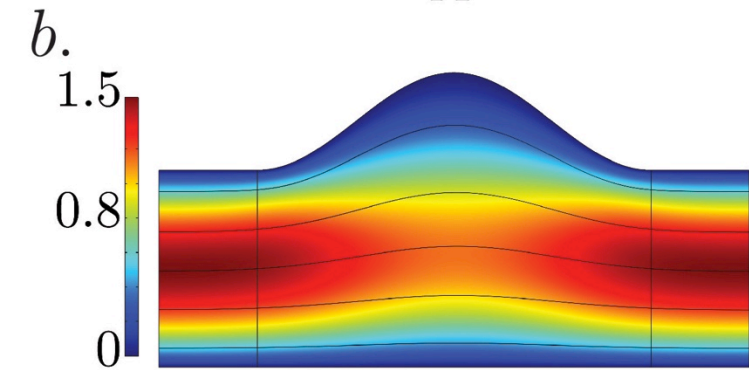
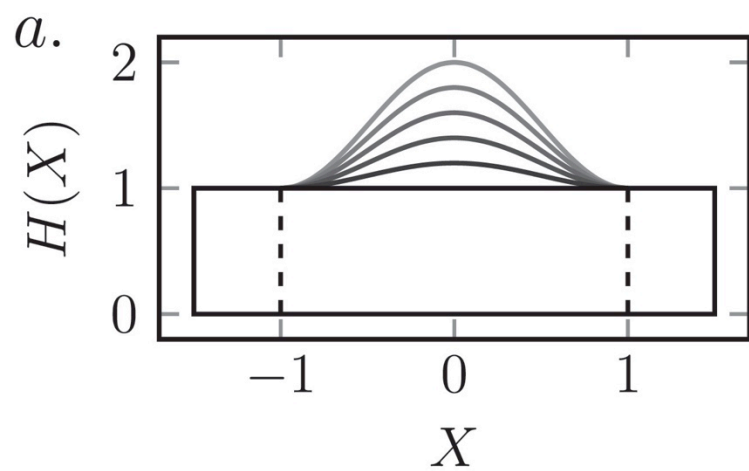
### Extended Lubrication Theory

$$\Delta P = \underbrace{\Delta P_0}_{\text{Lubrication Theory}} \left( 1 + \underbrace{\frac{4}{5} \lambda^2 \delta^2 - \frac{64}{225} \lambda^4 \delta^4 + \mathcal{O}(\delta^6)}_{\text{Perturbation to higher orders}} \right)$$



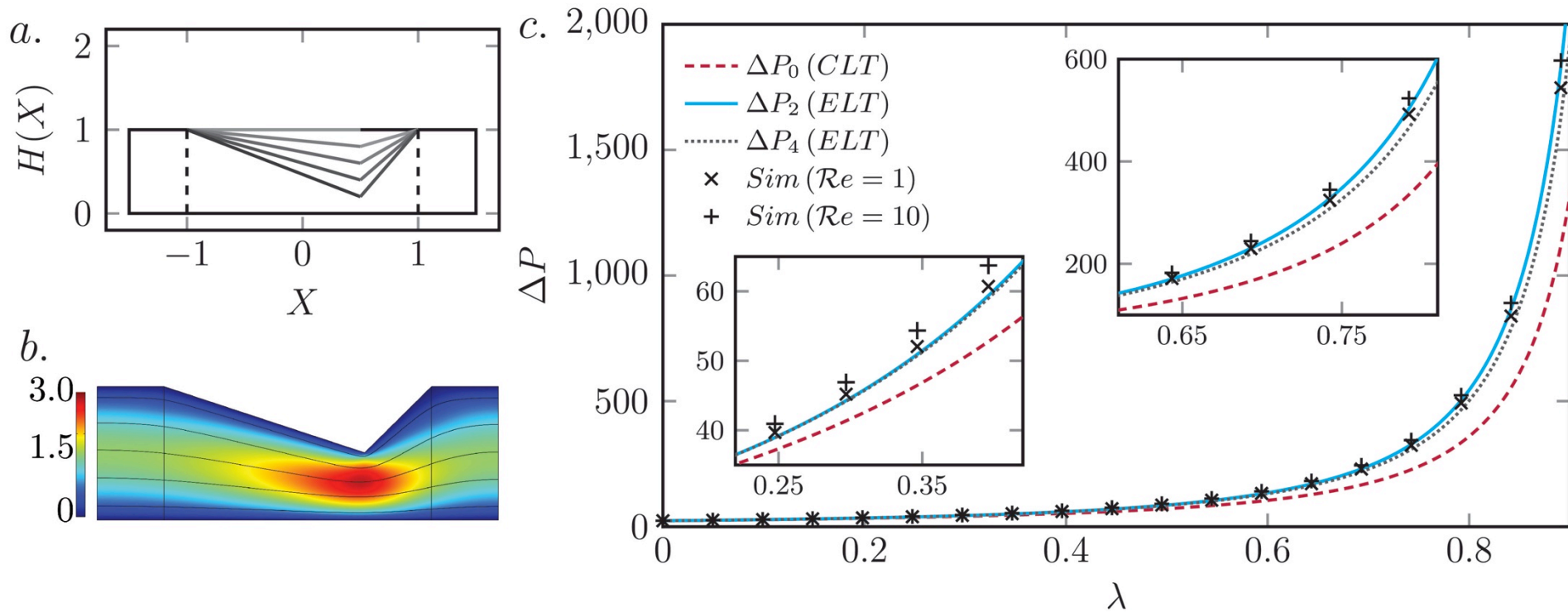


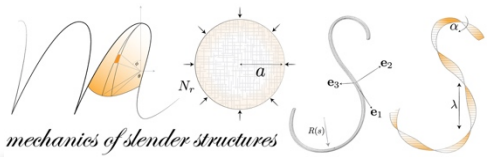
# Extended Lubrication



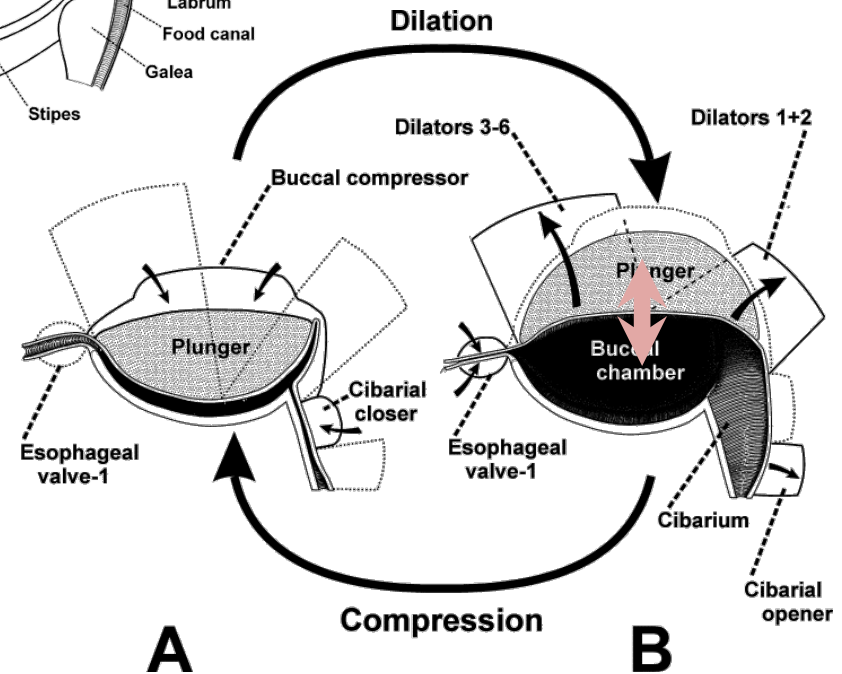
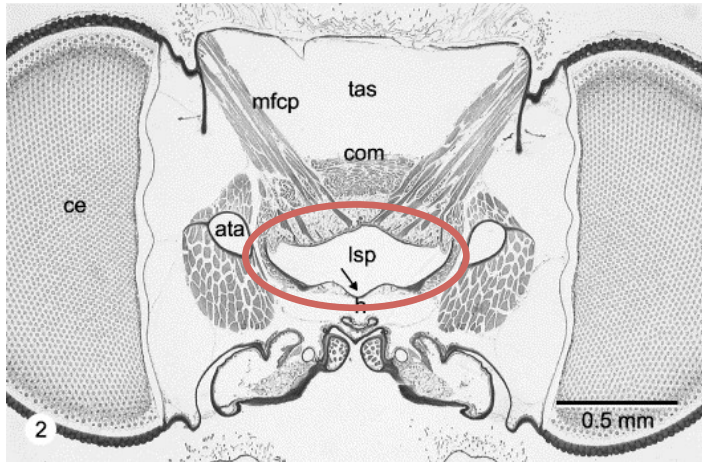
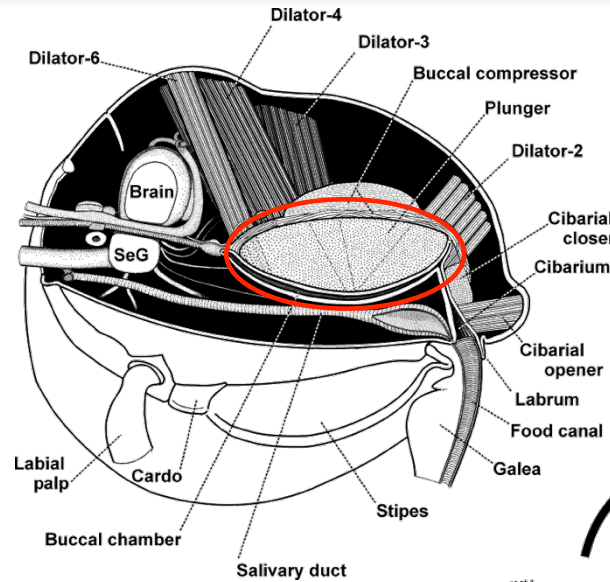


# Extended Lubrication





# Bioinspiration



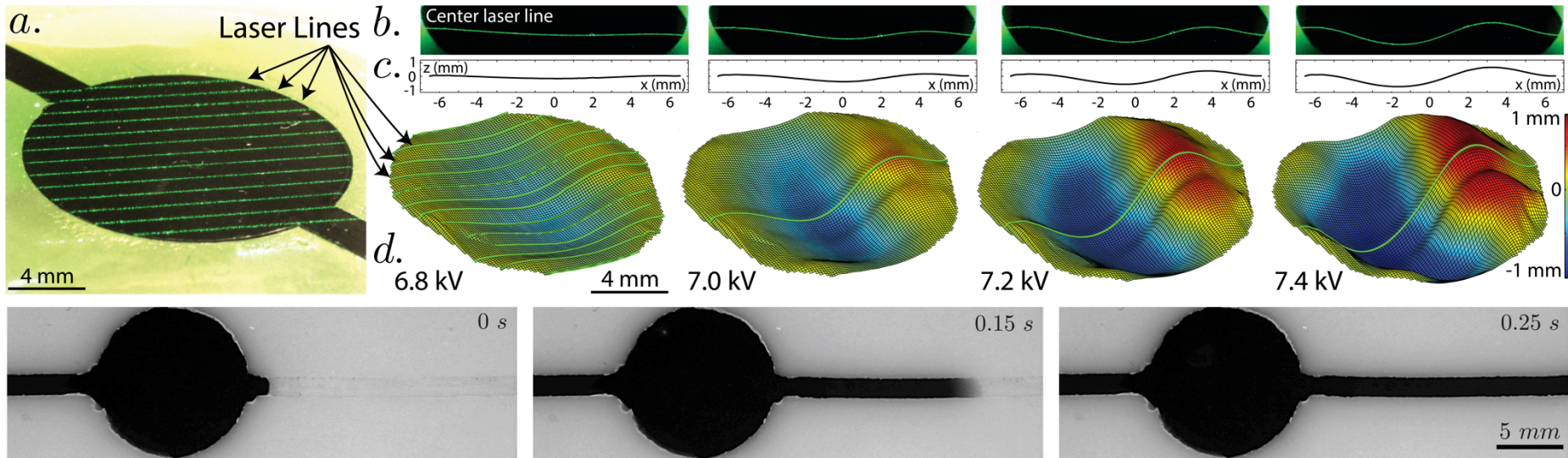
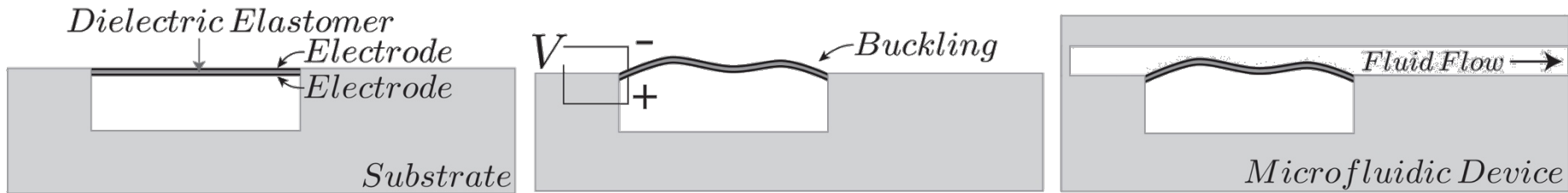
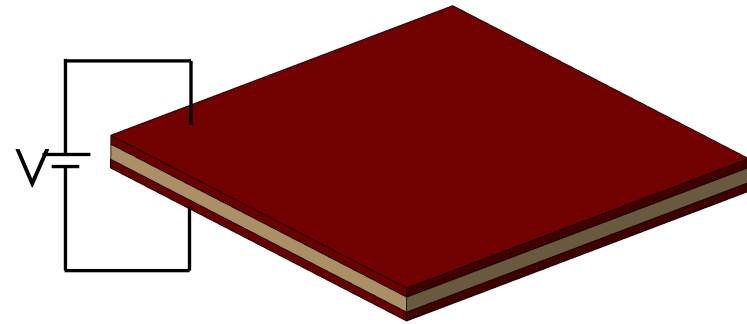
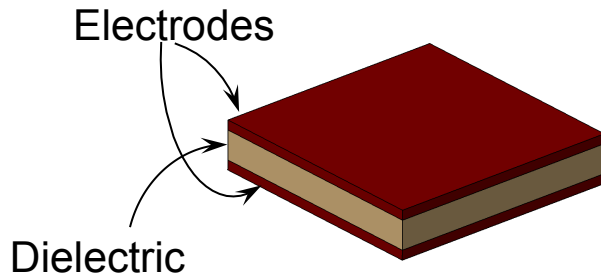
**Large, rapid, reversible diaphragm deformation for fluid pumping.**

Davis, N. T. and J. G. Hildebrand (2006). "Neuroanatomy of the sucking pump of the moth, *Manduca sexta* (Sphingidae, Lepidoptera)." *Arthropod Structure & Development*.

Eberhard, S. H. and H. W. Krenn (2005). "Anatomy of the oral valve in nymphalid butterflies and a functional model for fluid uptake in Lepidoptera." *Zoologischer Anzeiger - A Journal of Comparative Zoology* 243(4): 305-312.



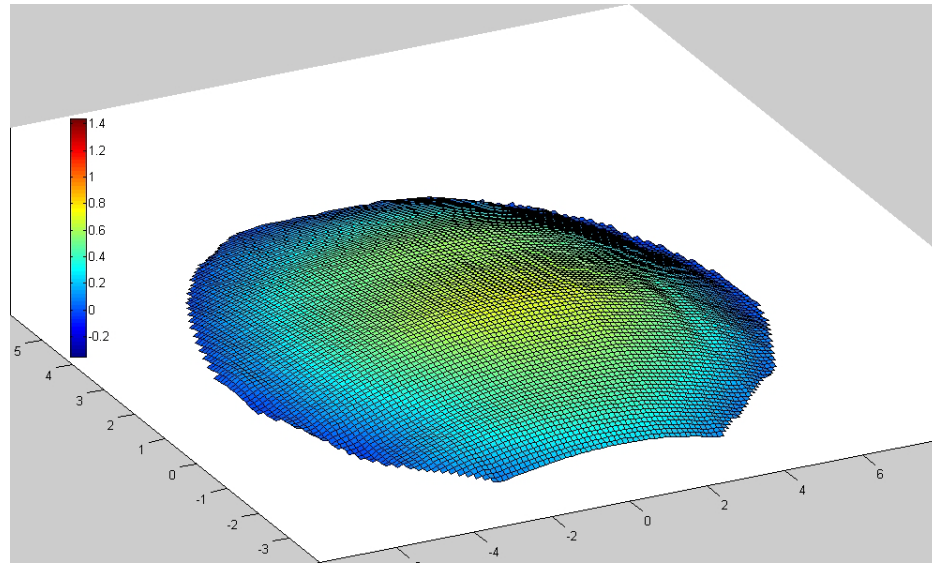
# Electrical Valves



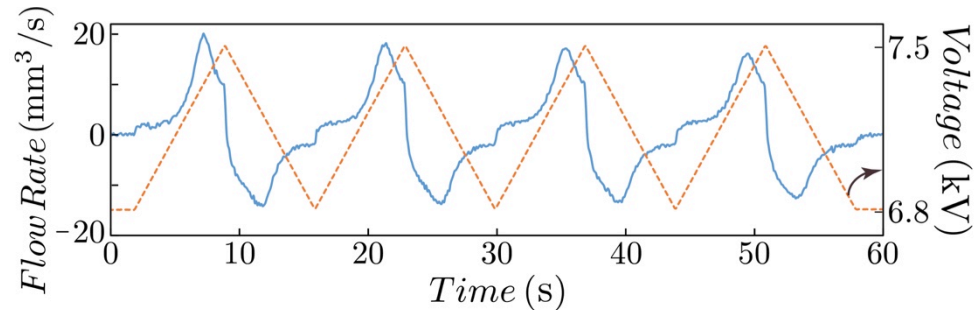
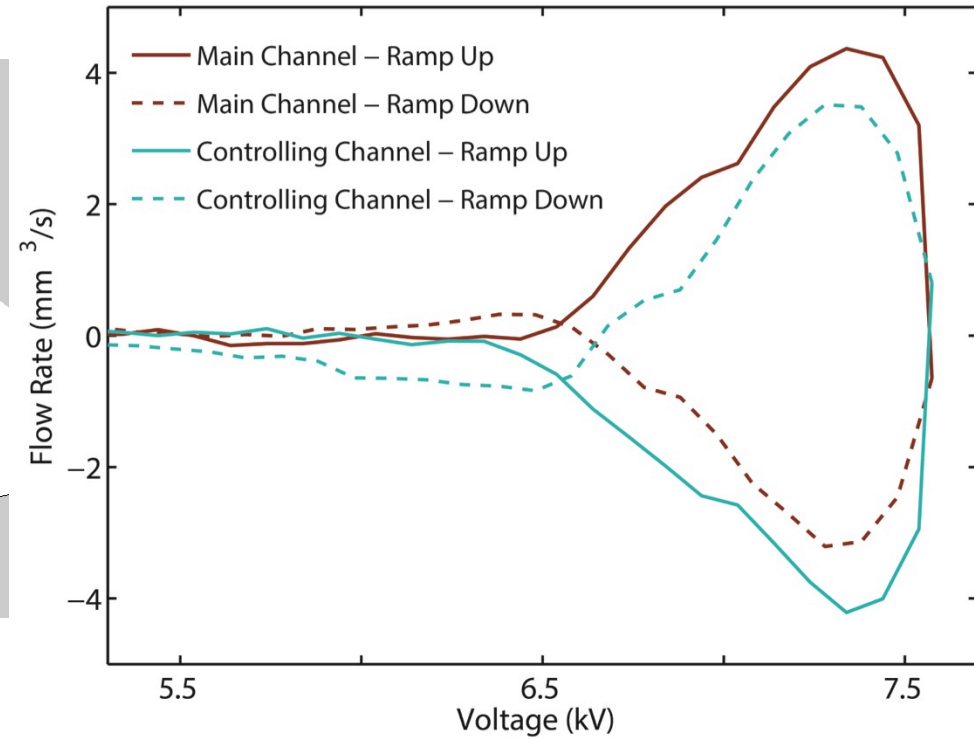
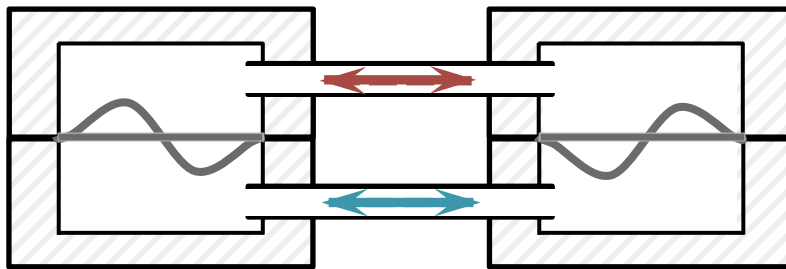


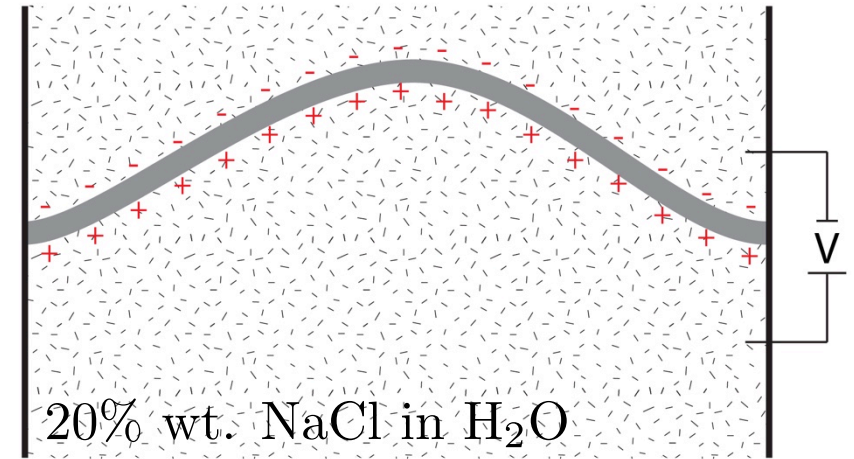
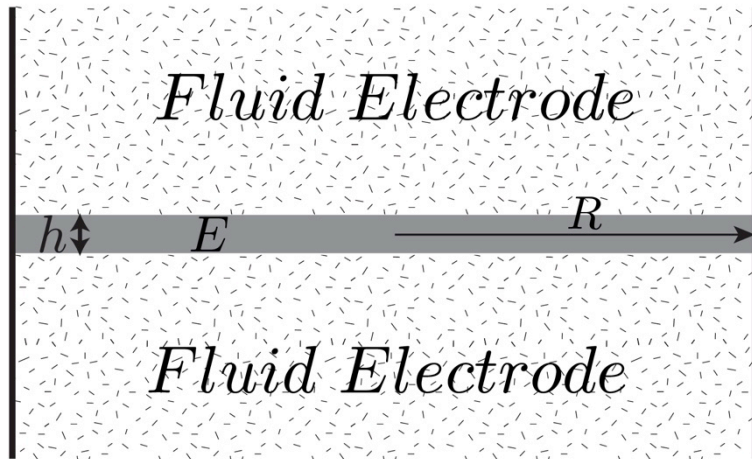


# Electrical Valves



**Coupled Chambers**





At what **voltage** will the plate **buckle**?

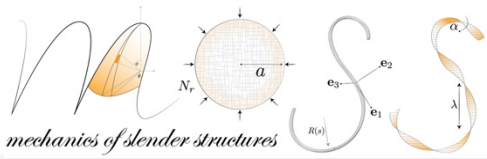
$$\varepsilon_r \approx \frac{\epsilon_0 \epsilon}{2E} \left( \frac{V}{h} \right)^2 \rightarrow \sigma_r \approx \frac{\epsilon_0 \epsilon}{2(1 - \nu^2)} \left( \frac{V}{h} \right)^2 \rightarrow \sigma_c = \frac{kEh^2}{12(1 - \nu^2)R^2}$$

Radial strain in a dielectric elastomer

Plane stress relation

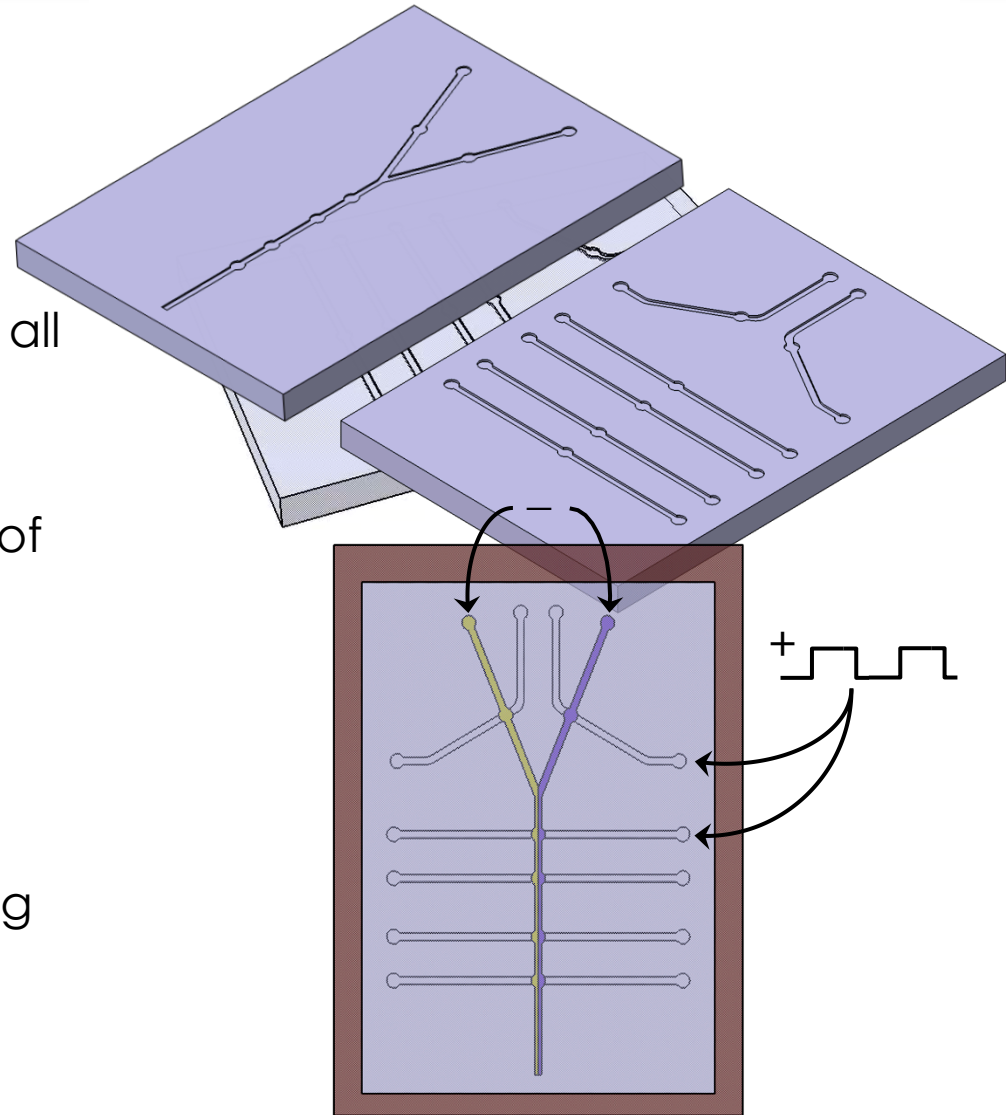
Linear stability analysis for a buckling plate

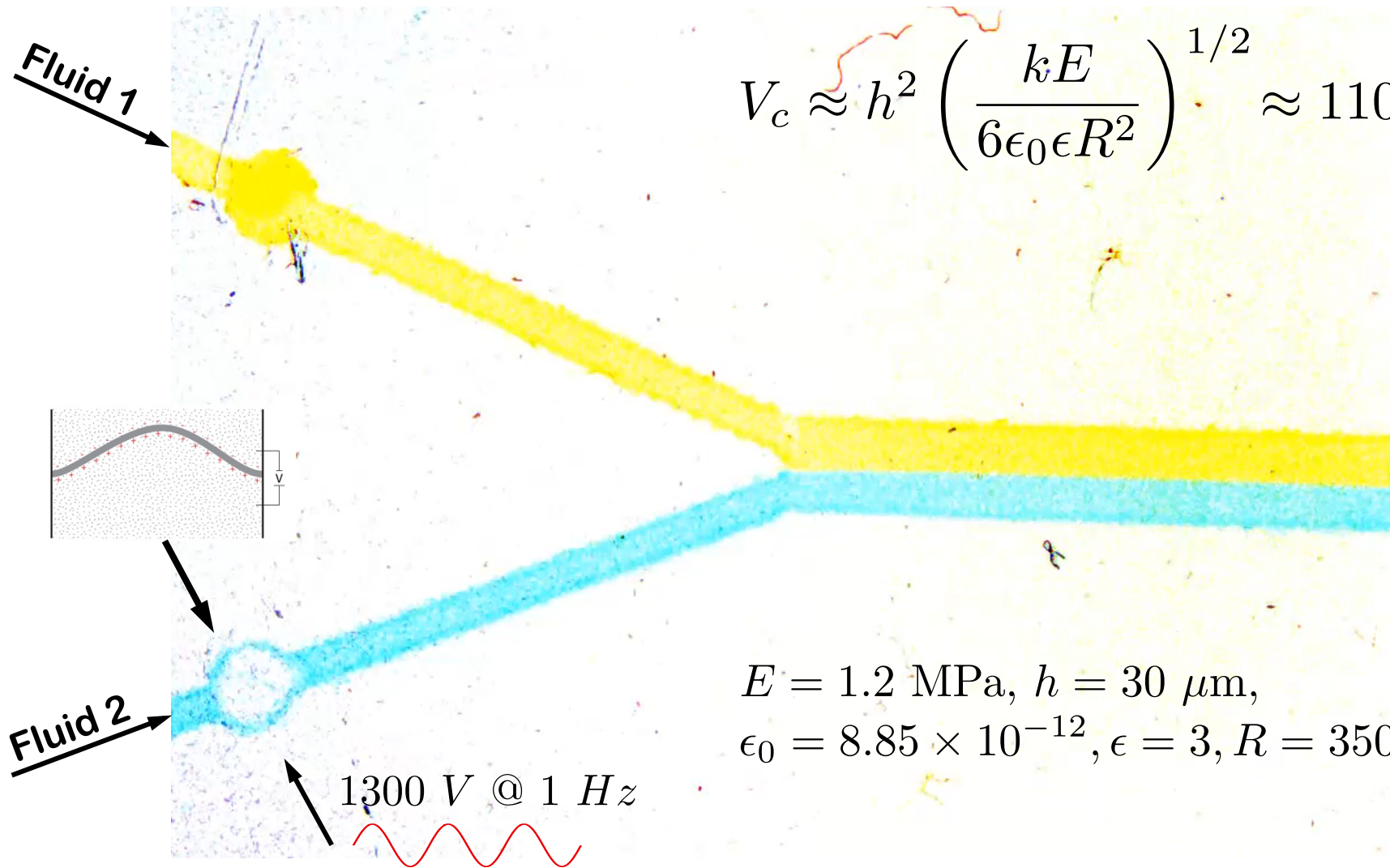
$$V_c \approx h^2 \left( \frac{kE}{6\epsilon_0 \epsilon R^2} \right)^{1/2}$$



# Microfluidic Fabrication

1. Prepare the **top** substrate with the **microfluidic** channel.
2. Prepare the **bottom** substrate with all **controlling** channels.
3. Make the thin, **dielectric film**.
4. Bond the substrates on both sides of the dielectric film.
  - Film will **buckle** at the intersection between channels.
5. Fill the channels with conductive fluids.
6. Apply a voltage to induce buckling



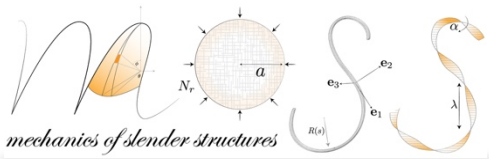


$$V_c \approx h^2 \left( \frac{kE}{6\epsilon_0\epsilon R^2} \right)^{1/2} \approx 1100 \text{ V}$$

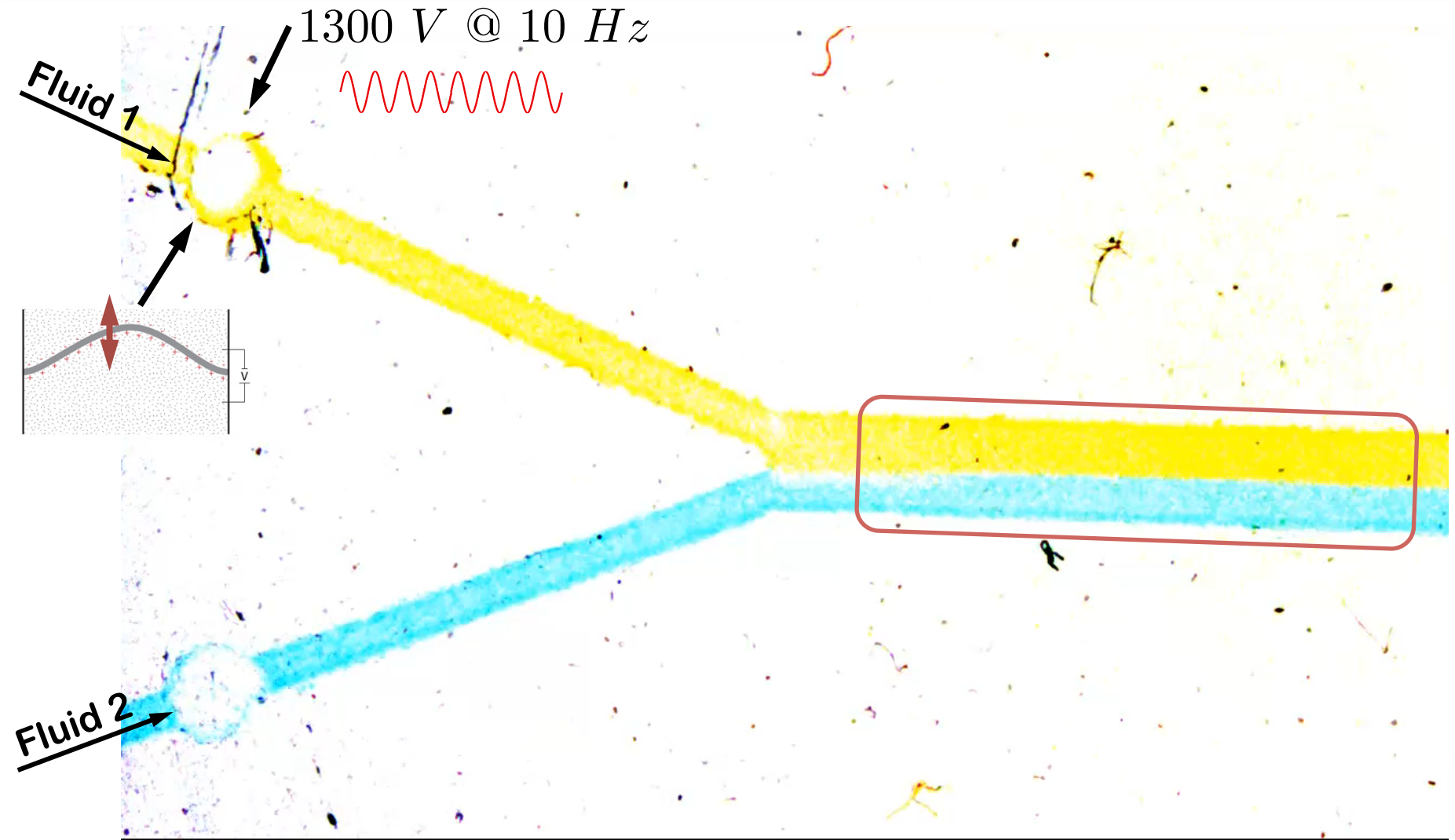
$$E = 1.2 \text{ MPa}, h = 30 \text{ }\mu\text{m},$$

$$\epsilon_0 = 8.85 \times 10^{-12}, \epsilon = 3, R = 350 \text{ }\mu\text{m}$$



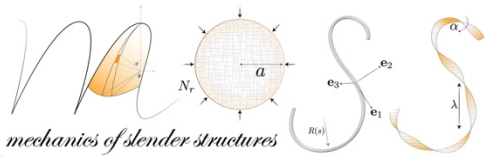


# Pumping



B. Tavakol, A. Chawan, and D. P. Holmes, "Buckling Instability of Thin Films as a Means to Control or Enhance Fluid Flow within Microchannels," in preparation, (2015).





## Confined Fluid Flow: Microfluidics and Capillarity

**Reynolds** Number: Inertia vs. Viscous effects

- Review of characteristic flows...

**Péclet** Number: Transport phenomena in a continuum

- Diffusion, separation, and mixing...

**Geometric** confinement: Controlling and manipulating fluid flow

- Microfluidic fabrication, valving, pumping...

**Capillary** Number: Viscosity vs. Surface tension

- Droplet formation, capillary rise, elasticity...



# Fluid Behavior

## Viscous Forces



Coiling honey

## Interfacial Forces



Wetting of water on a textured surface

**Capillary Number:** viscous/interfacial

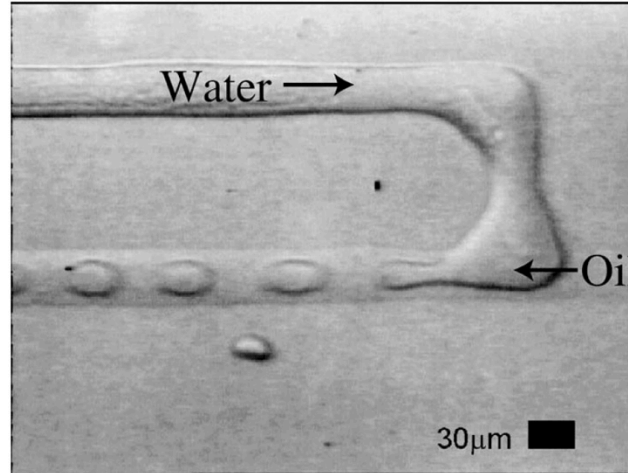


# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

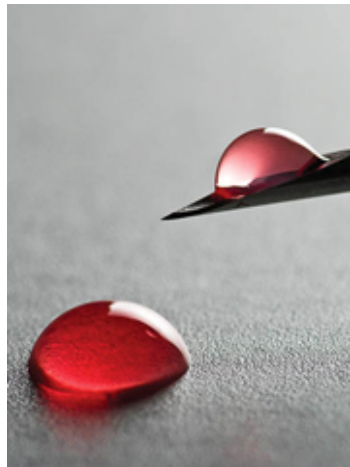


Monodisperse droplet generation

- Droplet emulsions in immiscible fluids
- Injection of water into stream of oil

**Interfacial tension** prevents the fluids from flowing alongside each other.

## Interfacial Forces



**Surface tension** acts to reduce the interfacial area.

$$\sigma_c \sim \gamma/R$$

**Viscous stresses** act to extend and drag the interface downstream.

$$\sigma_v \sim \mu U_0/h$$

**Characteristic droplet size:**

$$R \sim \frac{\gamma}{\mu U_0} h = \frac{h}{\mathcal{C}}$$

**Capillary number:**

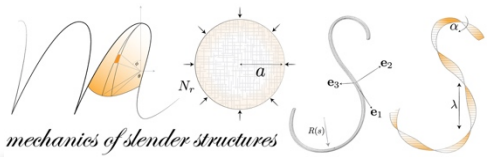
$$\mathcal{C} \equiv \frac{\mu U_0}{\gamma}$$

Squires, Todd M., and Stephen R. Quake. "Microfluidics: Fluid physics at the nanoliter scale." *Reviews of modern physics* 77.3 (2005): 977.

Thorsen, Todd, et al. "Dynamic pattern formation in a vesicle-generating microfluidic device." *Physical review letters* 86.18 (2001): 4163.

Honey: <http://www.honeyassociation.com/webimages/honey-dipper.jpg>

Droplet: <http://www.rycobel.be/en/technical-info/articles/1337/measuring-dynamic-absorption-and-wetting>

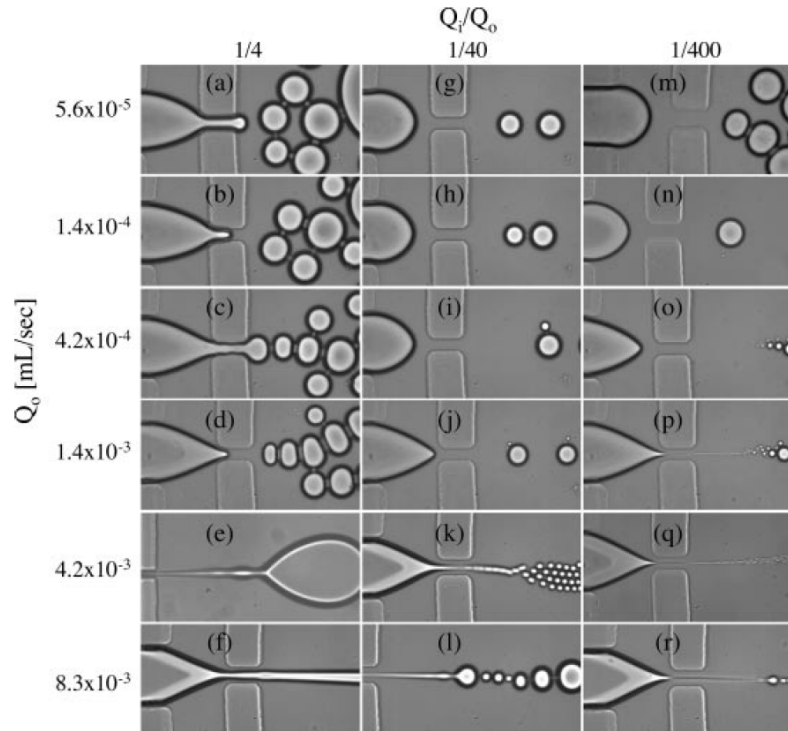


# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial



Drop formation in a flow-focusing configuration

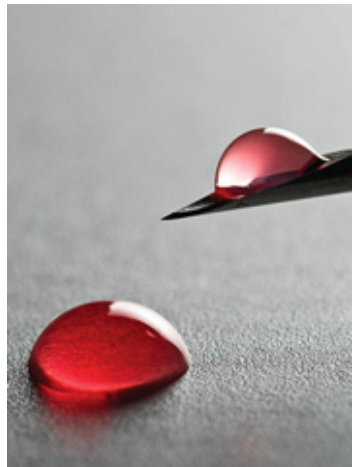
Water drops are formed in a silicone oil continuous phase.

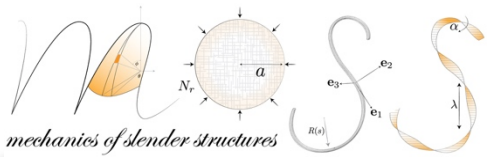
$Q_i$  is the flow rate of water  
 $Q_o$  is the flow rate of oil

Characteristic droplet size:

$$R \sim \frac{\gamma}{\mu U_0} h = \frac{h}{\mathcal{C}}$$

## Interfacial Forces



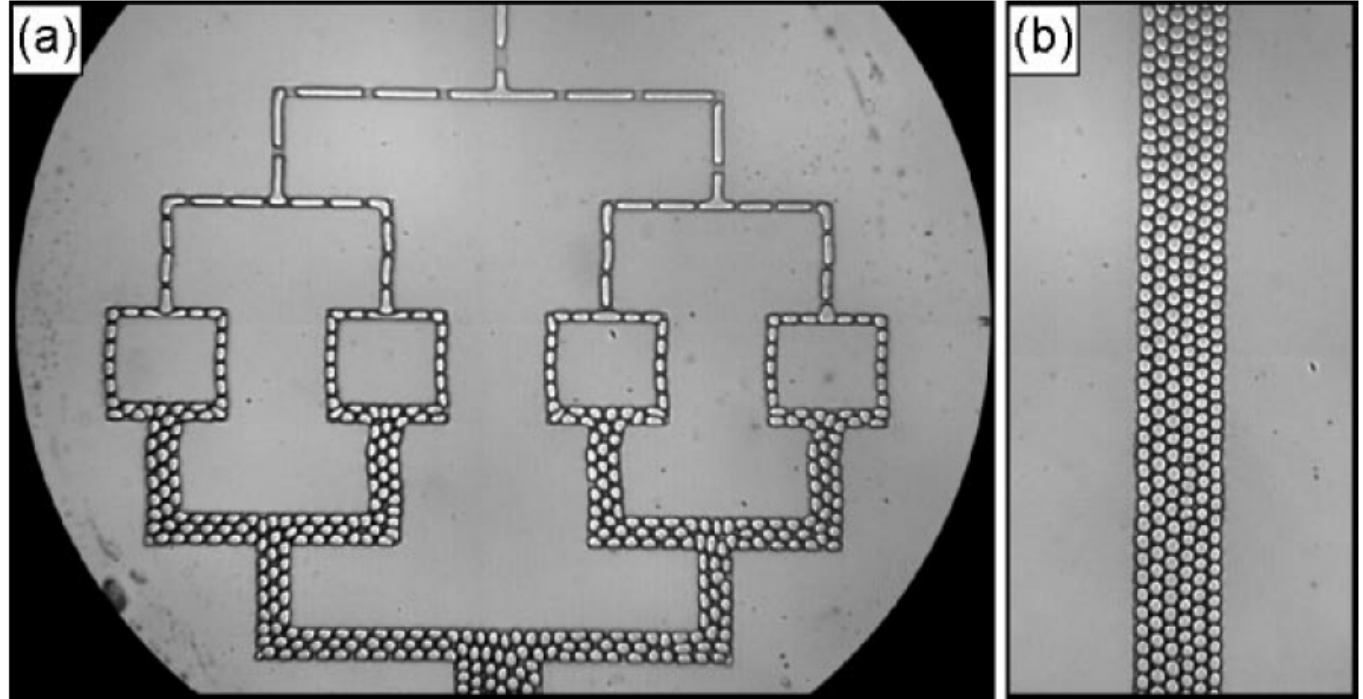


# Fluid Behavior

## Viscous Forces



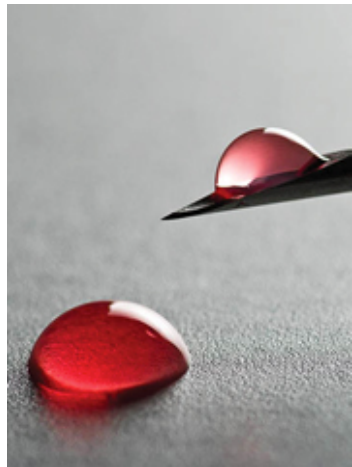
## Capillary Number: viscous/interfacial



## Sequential drop breakup at T-junctions

- Generally, it is difficult to form small drops at high-volume fraction.
- This “passive” route allows size reduction after fabrication.
- The ratio of two daughter droplets depends on the lengths of the arms off the T-junction.

## Interfacial Forces





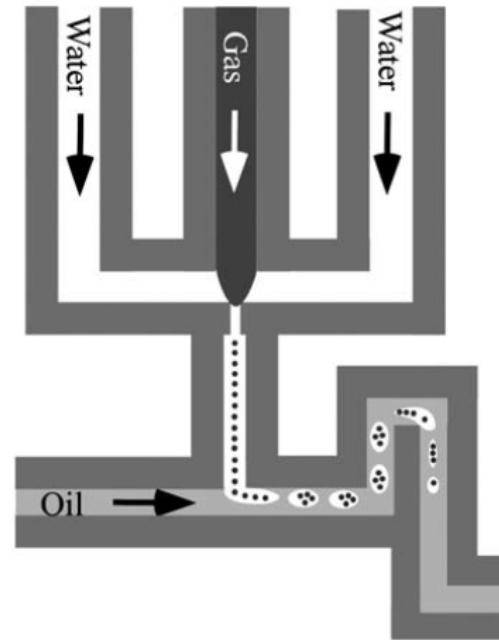


# Fluid Behavior

## Viscous Forces

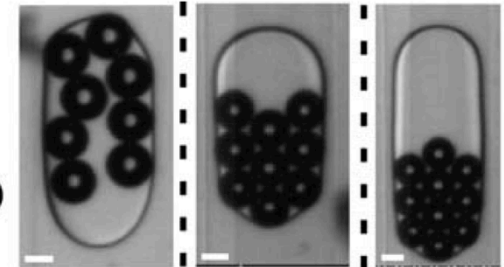


## Capillary Number: viscous/interfacial

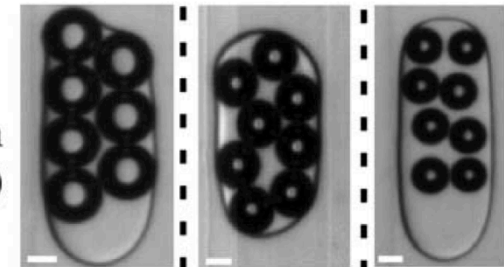


A

Upstream  
( $Z < 5$  mm)

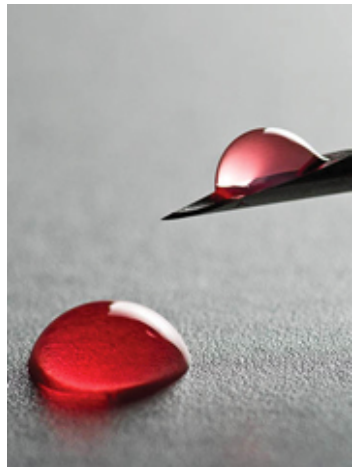


Downstream  
( $Z > 30$  mm)



$Q_w$  ( $\mu\text{l}/\text{min}$ ) = 3 : 9 : 13

## Interfacial Forces





# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

Large **surface-to-volume** ratios in microfluidic devices

- Makes surface effects increasingly important.
- Important when free fluid surfaces are present.

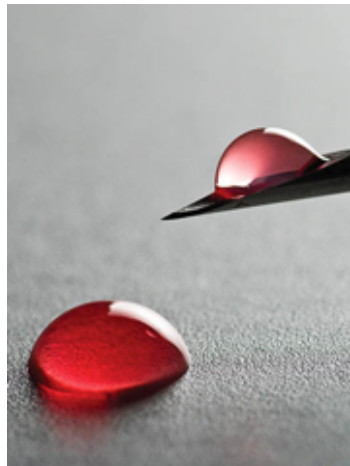
Surface tensions can exert significant stress

- Result in free surface deformations.
- Can drive fluid motion.

Capillary forces tend to draw fluid into wetting microchannels

- Occurs when **solid-liquid** interfacial **energy** is **lower** than the **solid-gas** interfacial **energy**.

## Interfacial Forces





# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

Capillary forces tend to draw fluid into wetting microchannels

**Dynamics** of surface tension-driven intrusion into a pipe of radius  $w$  occurs from **balance** of **capillary** and **viscous forces**.

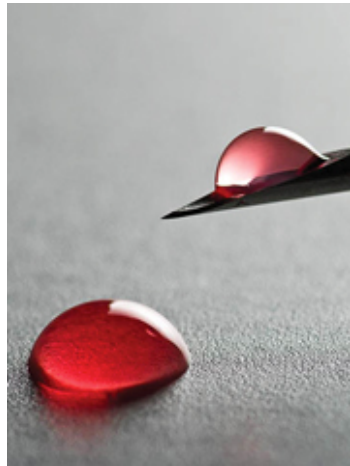
Curved meniscus at fluid-gas interface – Laplace pressure

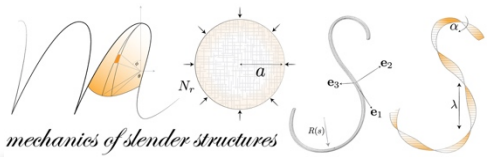
$$\Delta p \sim \Delta \gamma / w$$

Difference in surface energies (solid-liquid to solid-gas)

$$\Delta \gamma = \gamma_{sl} - \gamma_{sg}$$

## Interfacial Forces





# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

Capillary forces tend to draw fluid into wetting microchannels

**Dynamics** of surface tension-driven intrusion into a pipe of radius  $w$  occurs from **balance** of **capillary** and **viscous forces**.

Stoke's flow:

$$\mu \nabla^2 \mathbf{u} = \nabla p$$

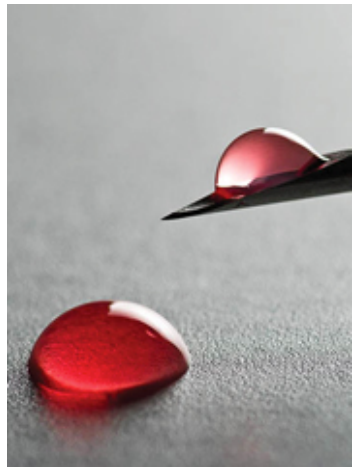
Scaling of the flow rate:

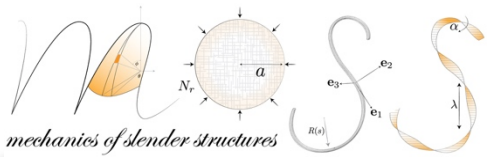
$$u \sim \frac{\Delta P w^2}{\mu L}$$

From the pressure – surface tension relationship:

$$u \sim \frac{\Delta \gamma w}{\mu L}$$

## Interfacial Forces





# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

Capillary forces tend to draw fluid into wetting microchannels

From the pressure – surface tension relationship:

$$u \sim \frac{\Delta\gamma w}{\mu L}$$

Capillary number determines the dynamics:

$$\mathcal{C} \sim w/z$$

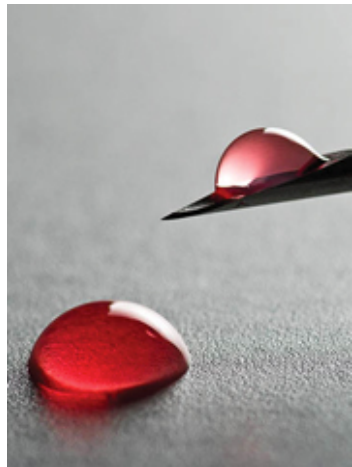
The column length changes as the fluid front moves: ( $u = \partial z / \partial t$ )

- Fluid invades the channel at an ever slowing rate:

$$z \sim \left( \frac{\Delta\gamma w}{\mu} t \right)^{1/2}$$

**Washburn  
equation**

## Interfacial Forces







# Fluid Behavior

## Viscous Forces



## Capillary Number: viscous/interfacial

Surface patterning can create “wall-less” microchannels which confine fluids.

Chemically treat a surface to make some areas hydrophilic and others hydrophobic.

**Spreading** along hydrophilic **stripe** is similar to the **Washburn** analysis, however the height of the fluid stripe:

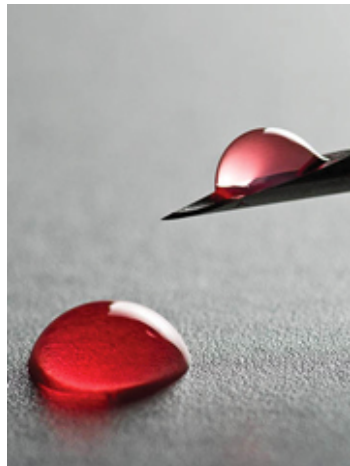
$$h \sim w^2 / R_r$$

- $R_r$  is the radius of curvature of the inlet reservoir drop.

$$z \sim \left( \frac{\Delta\gamma w^4}{\mu R_r^3} t \right)^{1/2}$$

- Spreading requires:  $\mathcal{C} \ll 1$

## Interfacial Forces





# Fluid Behavior

## Viscous Forces



**Capillary Number:** viscous/interfacial

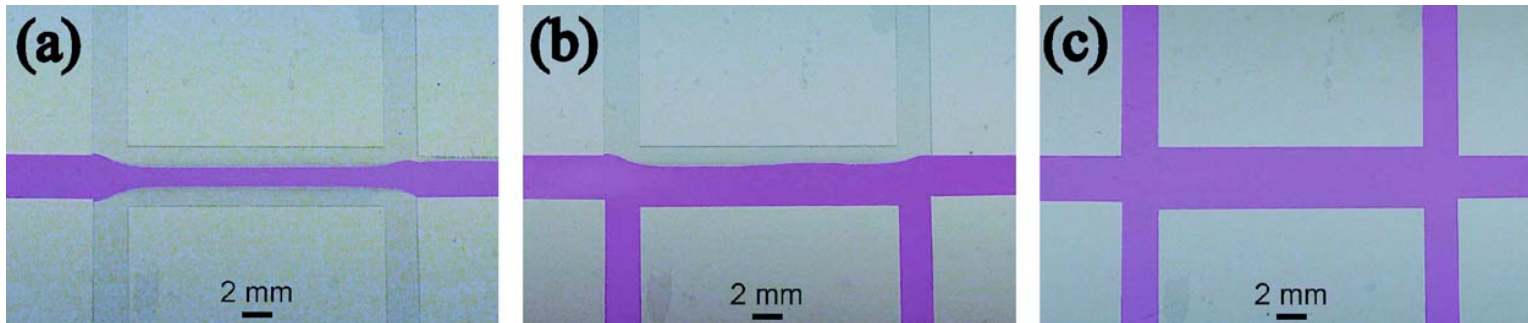
**Chemically treat** walls of **microfluidic channel** to make pressure-sensitive **valves**.

### Pressure sensitive pumping

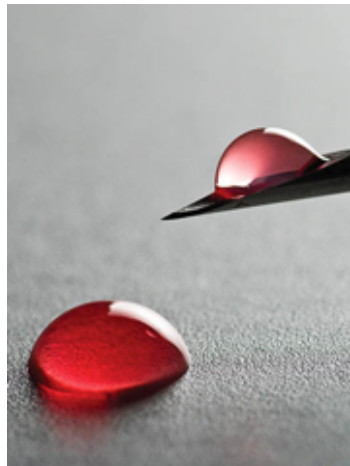
- From pressure drop across fluid channel (different from Quake valve)
- Fluid is confined to hydrophilic region if:

$$\Delta p \leq \gamma/w$$

Channel is patterned with hydrophilic stripe (center), weakly hydrophobic surface (bottom channels), and strongly hydrophobic (top channels).



## Interfacial Forces





# Capillary Rise

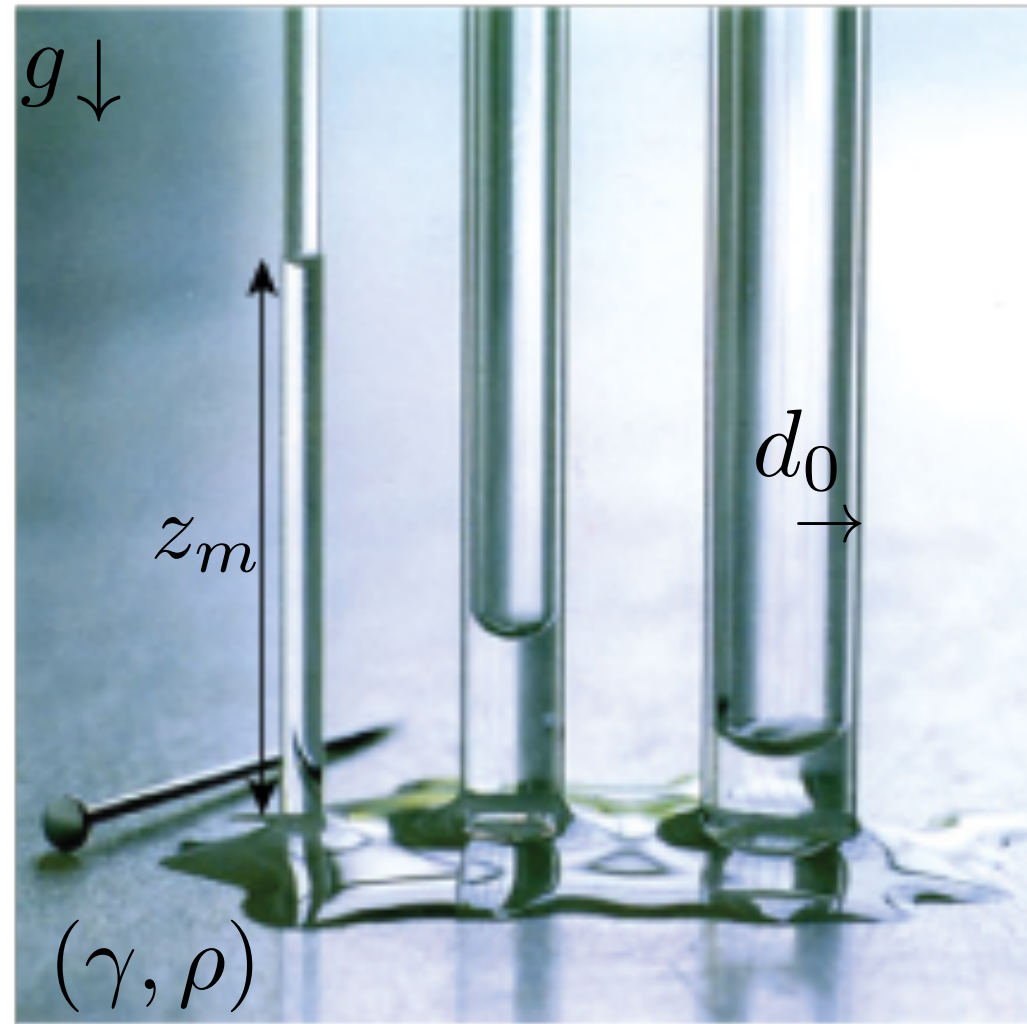
## Classical Problem:

- Noted as early as 15<sup>th</sup> by Leonardo da Vinci.
- Attributed to circulation in plants in 17<sup>th</sup> century by Montanari.

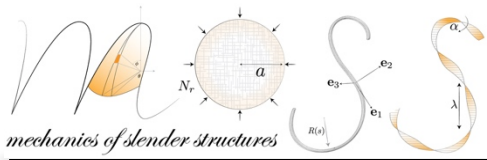
$$U = \underbrace{2\pi\gamma \cos \theta_e r z_m}_{\text{surface energy}} + \underbrace{\frac{1}{2}\rho g \pi r^2 z_m^2}_{\text{gravitational P.E.}}$$

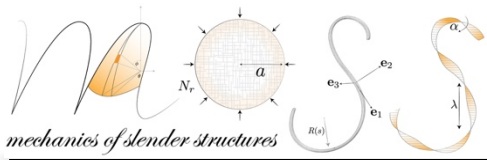
$$\frac{dU}{dz_m} = 0$$

$$l_{cg} \sim \frac{\gamma \cos \theta_e}{\rho g d_0}$$



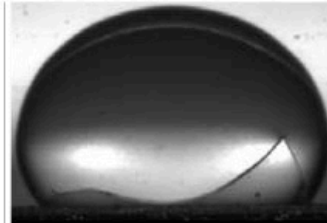
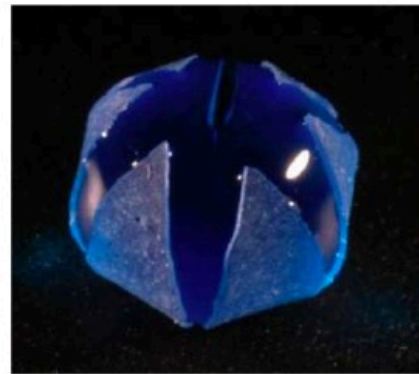
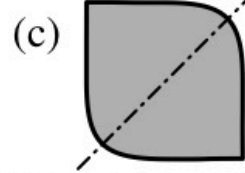
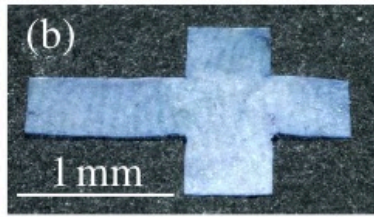
Balance: Surface Tension & Gravity



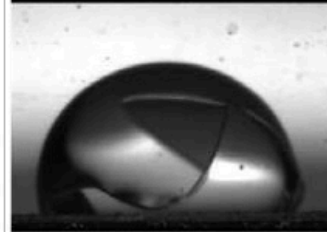
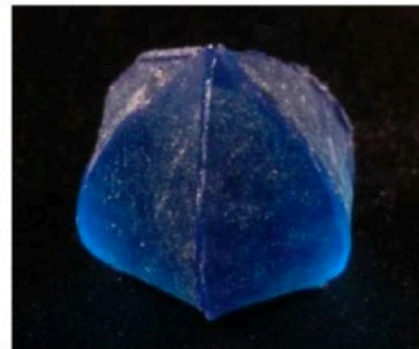




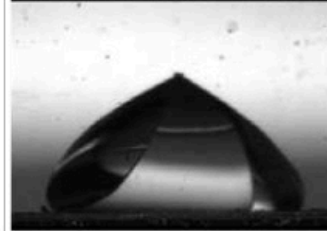
# Elastocapillarity



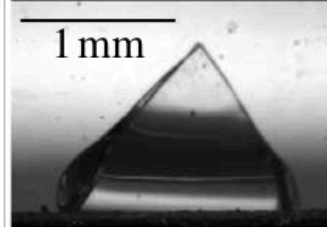
Fluid-structure interaction:



- Droplet bends and folds the sheet.



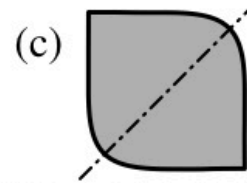
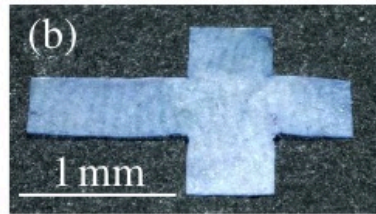
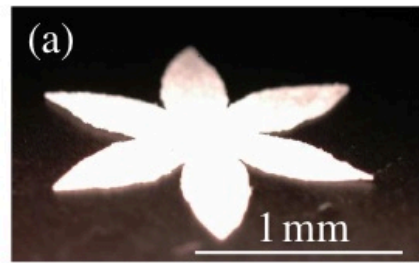
- Droplet is minimizing the amount of its surface in contact with air.



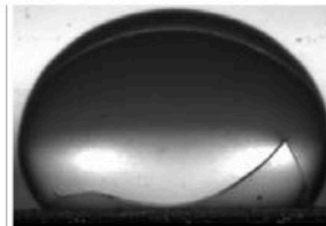
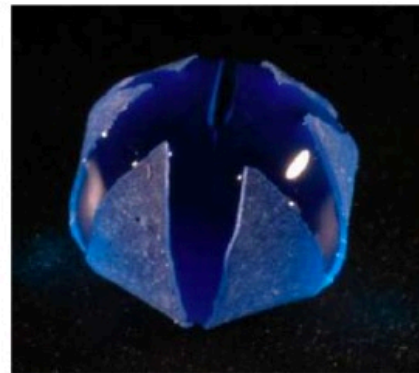
- Liquid-air surface area is minimized at the expense of bending the sheet.



# Elastocapillarity



Fluid-structure interaction:

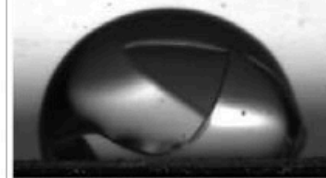


Elastic energy of a plate – bending:

$$\mathcal{U}_e = \frac{1}{2} \iint_P dx dy \int_{-h/2}^{h/2} dz (\sigma_{\alpha\beta} \varepsilon_{\alpha\beta})$$

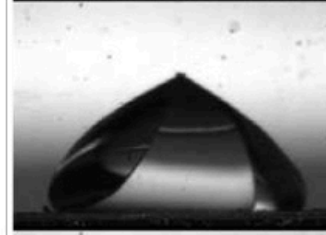
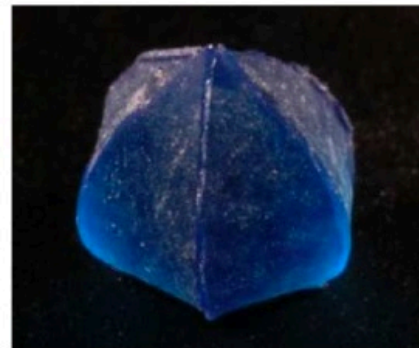
Relation between in-plane strain to out-of-plane bending:

$$\varepsilon_{\alpha\beta}(x) = z \frac{d^2 w}{dx^2} = \frac{z}{R}$$

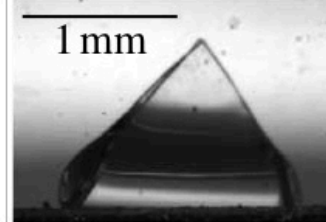


Bending energy:

$$\mathcal{U}_b = \frac{1}{2} \iint_P dx dy \frac{Eh^3}{12} \left( \frac{1}{R} \right)^2$$

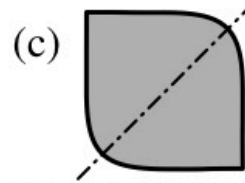
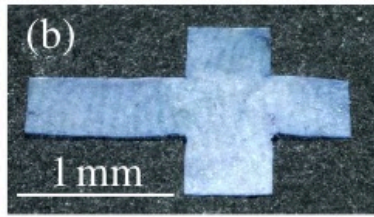
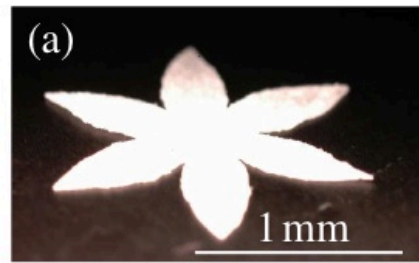


1 mm



$$\mathcal{U}_b \sim Eh^3$$

# Elastocapillarity



Fluid-structure interaction:

Bending energy:

$$U_b \sim Eh^3$$

Surface energy:

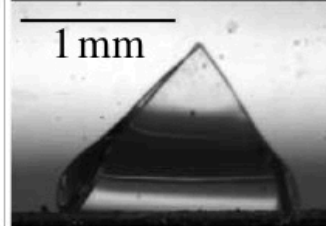
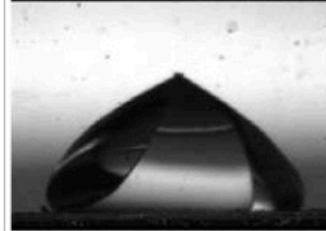
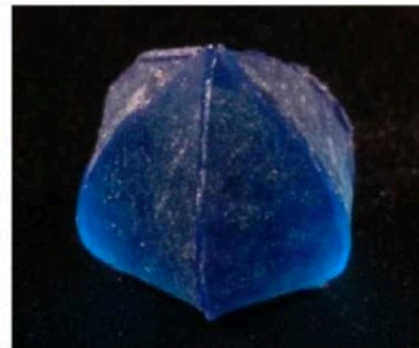
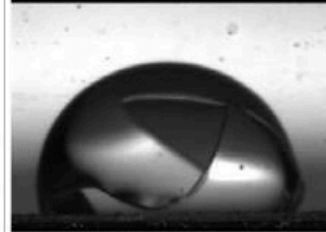
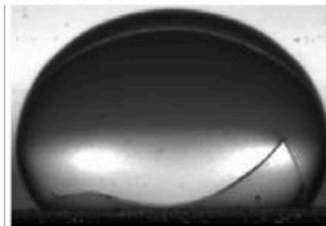
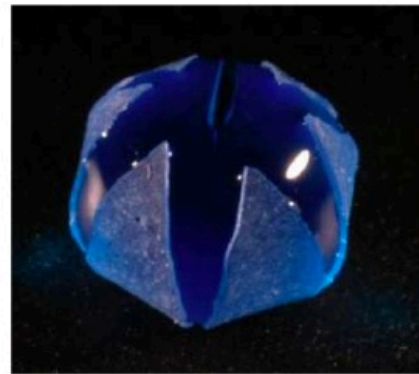
$$U_\gamma \sim \gamma L^2$$

Elastocapillary length:

$$\ell_{ec} \sim \sqrt{\frac{Eh^3}{\gamma}} \sim \sqrt{\frac{B}{\gamma}}$$

Elastocapillary bending of sheet:

$$7\ell_{ec} \leq L \leq 12\ell_{ec}$$







# Elastocapillarity

Capillary rise between flexible fibers.

$$\underbrace{B \frac{\partial^4 d}{\partial x^4}}_{\text{Beam bending}} = \underbrace{\gamma \kappa_f}_{\text{Laplace pressure}}$$

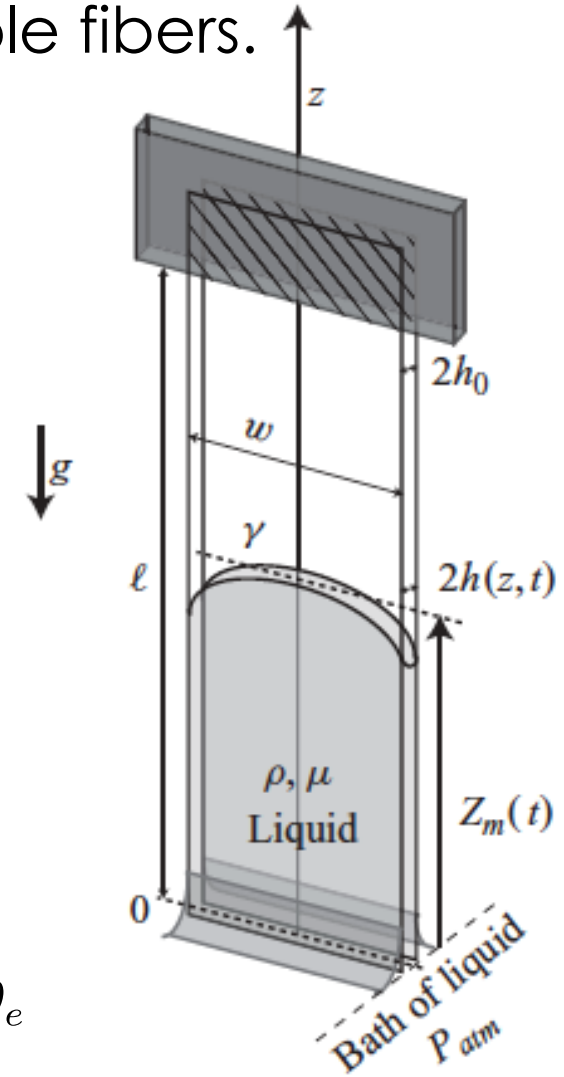
Curvature of meniscus

$$\kappa_f = \frac{1}{d(z_m)} \cos \theta_e + \frac{2}{b} \cos \theta_e$$

Assume gap is much smaller than width:  $d_0 \ll b$

Initial gap:  $d(z_m, t) = d_0$

Approximation of meniscus curvature:  $\kappa_f \approx \frac{1}{d_0} \cos \theta_e$



J. Bico et al (2004)



# Elastocapillarity

Capillary rise between flexible fibers.

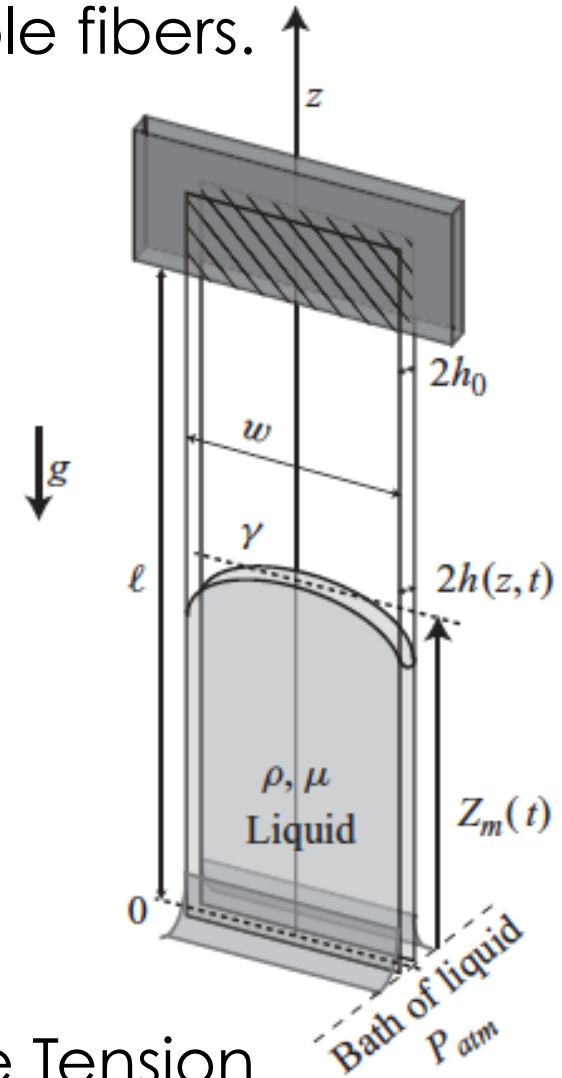
$$\underbrace{B \frac{\partial^4 d}{\partial x^4}}_{\text{Beam bending}} = \underbrace{\gamma \kappa_f}_{\text{Laplace pressure}}$$

Approximation:  $\kappa_f \approx \frac{1}{d_0} \cos \theta_e$

Scaling:  $B \frac{d_0}{\ell^4} \sim \frac{\gamma}{d_0} \cos \theta_e$

$$\ell_{ec} \equiv \left( \frac{B d_0^2}{\gamma \cos \theta_e} \right)^{1/4}$$

Balance: Bending & Surface Tension



J. Bico et al (2004)

H.-Y. Kim and L. Mahadevan, "Capillary rise between elastic sheets," *J. Fluid Mech.* **548**, 141-150, (2006).  
 J.M. Aristoff, C. Duprat, and H.A. Stone, "Elastocapillary Imbibition," *Int. J Nonlinear Mech.* **48**, 648-656, (2011).  
 C. Duprat, J.M. Aristoff, and H.A. Stone, "Dynamics of elastocapillary rise," *J. Fluid Mech.* **679**, 641-654, (2011).







# Elastocapillarity

Capillary rise between flexible fibers.

**Potential energies:**

Elastic energy

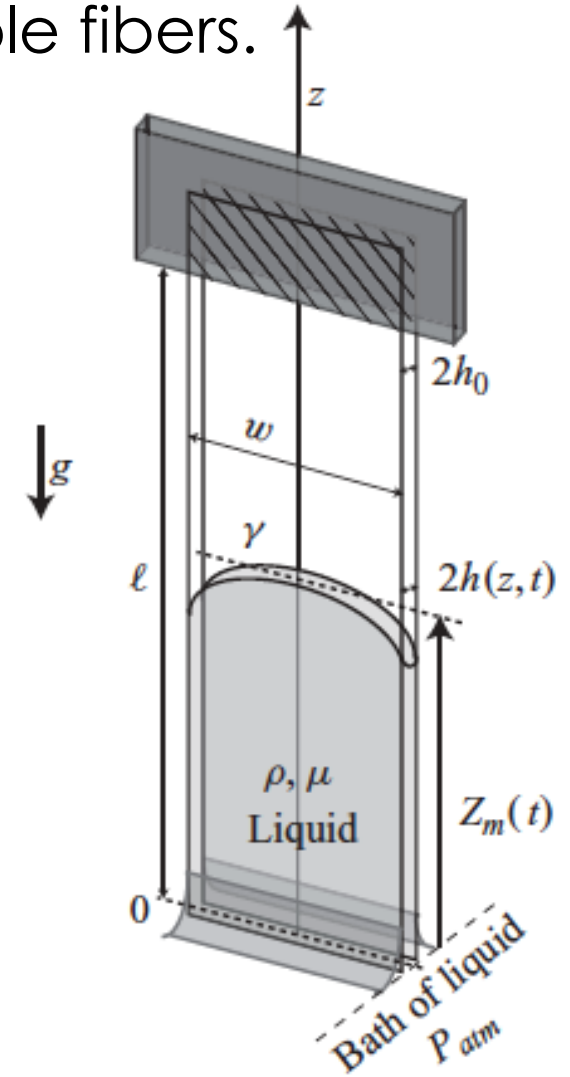
$$U_e \sim \frac{Bwh_0^2}{\ell^3}$$

Gravitational potential energy

$$U_g \sim \rho g w h_0 \ell^2$$

Surface energy

$$U_c \sim w \ell \gamma$$





# Elastocapillarity

Capillary rise between flexible fibers.

**Characteristic length scales:**

Elastocapillary length

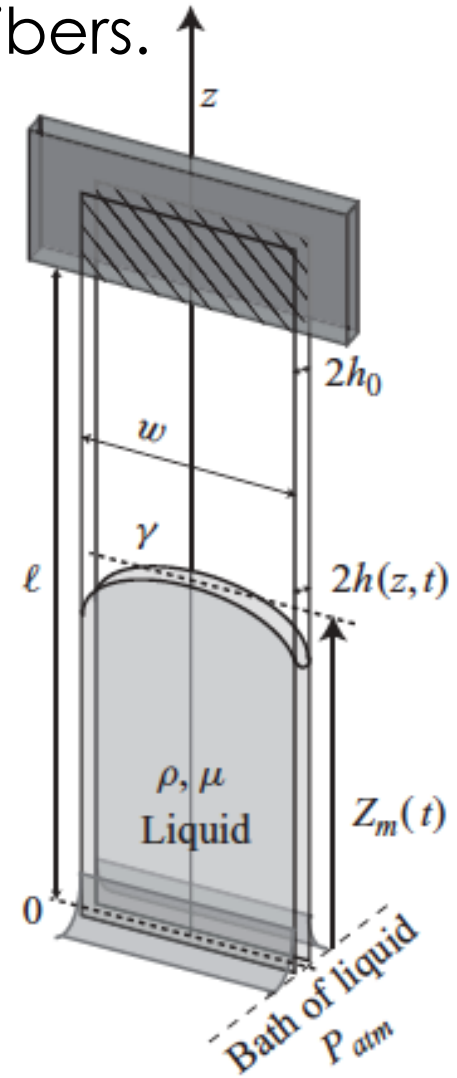
$$l_{ec} = \frac{U_e}{U_c} = \left( \frac{Bh_0^2}{\gamma} \right)^{1/4}$$

Capillary gravity length

$$l_{cg} = \frac{U_c}{U_g} = \frac{\gamma}{\rho gh_0}$$

Elastogravity length

$$l_{eg} = \frac{U_e}{U_g} = \left( \frac{Bh_0}{\rho g} \right)^{1/5}$$





# Elastocapillarity

Capillary rise between flexible fibers.

## Dimensionless parameters

Bond number:

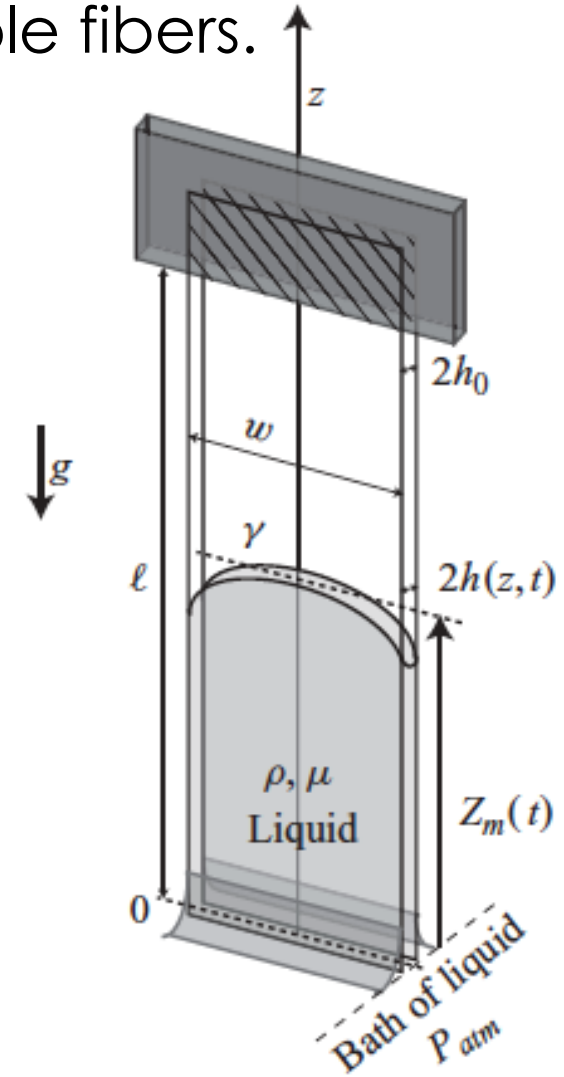
$$\mathcal{B} = \frac{l}{l_{cg}}$$

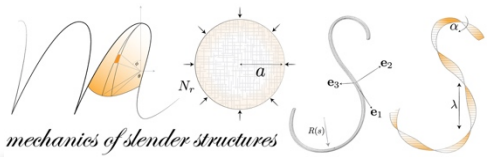
Elastocapillary number

$$\mathcal{E} = \left( \frac{l}{l_{ec}} \right)^4$$

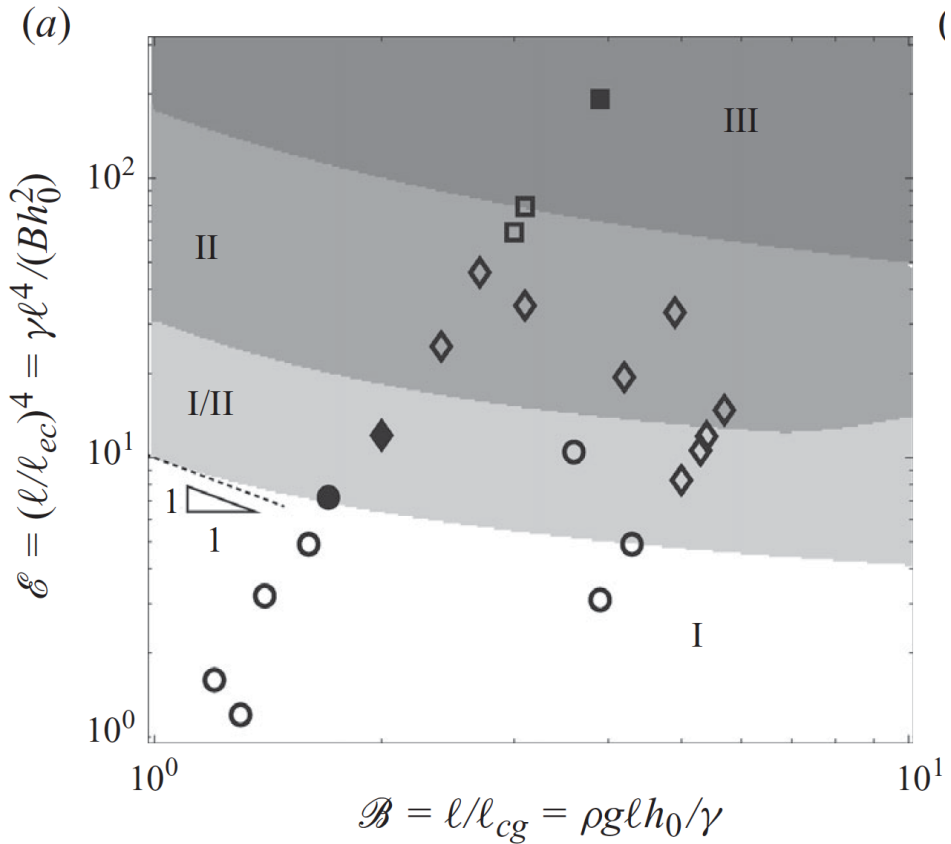
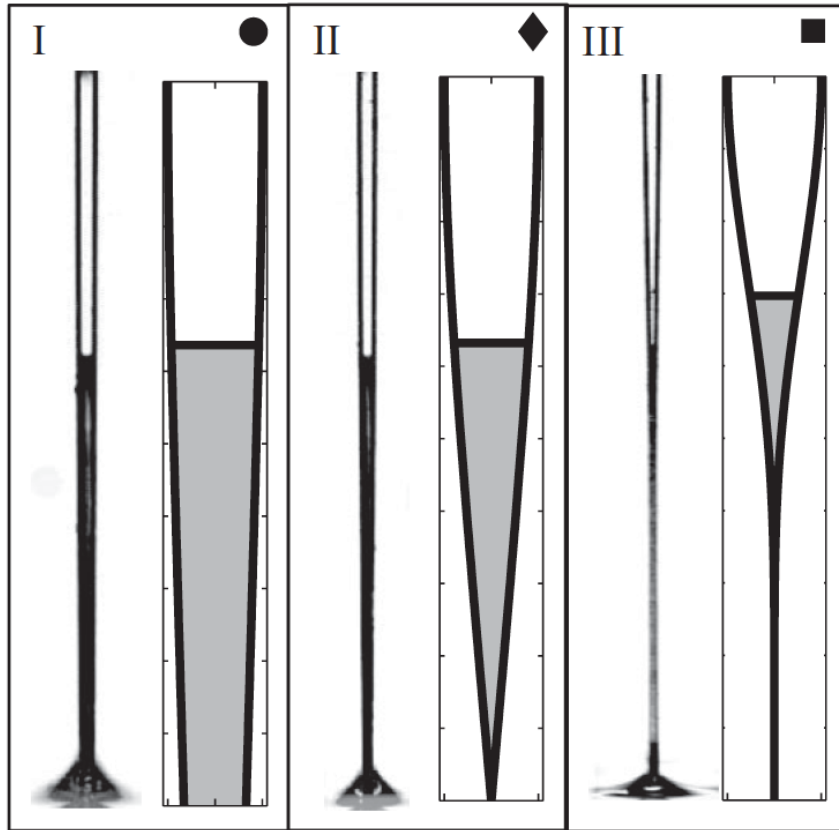
Elastogravity number

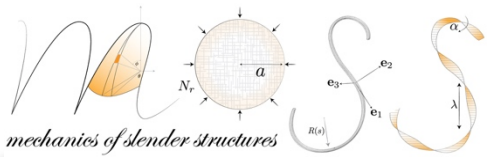
$$\mathcal{G} = \mathcal{B}\mathcal{E} = \left( \frac{l}{l_{eg}} \right)^5$$



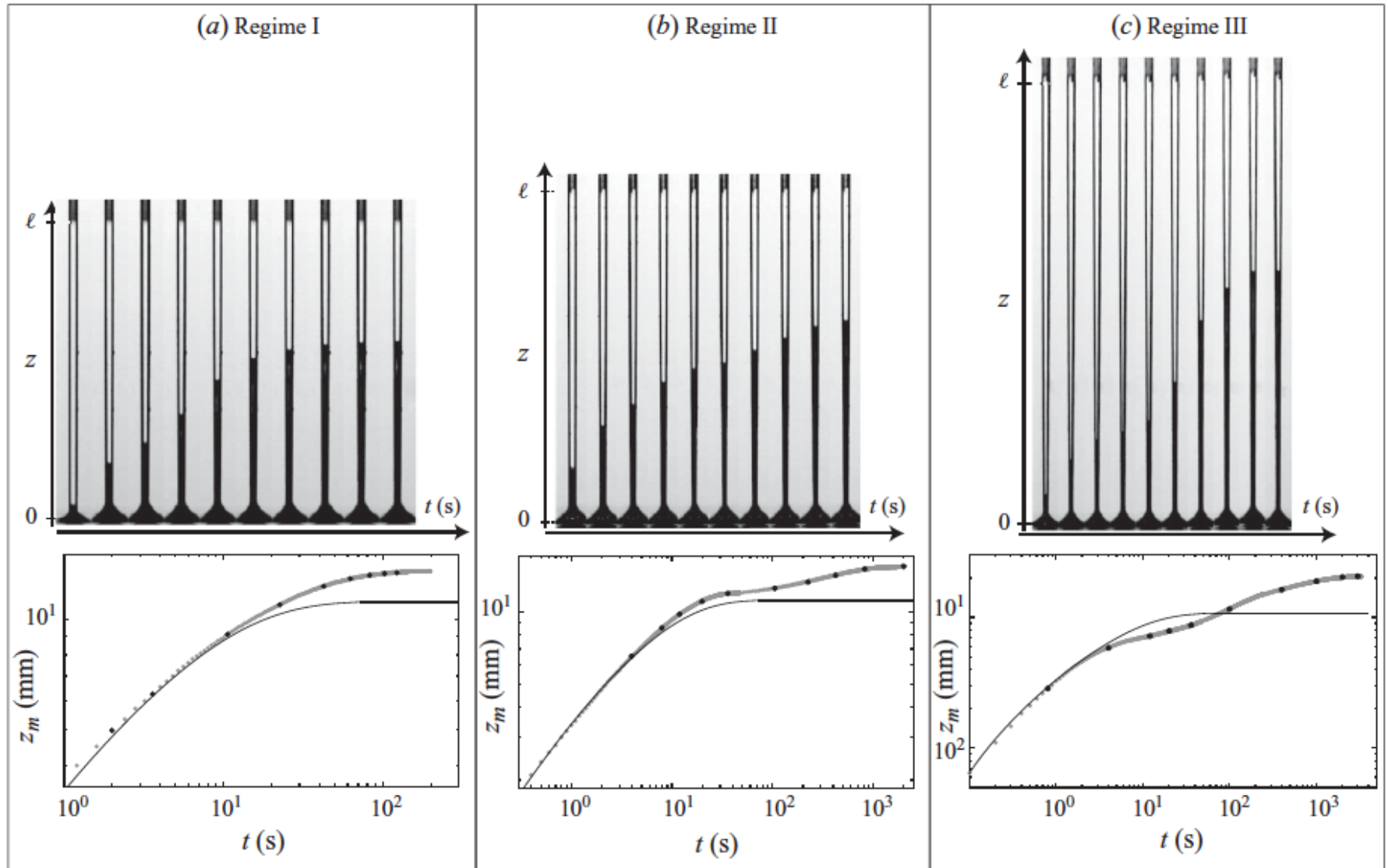


# Elastocapillarity





# Elastocapillarity

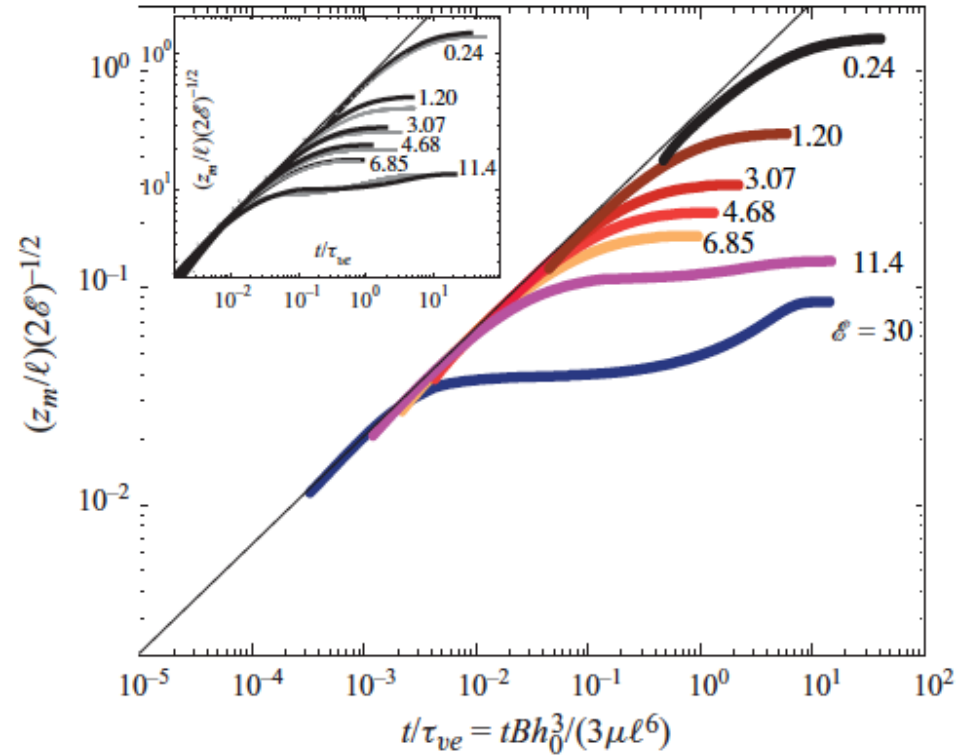
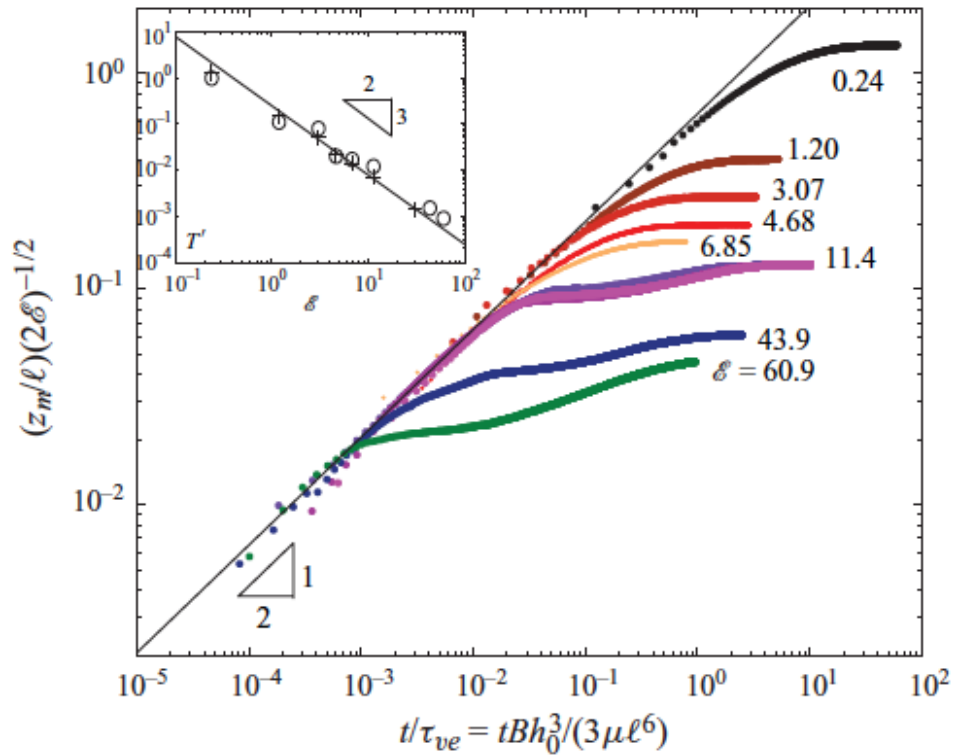


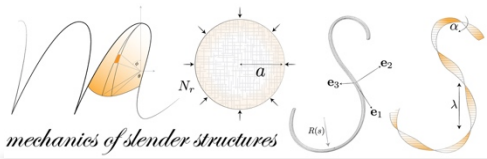
H.-Y. Kim and L. Mahadevan, "Capillary rise between elastic sheets," *J. Fluid Mech.* **548**, 141-150, (2006).  
 J.M. Aristoff, C. Duprat, and H.A. Stone, "Elastocapillary Imbibition," *Int. J. Nonlinear Mech.* **48**, 648-656, (2011).  
 C. Duprat, J.M. Aristoff, and H.A. Stone, "Dynamics of elastocapillary rise," *J. Fluid Mech.* **679**, 641-654, (2011).



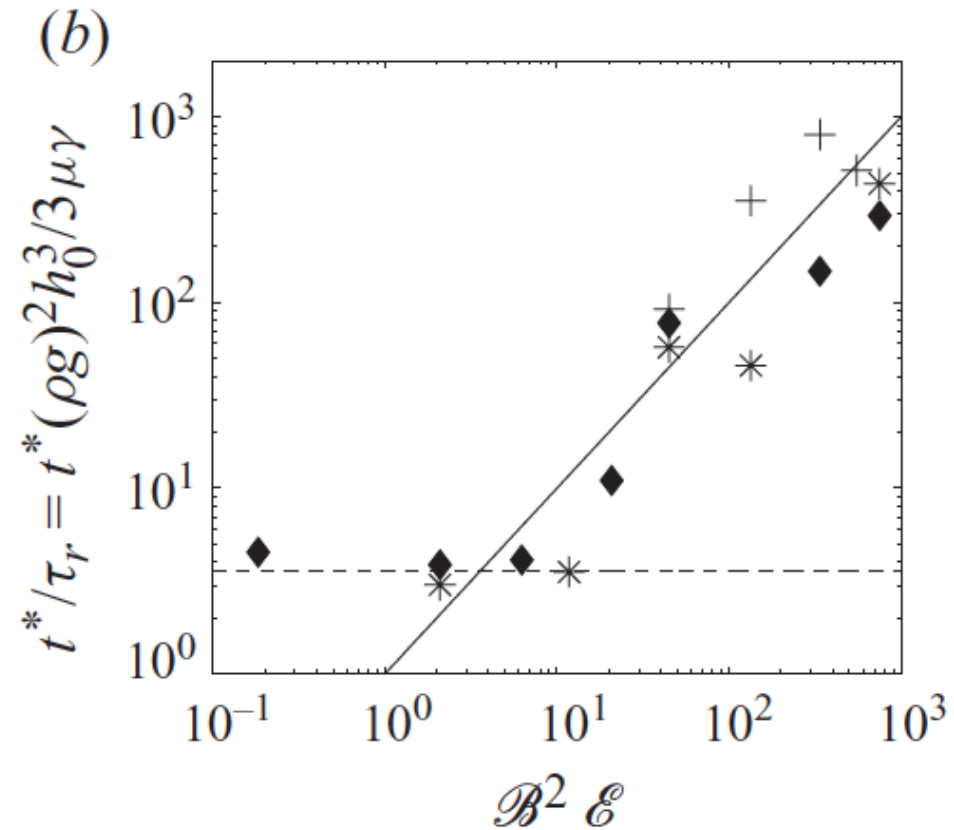
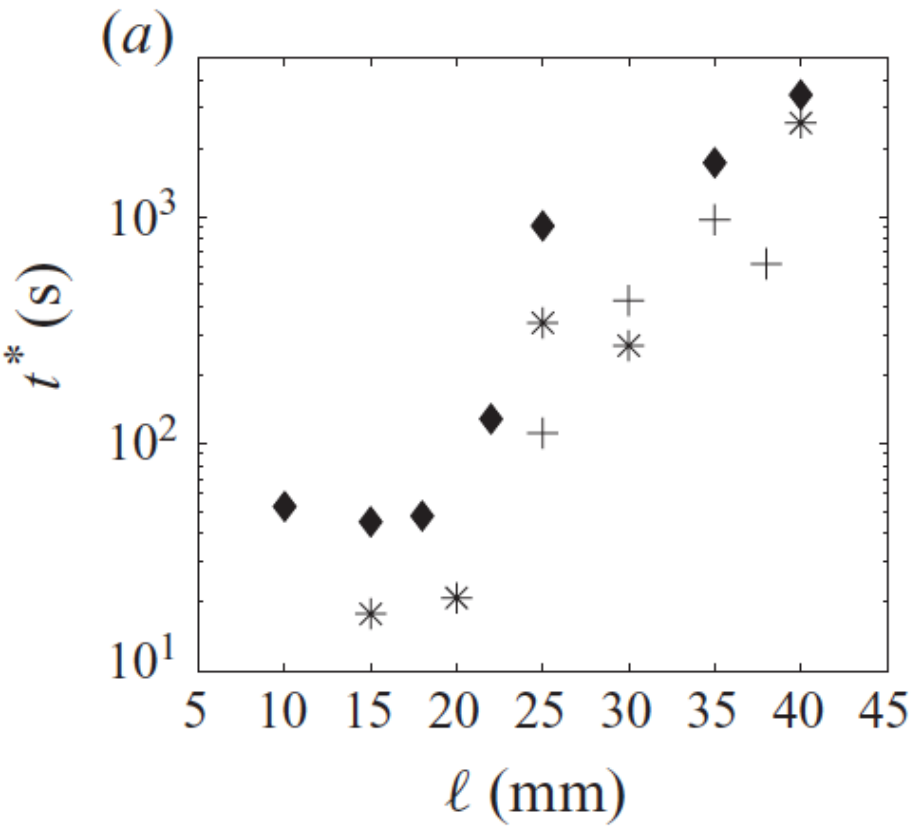


# Elastocapillarity





# Elastocapillarity





# Capillarity & Swelling



Solid: Polyvinylsiloxane

Fluid: Silicone Oil (5 cSt)

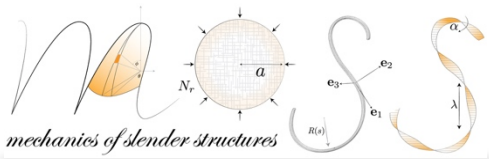
20x faster than real time

$E \approx 1 \text{ MPa}$  (PVS)

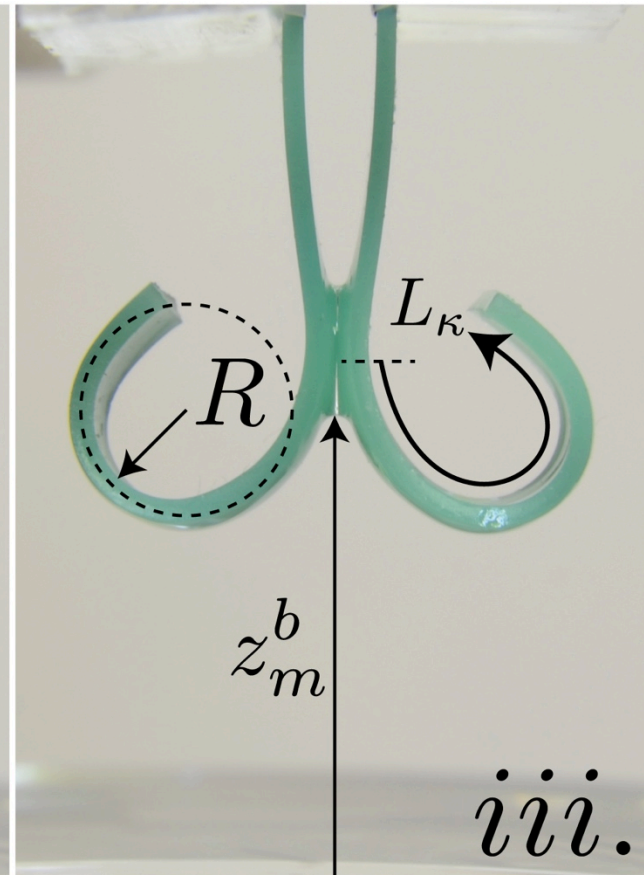
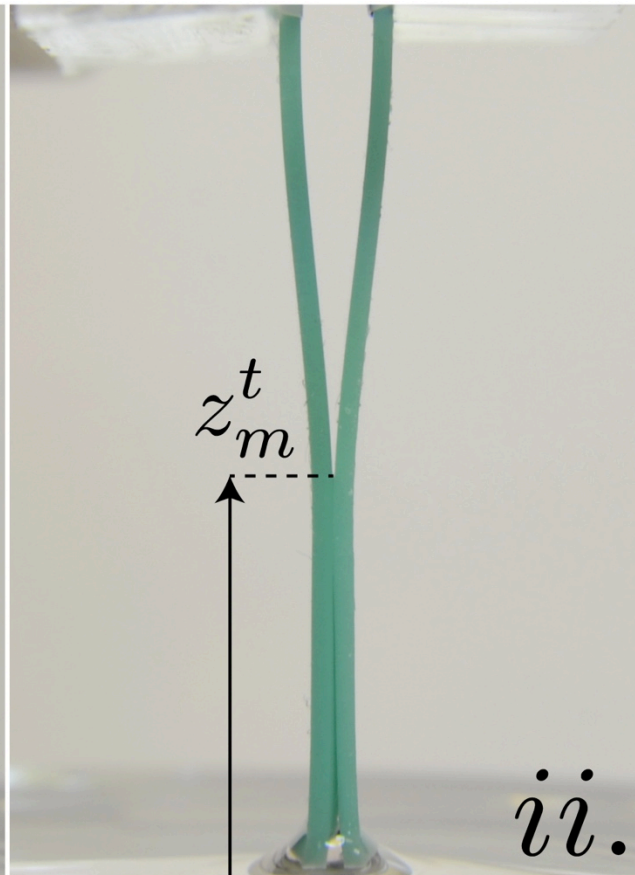
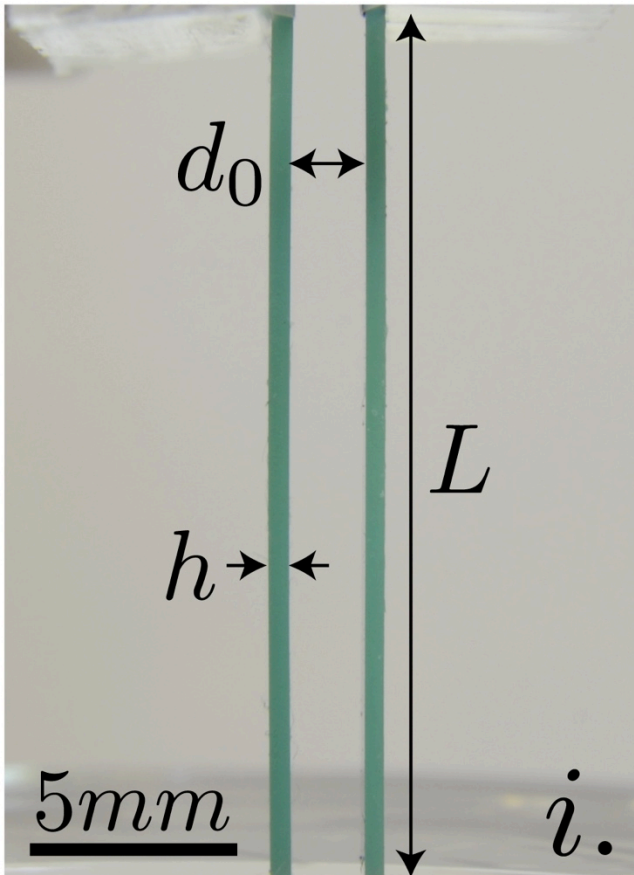
$L = 20 \text{ mm}$

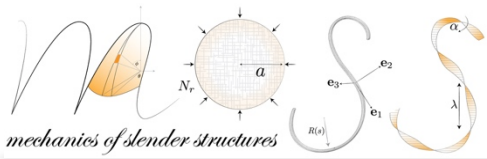
$d \approx 2 \text{ mm}$

$h \approx 0.5 \text{ mm}$

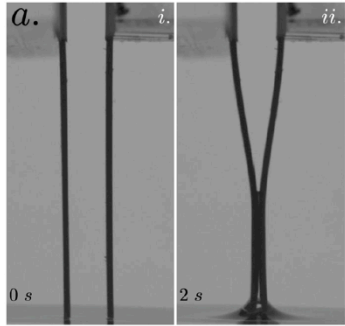


# Capillarity & Swelling

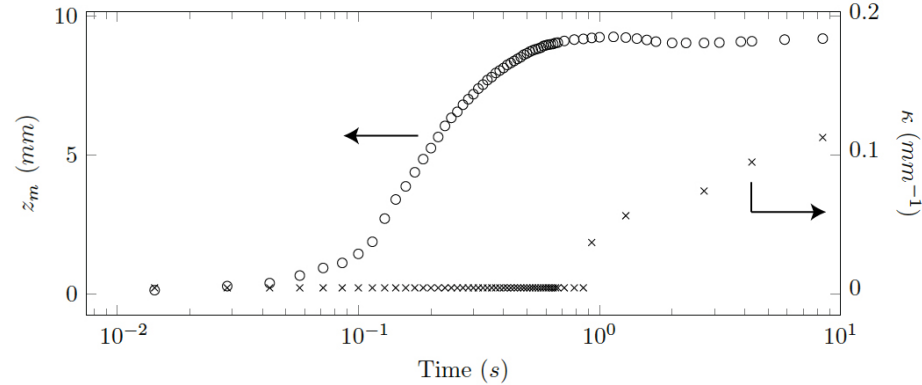
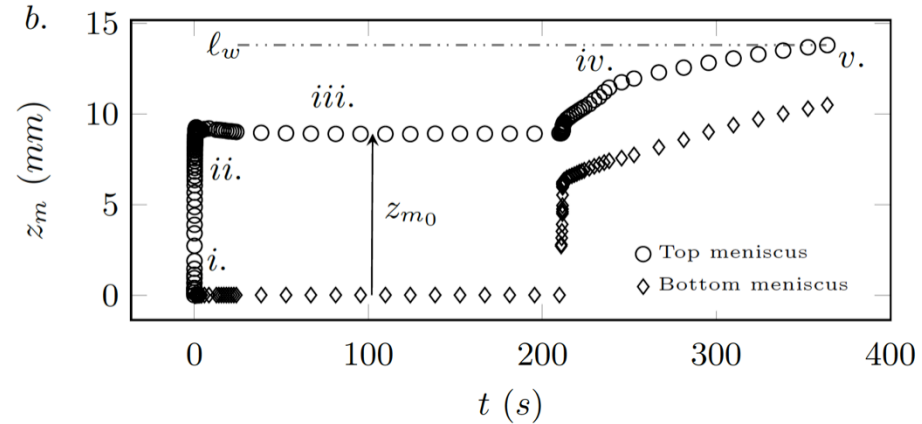
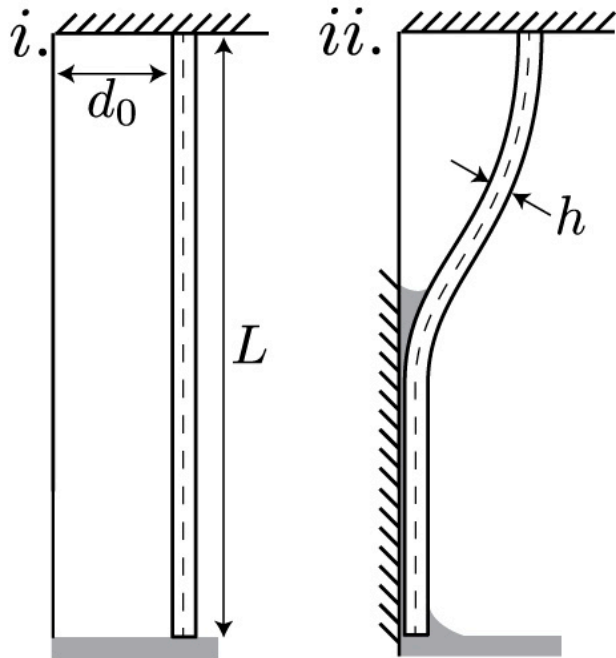




# Capillarity & Swelling



1. Elastocapillary rise between flexible fibers.



$$B \frac{\partial^4 d}{\partial x^4} = \gamma \kappa_f$$

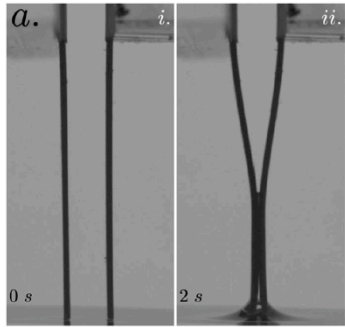
$$B \frac{d_0}{\ell^4} \sim \frac{\gamma}{d_0} \cos \theta_e$$

$$\ell_{ec} \equiv \left( \frac{B d_0^2}{\gamma \cos \theta_e} \right)^{1/4}$$

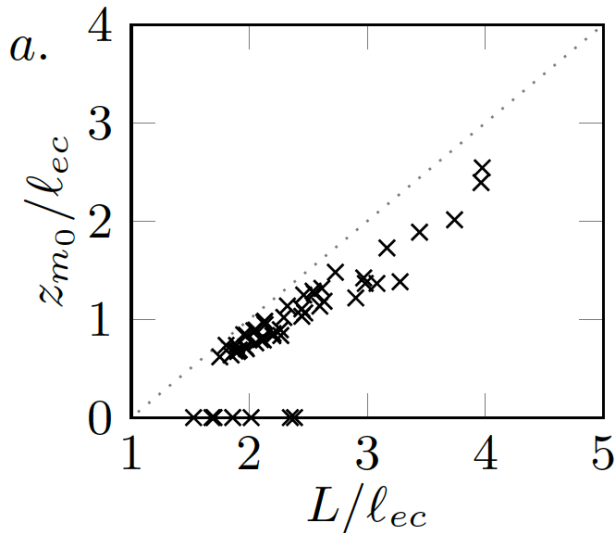




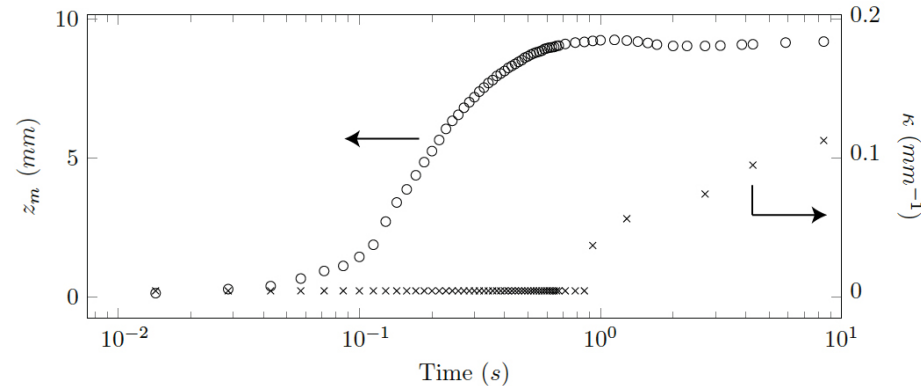
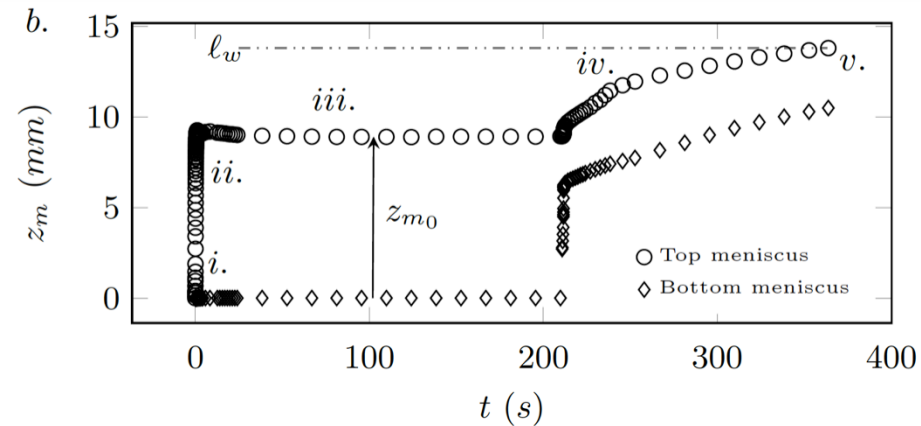
# Capillarity & Swelling



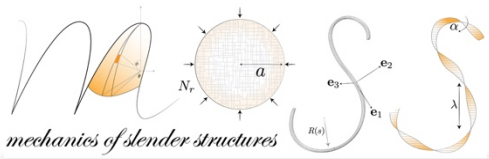
1. Elastocapillary rise between flexible fibers.



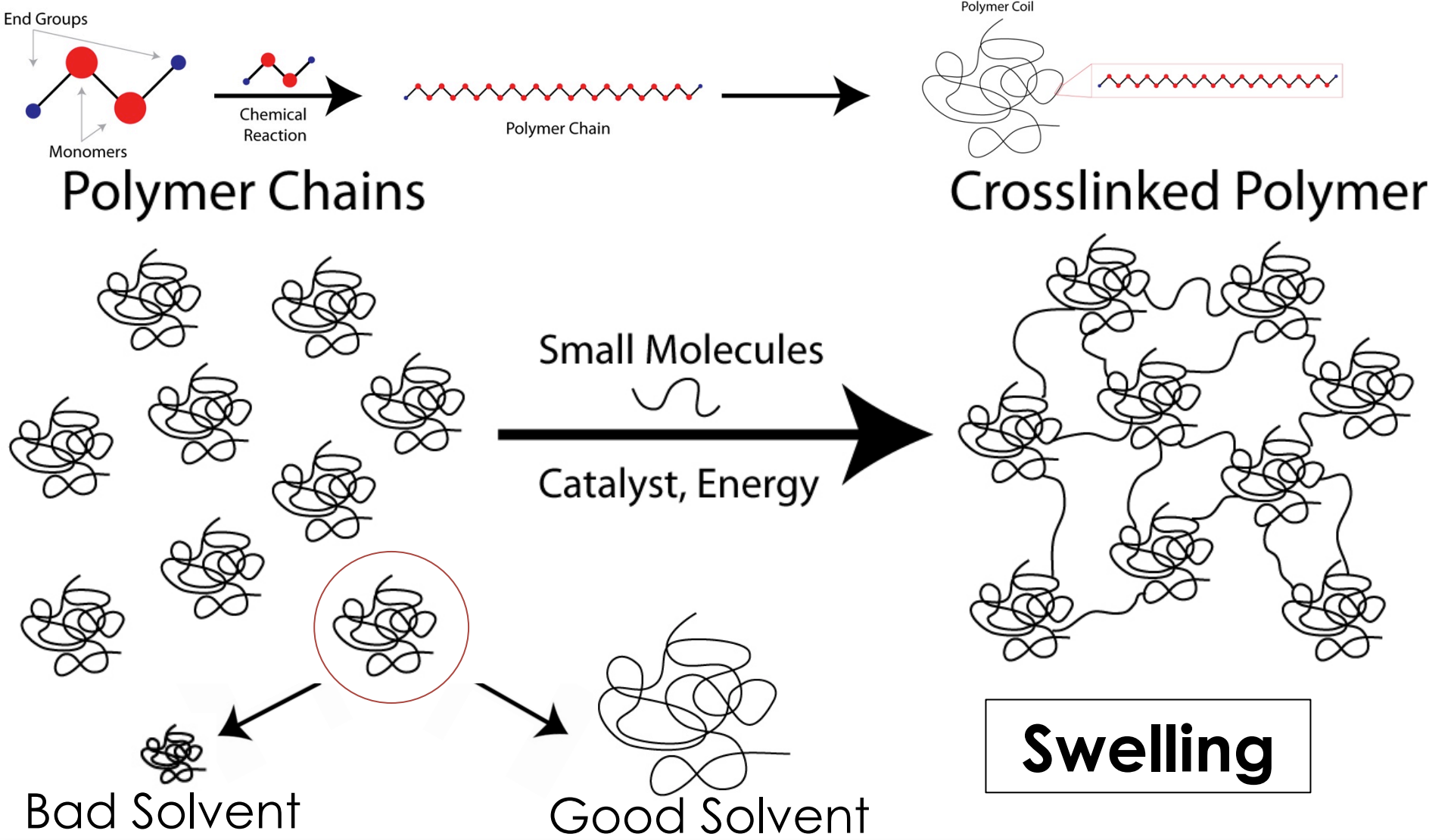
Stationary meniscus height rises linearly with elastocapillary length.



$$\ell_{ec} \sim \left( \frac{Bd_0}{\gamma} \right)^{1/4}$$

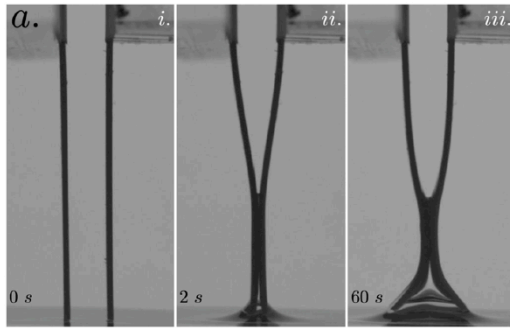


# Polymers & Swelling

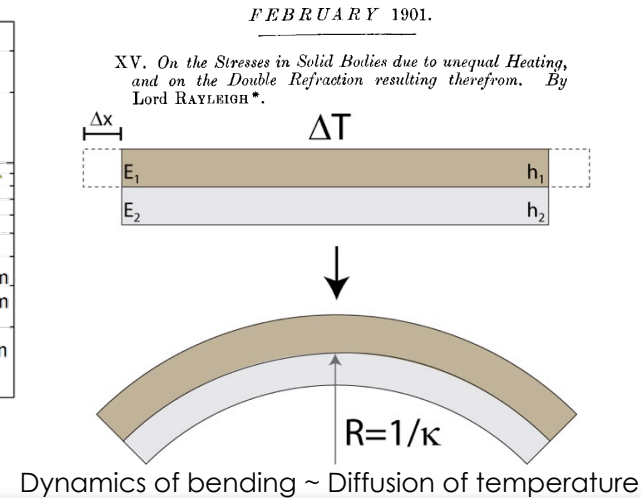
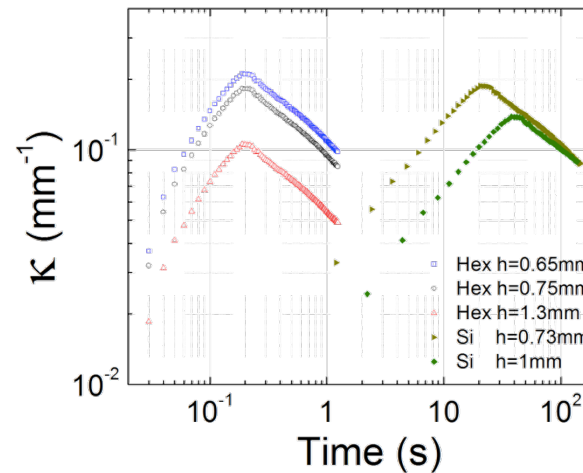
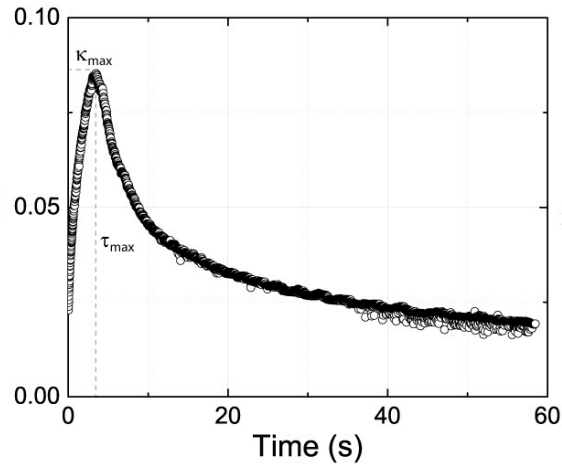
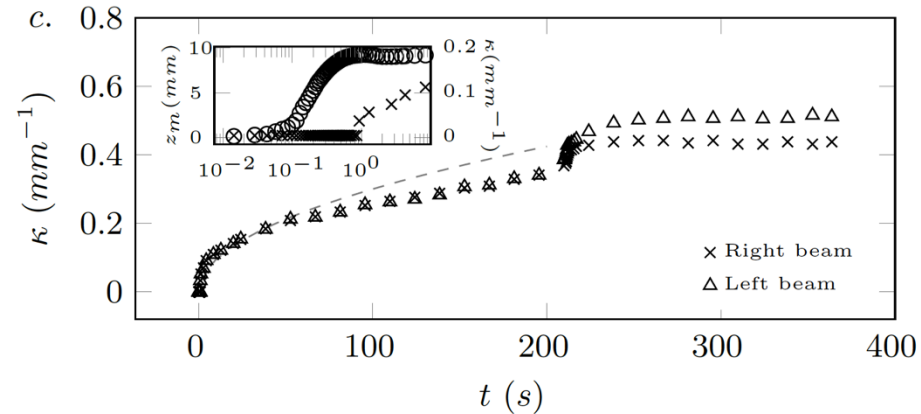
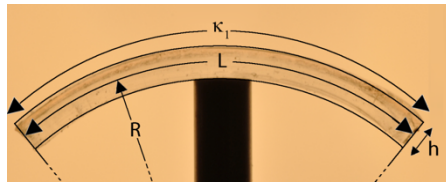




# Capillarity & Swelling

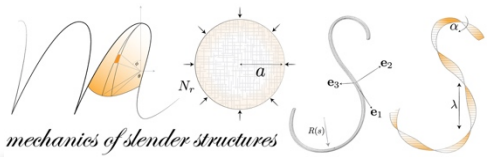


## 2. Swelling-induced bending.

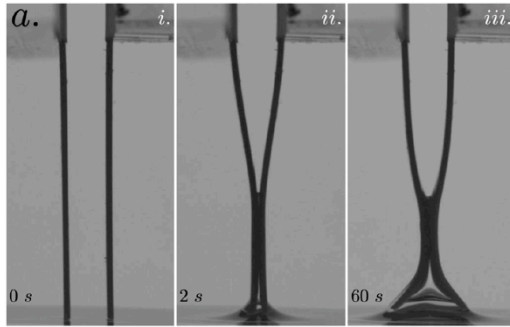


D.P. Holmes, A. Pandey, and S. Protière, *In Preparation*, (2015).

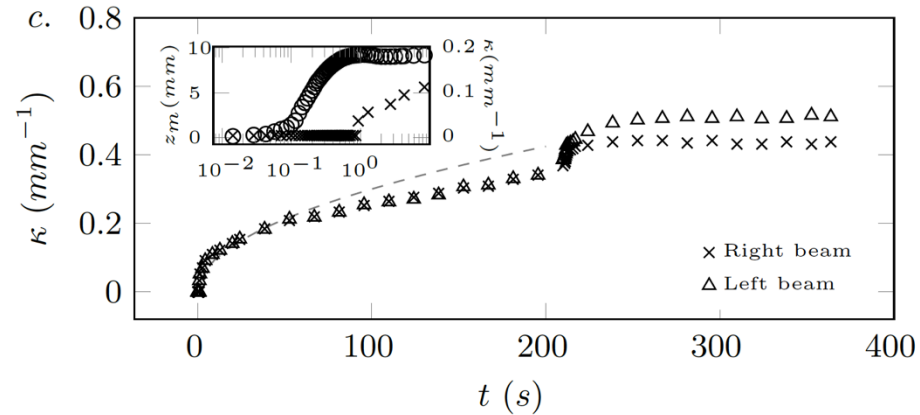
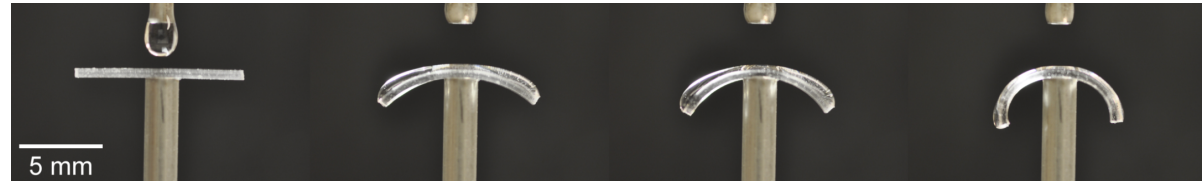
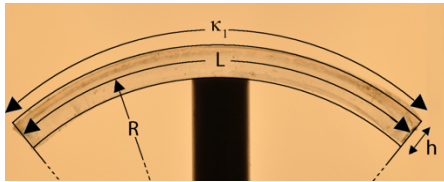
D.P. Holmes, M. Roché, T. Sinha, and H.A. Stone. "Bending and Twisting of Soft Materials by Non-homogenous Swelling" *Soft Matter*, **7**, 5188, 2011.



# Capillarity & Swelling



## 2. Swelling-induced bending.



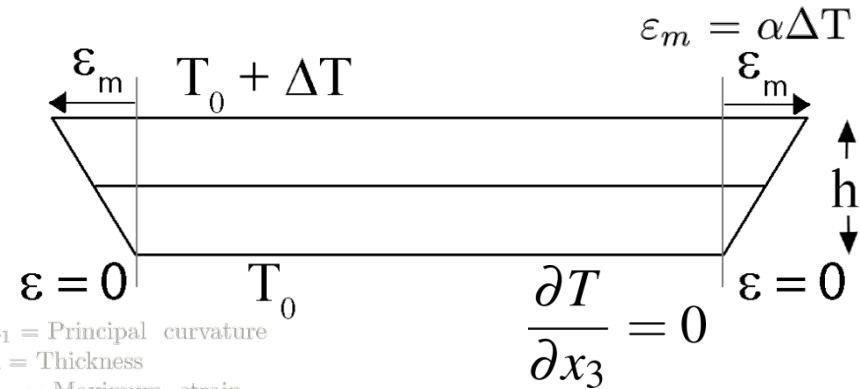
- Thermal diffusion through the beam thickness.
- Shape obtained by minimizing the bending moment in the beam.
- Beam curvature as temperature diffuses.

$$\frac{\kappa_1 h}{\epsilon_m(1+\nu)} = 1.33e^{-\frac{\pi^2 t/\tau}{4}} - 0.77e^{-\frac{9\pi^2 t/\tau}{4}} + \dots$$

- Poroelastic time scale

$$\tau_p \approx \frac{\mu h^2}{kE}$$

$\mu$  = Solvent viscosity  
 $h$  = Thickness  
 $k$  = Permeability ( $k \approx 10^{-18} \text{ m}^2/\text{s}$ )  
 $E$  = Elastic modulus ( $E = 10^6 \text{ Pa}$ )

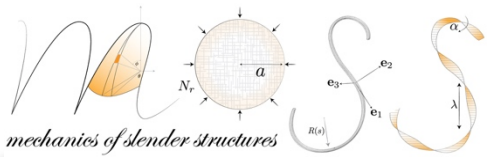


$\kappa_1$  = Principal curvature  
 $h$  = Thickness  
 $\epsilon_m$  = Maximum strain  
 $\nu$  = Poisson's ratio

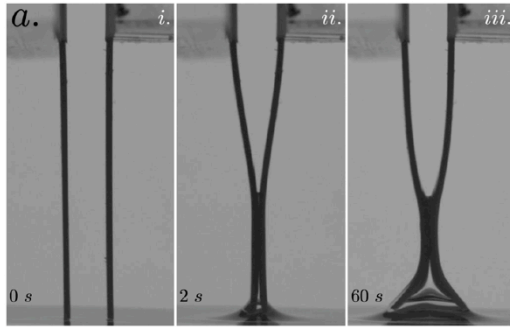
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D.P. Holmes, M. Roché, T. Sinha, and H.A. Stone. "Bending and Twisting of Soft Materials by Non-homogenous Swelling" *Soft Matter*, **7**, 5188, 2011.

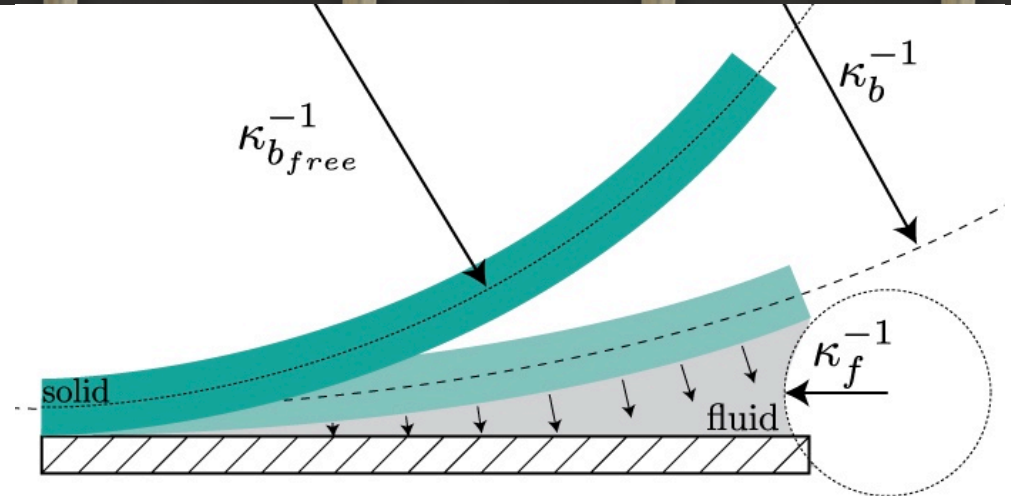
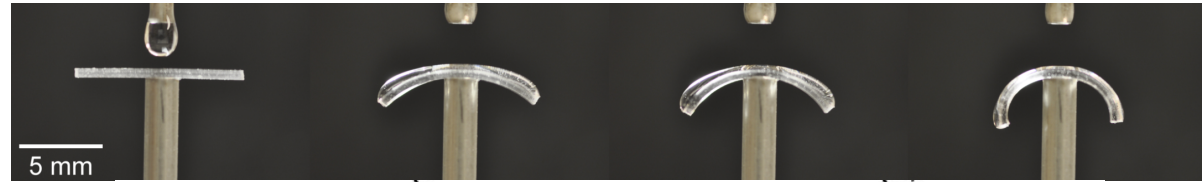
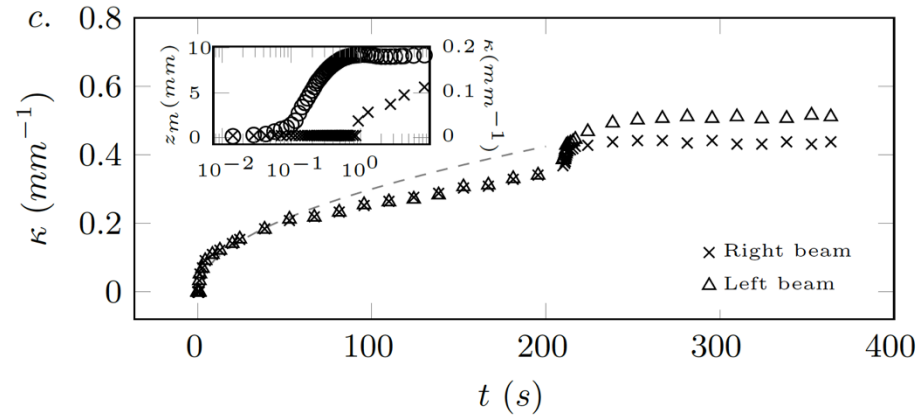
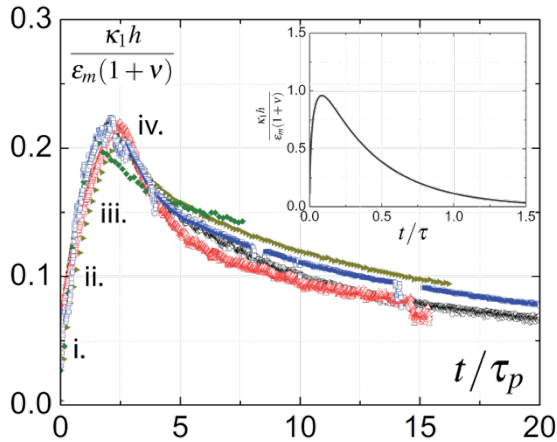
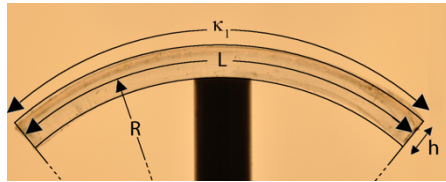




# Capillarity & Swelling



## 2. Swelling-induced bending.



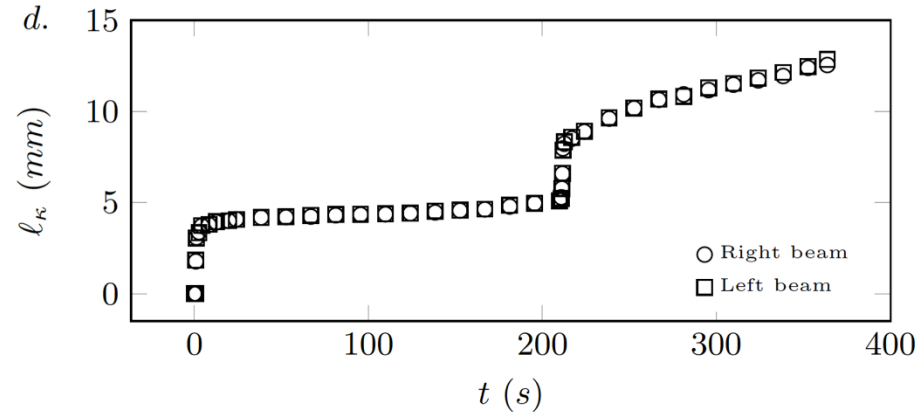
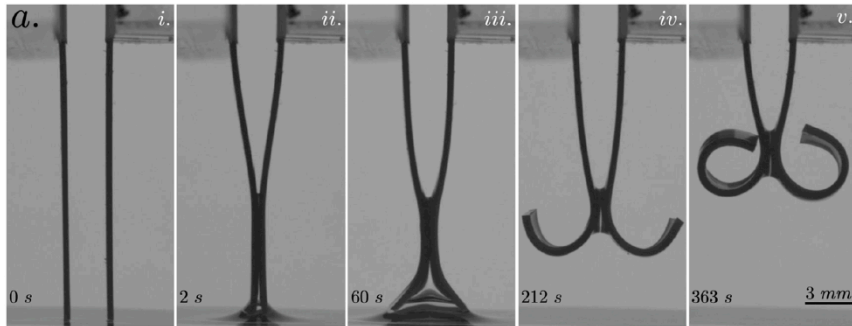
D.P. Holmes, A. Pandey, and S. Protière, *In Preparation*, (2015).

D.P. Holmes, M. Roché, T. Sinha, and H.A. Stone. "Bending and Twisting of Soft Materials by Non-homogenous Swelling" *Soft Matter*, **7**, 5188, 2011.

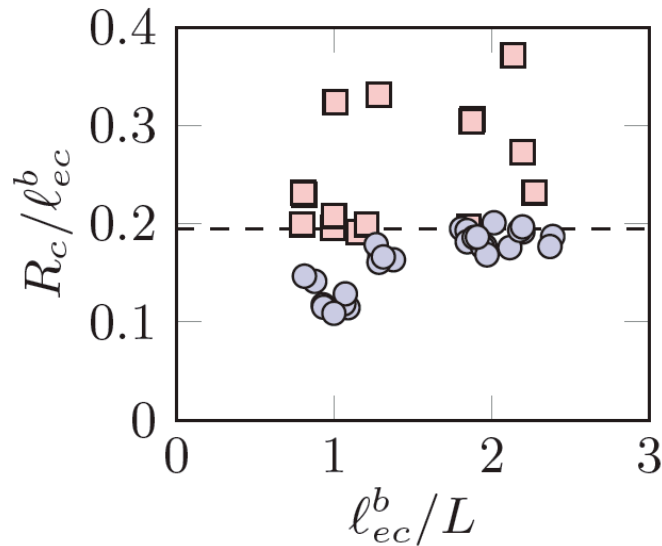




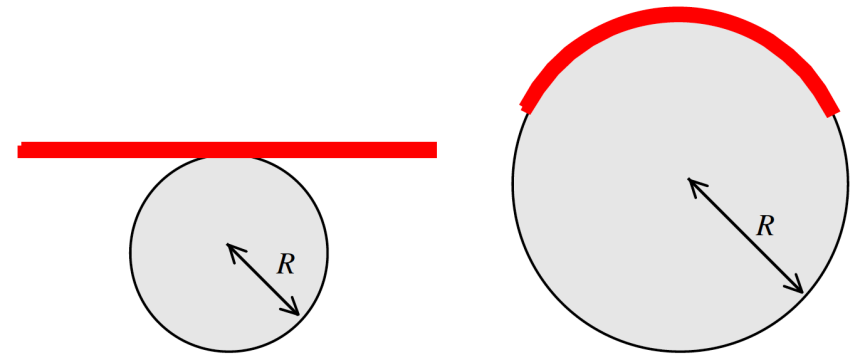
# Capillarity & Swelling



3. Bending dominates surface tension.



Peeling occurs if the curvature exceeds the bending capillary length.



Bending energy

$$U_b \sim \frac{1}{2} B L b \kappa_b^2$$

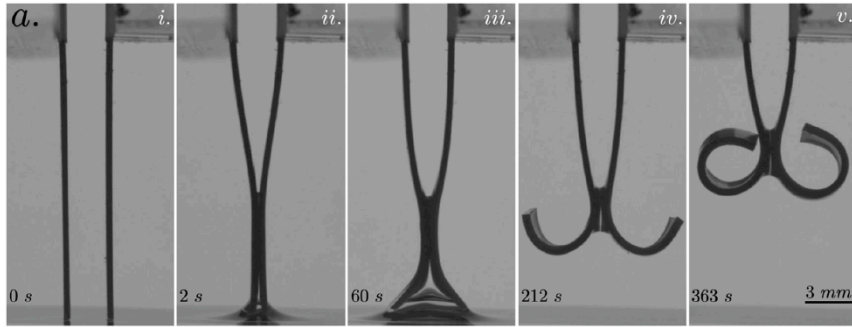
Surface energy

$$U_\gamma \sim 2\gamma L b$$

$$l_{ec}^b \sim \left( \frac{B}{\gamma} \right)^{1/2}$$



# Capillarity & Swelling



## 1. Elastocapillary rise between flexible fibers.

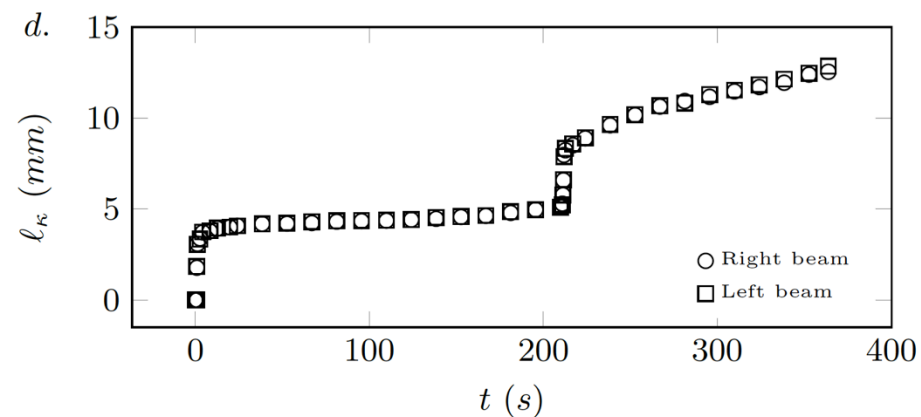
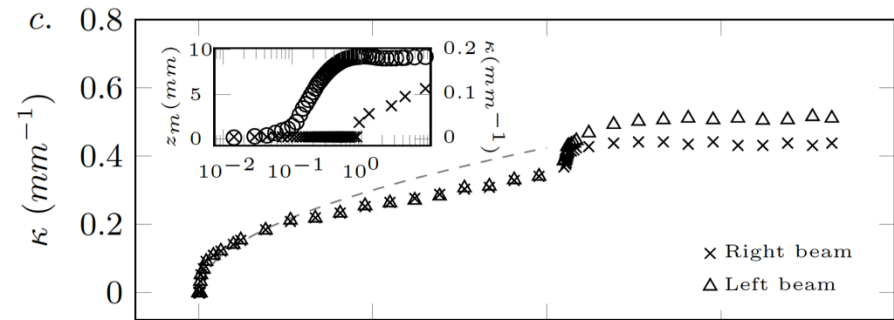
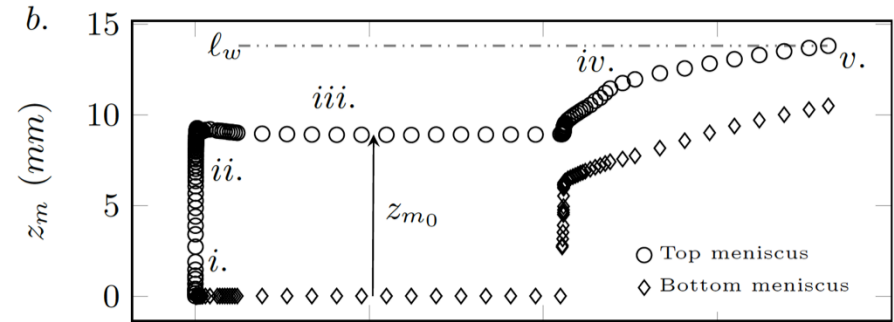
At short times, elastocapillary rise dominates the deformation.

## 2. Swelling-induced bending.

Bending is constrained by surface tension, as the beam bends with a lower curvature than a free swelling beam.

## 3. Bending dominates surface tension.

Separation occurs as the "natural" curvature of the beam exceeds the fluids ability to confine it.





# Capillarity & Swelling

$$U \sim \underbrace{U_m}_{\text{stretching}} + \underbrace{U_b}_{\text{bending}} - \underbrace{U_\gamma}_{\text{surface}}$$

Stretching energy

$$U_m = Eh \int_A \varepsilon^2 dA$$



$$Ehlb\varepsilon^2$$

Bending energy

$$U_b = EI \int_A \kappa^2 dA$$



$$\frac{Eh^3lb}{12} \kappa^2$$

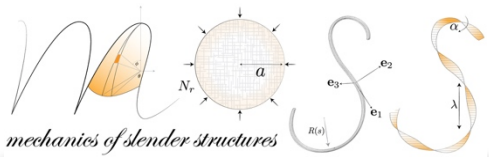
Surface energy

$$U_\gamma = \ell_w \int_A \gamma dA$$



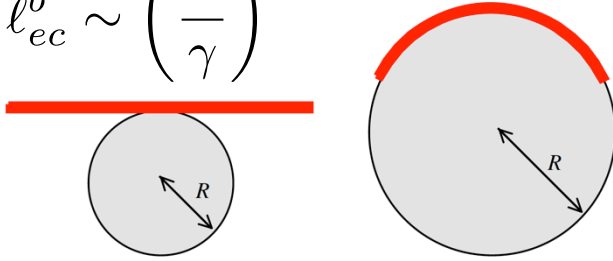
$$\kappa_f \ell_{ec}^b lb \gamma l_w$$

$$Eh\varepsilon^2 + \frac{Eh^3}{12} \kappa_b^2 = \gamma \kappa_f \ell_{ec}^b \quad \xrightarrow[\kappa_b \sim L \quad \kappa_f \sim d_0^{-1}]{S = B/EhL^2b = h^2/12L^2} \quad S \sim \frac{\varepsilon^2}{(L/\ell_{ec})^2 - 1}$$

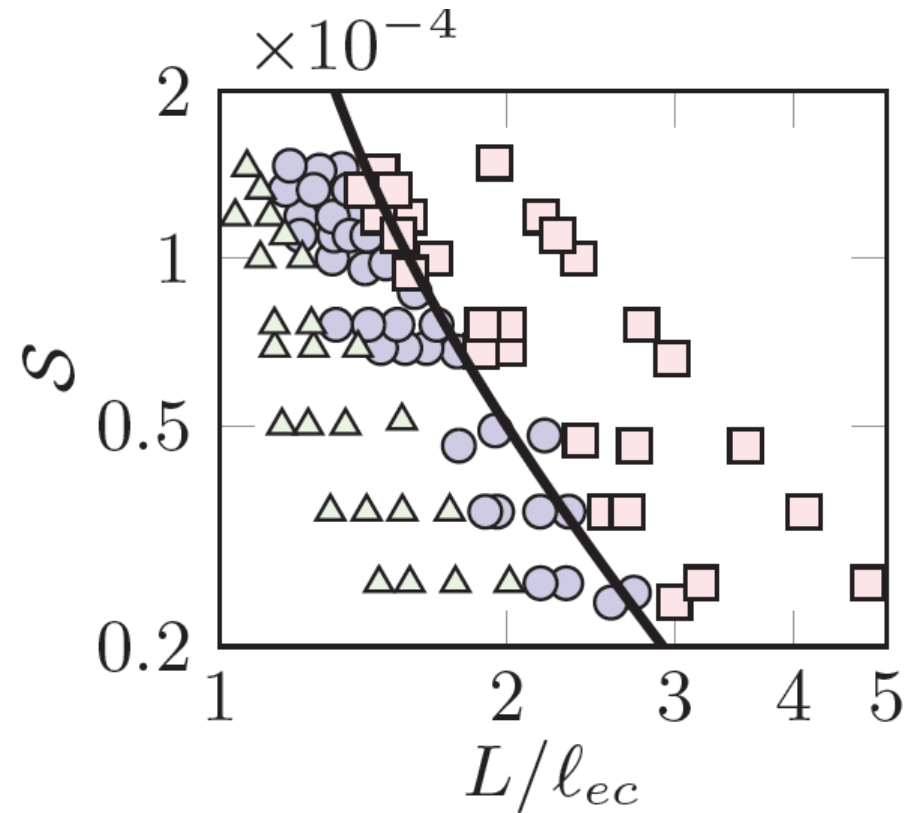
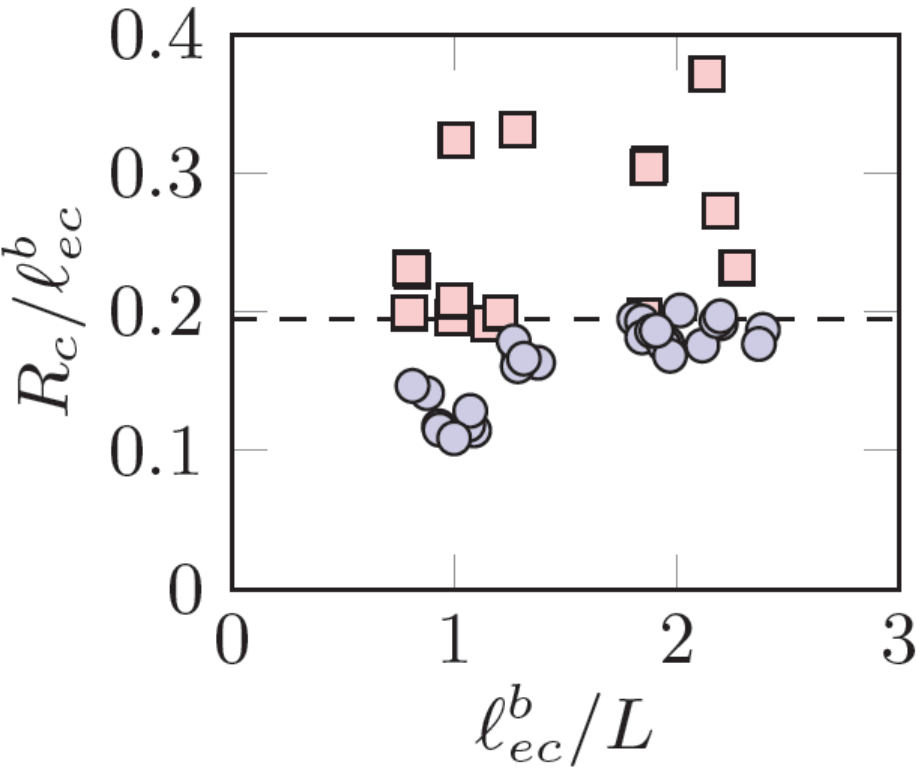


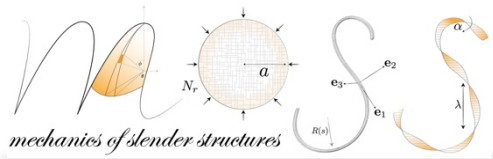
# Capillarity & Swelling

$$\ell_{ec}^b \sim \left( \frac{B}{\gamma} \right)^{1/2}$$



$$S \sim \frac{\varepsilon^2}{(L/\ell_{ec})^2 - 1}$$





# Baobab Flowering







## Confined Fluid Flow: Microfluidics and Capillarity

**Reynolds** Number: Inertia vs. Viscous effects

- Review of characteristic flows...

**Péclet** Number: Transport phenomena in a continuum

- Diffusion, separation, and mixing...

**Geometric** confinement: Controlling and manipulating fluid flow

- Microfluidic fabrication, valving, pumping...

**Capillary** Number: Viscosity vs. Surface tension

- Droplet formation, capillary rise, elasticity...