

I. Protein Structure

Lecture 9 (10/2/23)

A. Primary

1. Peptide Bond
 - a. Planar, strong, ϕ/ψ angles
2. Determination
 - a. Sequence determination; CHEMICAL
 - i. aa composition; Divide & conquer; Edman degradation
 - b. Sequence determination; PHYSICAL
 - i. Tandem Mass Spectrometry for proteins
 - c. Sequence determination; BIOLOGICAL
 - i. Genome sequenced; need partial sequence
 - d. Determination of Disulfide bonds

B. Secondary

1. Conformational structure; Levinthal paradox
2. Pauling & Corey's predictions
 - a. α -Helix
 - b. β -sheets/strands
 - c. Connections between β -strands
 - d. Connections between α -helices; angle not important
3. Super secondary structure
 - Reading: Ch4; 119-122, 125-127, 131-133; 114-115, 120-121, 123-124

C. Tertiary

1. Picturing and classifications
 - Homework #9
 2. Topology
 3. Domains
 4. Intrinsically disordered
 5. Stability
- NEXT**
- Reading: Ch4; 116-117, 126-128
 - Homework #10

Determination of primary structure

THREE basic ways to know the primary structure:

CHEMICAL

Edman Degradation requires >100 pmole (1-5 μ g)

PHYSICAL

MS/MS requires >1 -10 pmole (100-500 ng)

BIOINFORMATICAL

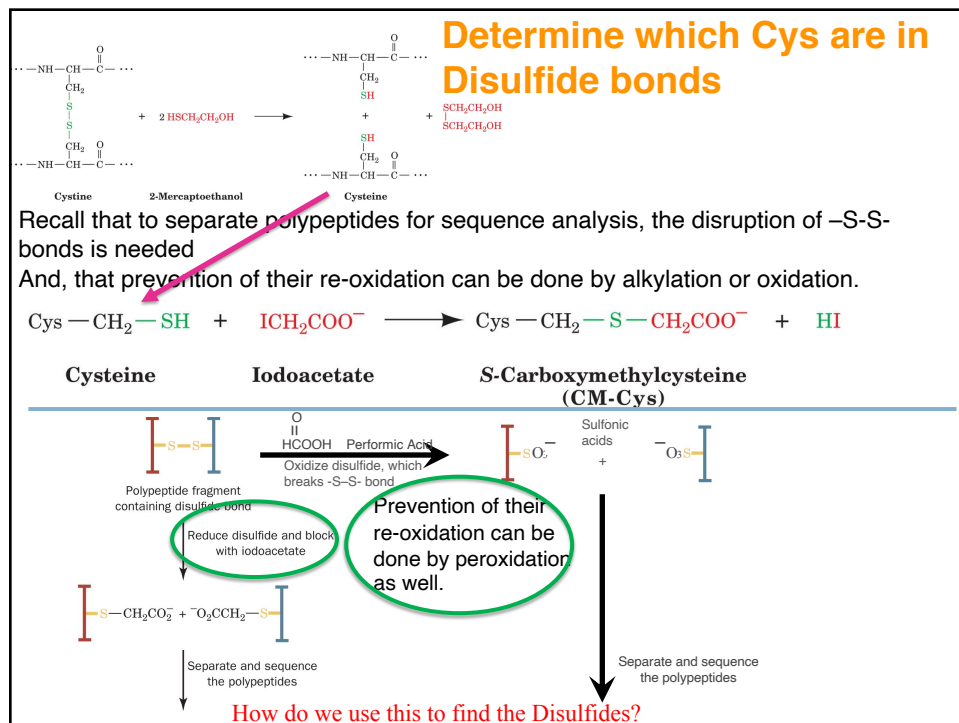
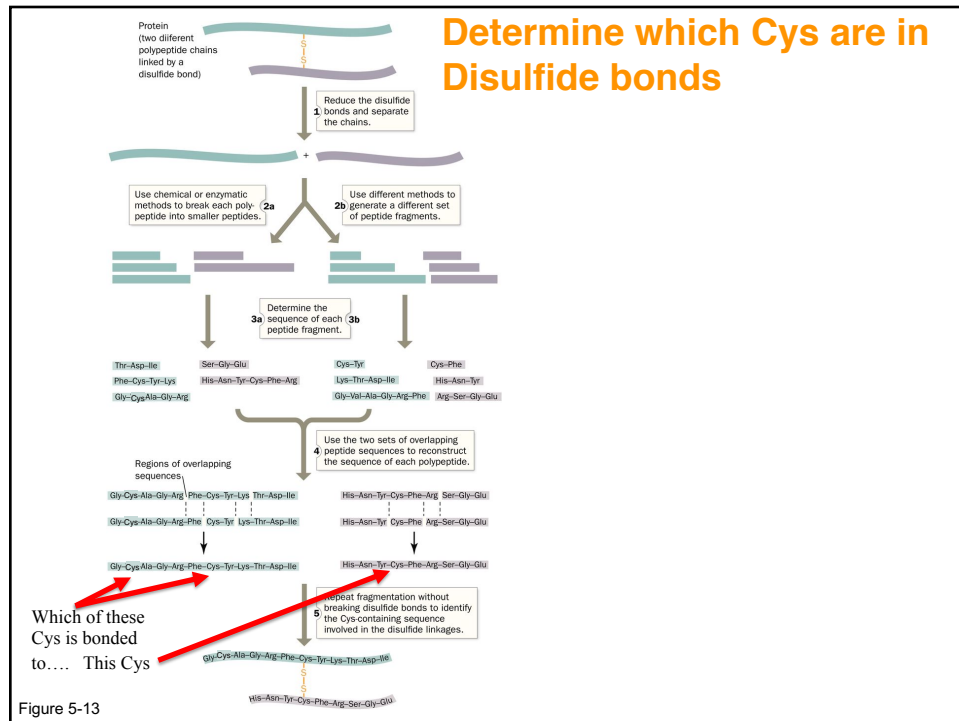
We just went through the CHEMICAL and PHYSICAL.

The BIOINFORMATICAL method requires information from chemical or physical, but only a limited amount of sequence.

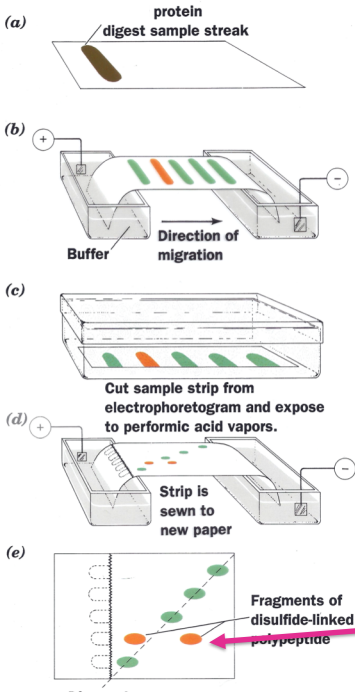
- Example: a sequence of 6 AA is only possible as one of 20^6 possible hexa-peptide sequences (1 of 64×10^6).
- There are no more than 50,000 protein-coding genes with ≤ 400 AA on average. This is $\sim 20 \times 10^6$ possible unique sequences.
- So, a hexamer is likely to appear only once; an octomer even rarer.
- Once you have at least 6-8 AA sequence, you can compare that to all possible proteins encoded in the **entirety of the gene sequences** (**genome**) for a species for which the **genome** is known. Then using appropriate bioinformatic tools, you can derive the entire protein sequence.

There is one remaining issue: Where are the Disulfides, if any?

.....This requires chemical and/or physical methods



Determine which Cys are in Disulfide bonds



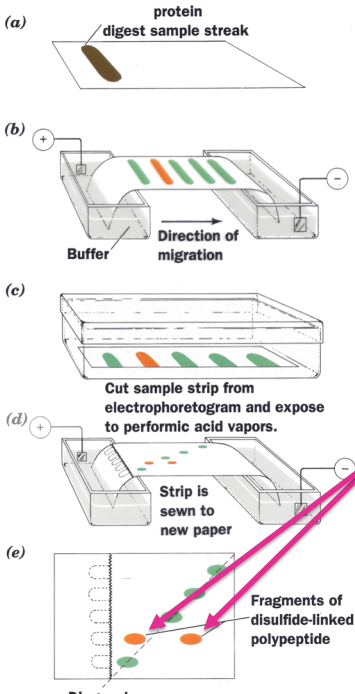
The diagram illustrates the 2D-diagonal electrophoresis process in five steps: (a) protein digest sample streak, (b) electrophoresis in a buffer with a direction of migration arrow, (c) cutting a sample strip from the electrophoretogram, (d) exposing the strip to performic acid vapors and sewing it to new paper, and (e) the resulting 2D gel showing fragments of disulfide-linked polypeptides off the diagonal.

We change the protection step for sulfhydryl groups.

- Cleave/Protect (reduction/alkylation or oxidation) AFTER fragmentation
- Separate fragments as before, but any linked by $-S-S-$ will not separate and remain together (e.g., orange peptide).
- THEN break $-S-S-$ bonds, and re-separate.

Determine the sequence of those peptides that fall off the diagonal by either Edman degradation or tandem MS/MS. Technique is called "2D-diagonal electrophoresis."

Determine which Cys are in Disulfide bonds

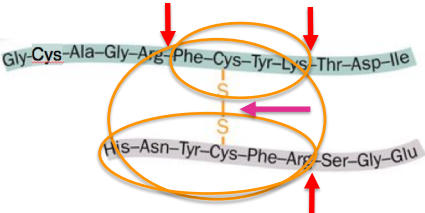


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The diagram shows two polypeptide chains linked by two disulfide bonds (S-S). The top chain is Gly-Cys-Ala-Gly-Arg-Phe-Cys-Tyr-Lys-Thr-Asp-Ile. The bottom chain is His-Asn-Tyr-Cys-Phe-Arg-Ser-Gly-Glu. Red arrows indicate the cleavage sites for the disulfide bonds.

Protein Structure

Conformational Structure

How does the polypeptide chain fold?

Protein Structure

Conformational Structure

How does the polypeptide chain fold?

- 1) **primary structure** – sequence of amino acids
- 2) **secondary structure** – small units of repetitive structure
- 3) **tertiary structure** – overall 3D shape
- 4) **quaternary structure** – shape of ≥ 2 chains

4 levels of protein structure

In order to understand these levels of structure, you need to understand the nature of the polymer first.

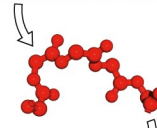
In other words, the linkage or
PEPTIDE BOND

Primary structure
The sequence of amino acid residues

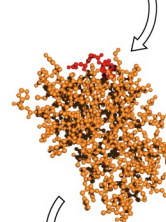
-Glu-Ser-Phe-Gly-Asp-

Secondary structure

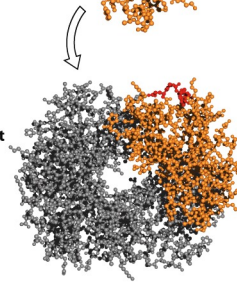
The localized conformation of the polypeptide backbone



Tertiary structure
The three-dimensional structure of an entire polypeptide, including all its side chains



Quaternary structure
The spatial arrangement of polypeptide chains in a protein with multiple subunits



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Protein Structure-Secondary

The 4 S's for secondary structure:

Size	-dependent on number of amino acids
Solubility	-dependent on AA composition and shape
Stability	-complex and not well understood
Shape	

Why is there Secondary Structure?

Protein Structure-Secondary

The Levinthal Paradox (1969):

Theoretical calculation:

Consider just the α -carbon backbone.....

ASSUME there are 4 clearly different angles allowed of all the angles at the α -carbon (ϕ and ψ), then each residue has $2 \times 4 = 8$ degrees of freedom.

For a protein of 100 residues, there are 8^{100} possible conformations to "test" for optimal energetics

$8^{100} = 2 \times 10^{90}$ different conformations

At 1.1 quintillion "tests" per second ($0.91/\text{attosec}$)*, this is 1.8×10^{71} seconds to find the best.

$\Rightarrow 5.7 \times 10^{63}$ years

Well The age of the universe is 14×10^9 years

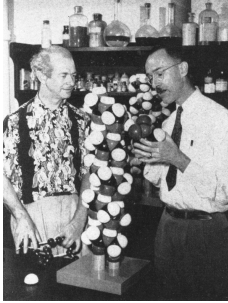
The shortcut proteins use to fold is the use of 2° structure where most of these degrees of freedom are prescribed by a regular structure.

* 10^{18} FLOPS is fastest supercomputer

What are these "regular structures?"

Secondary Structure

Protein Structure-Secondary



In the early 1950's, Linus Pauling and Robert Corey predicted some rules that proteins should follow to find the lowest energy conformation.

- 1) The peptide bond must be planar without free rotation
- 2) The degree of H-bonding should be maximized to achieve the lowest energy state [consider energetic consequences in the (unfolded)^{water} \rightleftharpoons (folded)^{water} transition]
- 3) The best H-bonds are linear
- 4) There should be repeating units of conformation (same) as you go from one residue to the next

Using these rules they predicted two basic structures:

α -helix

β -sheet

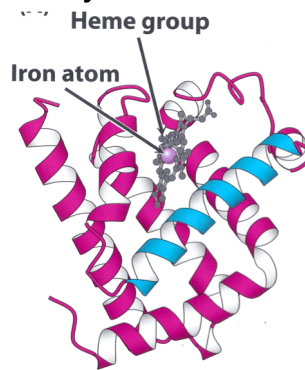
Protein Structure-Secondary



John Cowdery
Kendrew
(1917 - 1997)

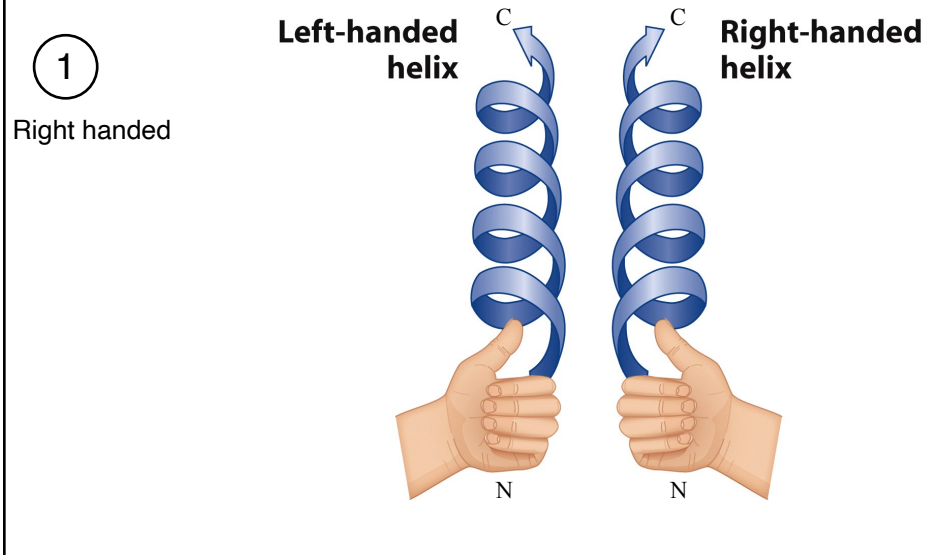
There were no known protein structures until 1957, when Kendrew solved the structure of myoglobin:

Imagine the excitement when indeed there were the very helices Pauling predicted!



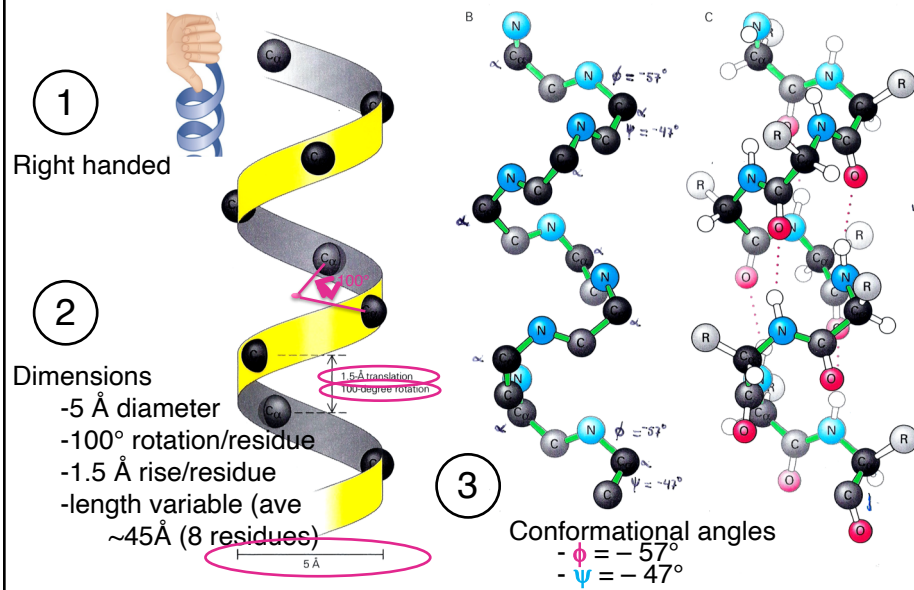
Protein Structure-Secondary

The α Helix



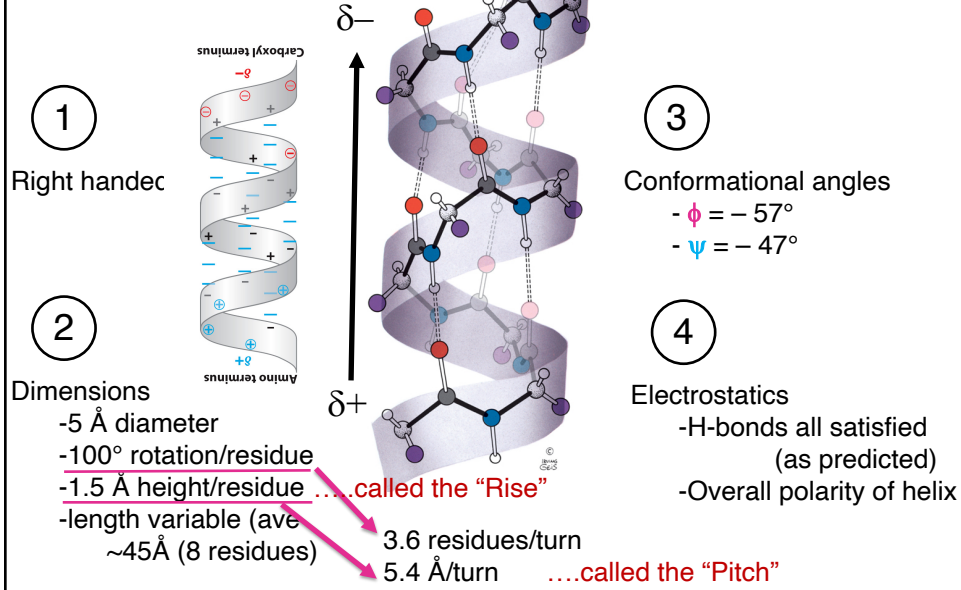
Protein Structure-Secondary

The α Helix



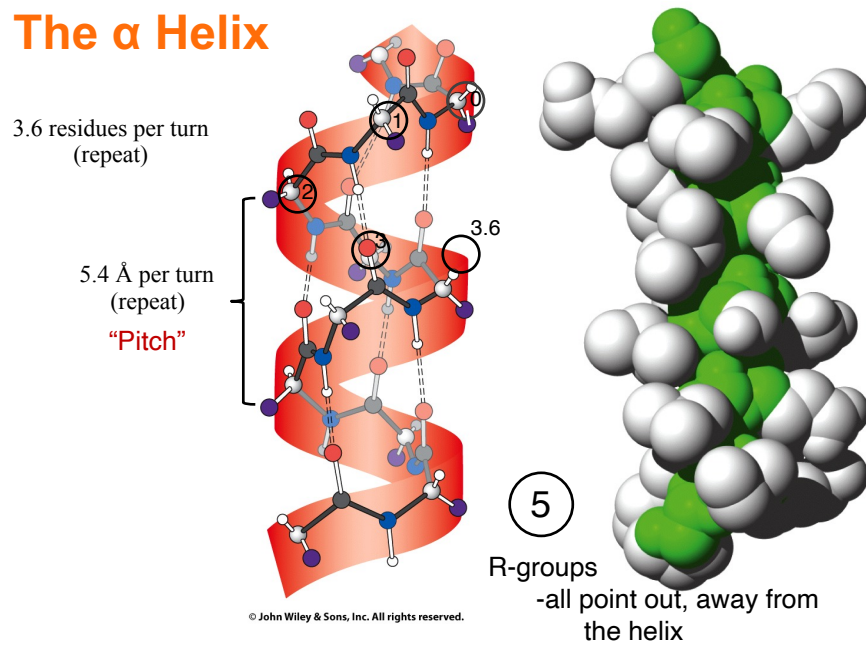
Protein Structure-Secondary

The α Helix



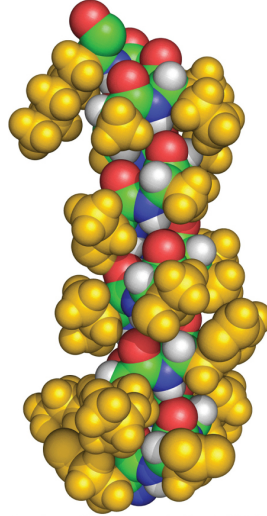
Protein Structure-Secondary

The α Helix



Protein Structure-Secondary

The α Helix: Space Filling Model



Helices can be hydrophobic

Helices can be hydrophilic

Helices can be mixed: amphipathic

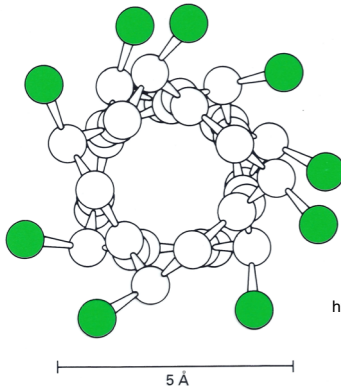
Based on an X-ray structure by Ilme Schlichting, Max Planck Institut für Molekulare Physiologie, Dortmund, Germany. PDBid 1A6M (for the definition of "PDBid" see Section 6-2E).

Oxy-Myoglobin
PDBid 1A6M

Protein Structure-Secondary

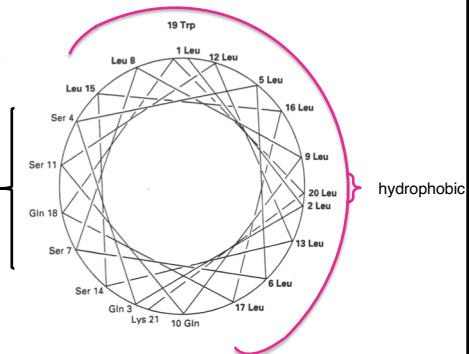
The α Helix

Helices can be mixed: amphipathic

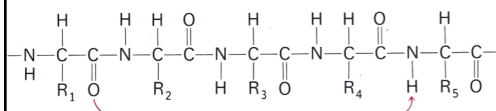


hydrophilic

LLQSLLSLLQSLLSLLLQW



hydrophobic



Protein Structure-Secondary

Table 6-1 Propensities of Amino Acid Residues for α Helical Conformations

Residue	P_{α}
Ala	1.42
Arg	0.98
Asn	0.67
Asp	1.01
Cys	0.70
Gln	1.11
Glu	1.51
Gly	0.57
His	1.00
Ile	1.08
Leu	1.21
Lys	1.16
Met	1.45
Phe	1.13
Pro	0.57
Ser	0.77
Thr	0.83
Trp	1.08
Tyr	0.69
Val	1.06

Source: Chou, P.Y. and Fasman, G.D., *Annu. Rev. Biochem.* **47**, 258 (1978).

Conformational angles

- $\phi = -57^{\circ}$
- $\psi = -47^{\circ}$

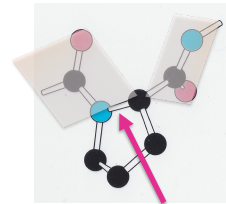
The α Helix: Propensities

Like:

- prefer small, medium, hydrophobic/charged
- no steric hindrance at C_{β}
- Glu, Met, Ala, Leu, Lys, Phe, Gln

Don't Like:

- Pro
- Gly



Pro has fixed $\phi = -65^{\circ}$

Gly is opposite: has too many degrees of freedom

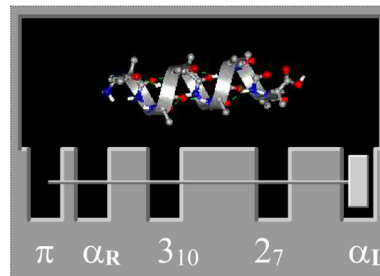
92

Protein Structure-Secondary

The α Helix: other helices

Increasing pitch (distance per turn) & rise (distance per residue) \rightarrow

- The α -helix is a common secondary structure in proteins.
- The α -helix is a spiral structure with defined dimensions, rise and pitch.
- The α -helix is stabilized by hydrogen bonds between residues in the helix.
- Several types of helices (such as the π , α_R , 3_{10} , 2_7 and α_L) can form intramolecular hydrogen bonds.

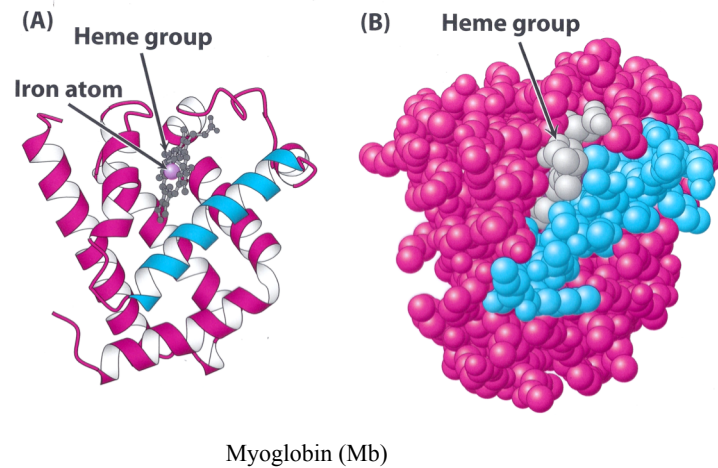


	4.1 ₁₆	3.0 ₁₀	
Repeat units/turn \rightarrow	3.6 ₁₃	2.2 ₇	
# of atoms in one turn \rightarrow			

Protein Structure-Secondary

The α Helix: examples

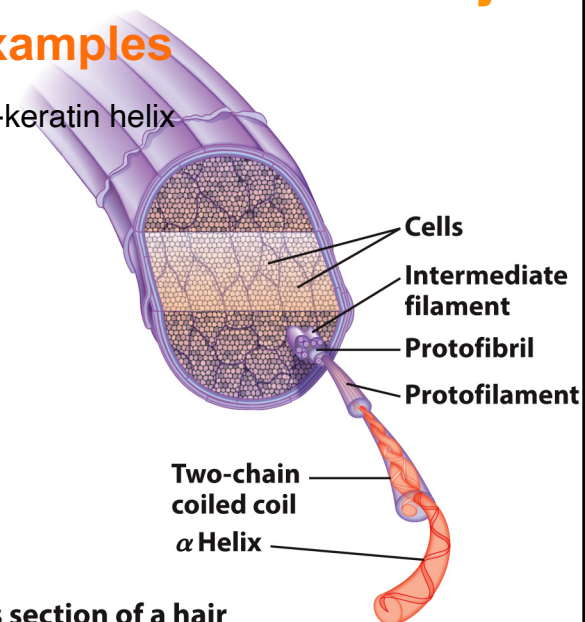
Globular Proteins



Protein Structure-Secondary

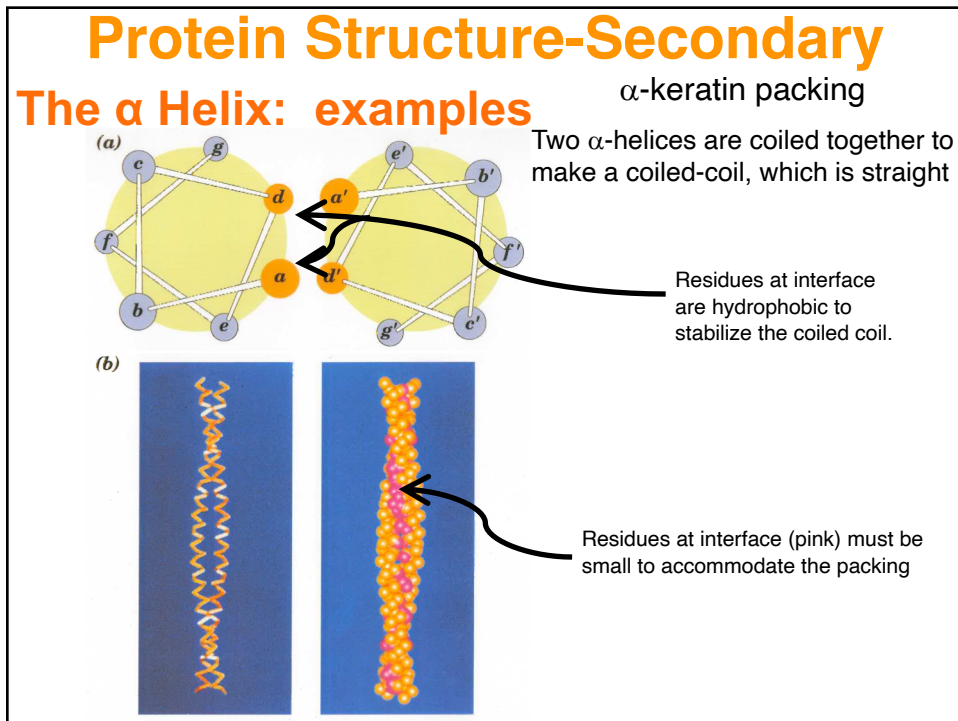
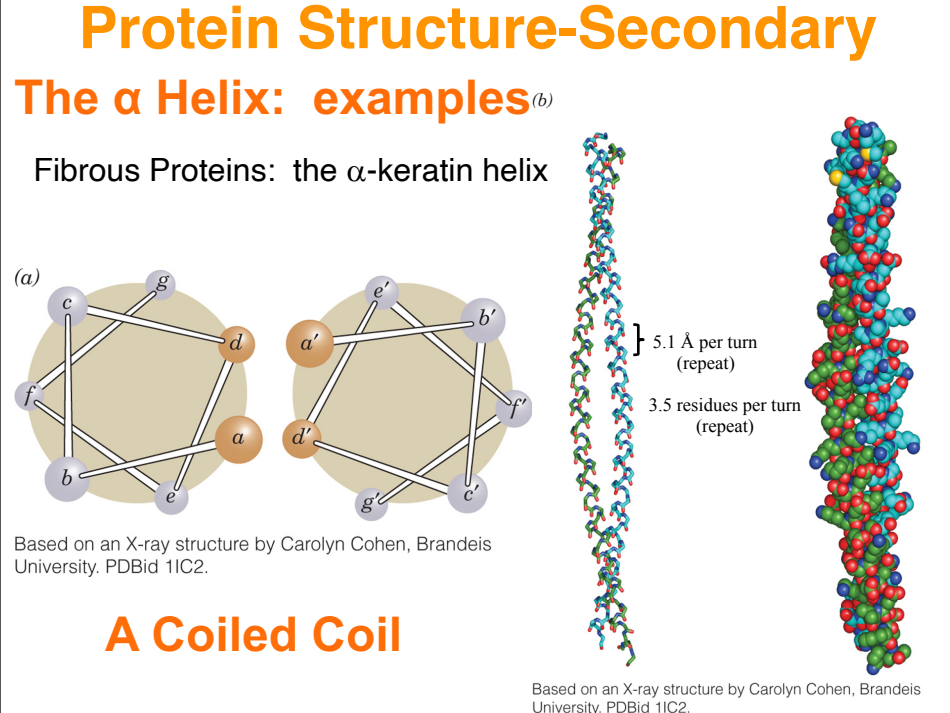
The α Helix: examples

Fibrous Proteins: the α -keratin helix



Cross section of a hair

Figure 4-11b



Protein Structure-Secondary

The α Helix: examples

Fibrous Proteins: the α -keratin helix

a-b-c-d-e-f-g

$h\phi$ $h\phi$ $+/-$ $+/-$

(L,V,A,G) (E,D,R)

b-c-&f tend to be Ser, Gln, Cys

(a)

(b)

A Coiled Coil

Protein Structure-Secondary

a&d

~25-30%

b,c,&f

~33%

e&g

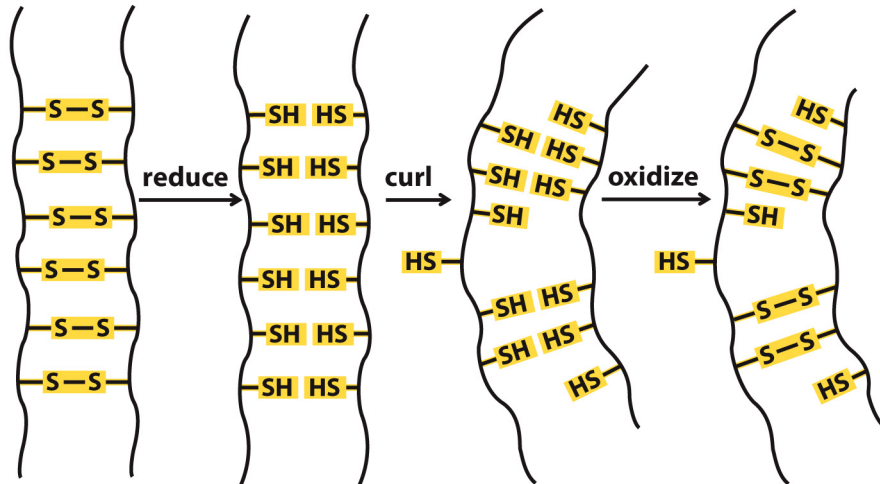
~15-20%

Notice the >10% Cys

Amino Acid	α -Keratin (Wool)
Gly	8.1
Ala	5.0
Ser	10.2
Val + Gln	12.1
Cys	11.2
Pro	7.5
Arg	7.2
Leu	6.9
Thr	6.5
Asp + Asn	6.0
Val	5.1
Tyr	4.2
Ile	2.8
Phe	2.5
Lys	2.3
Trp	1.2
His	0.7
Met	0.5

Note: The three most abundant amino acids in each protein are indicated in red. Values given are in mole percent.

Chemistry of Permanent Waving



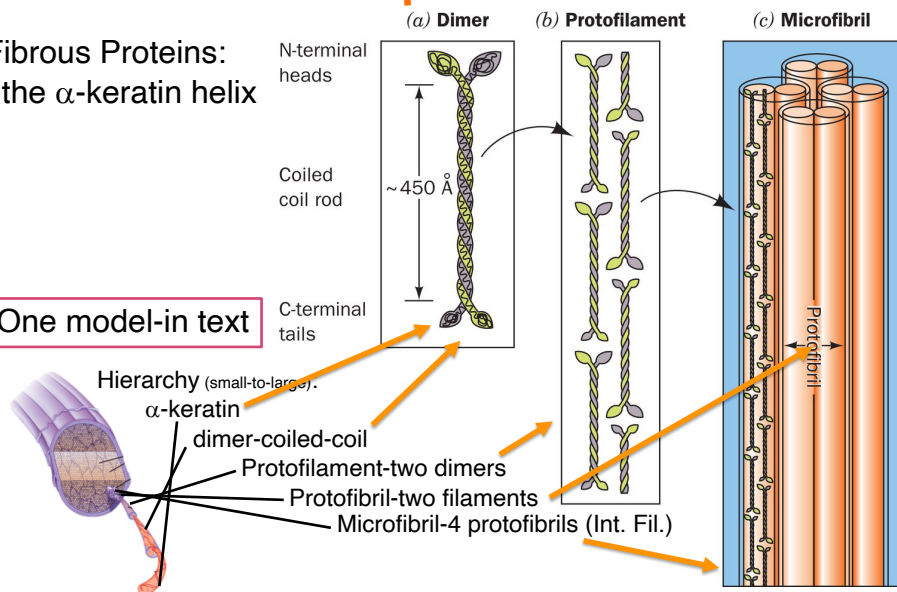
Box 4-2
Lehninger Principles of Biochemistry, Seventh Edition
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Protein Structure-Secondary

The α Helix: examples

Fibrous Proteins:
the α -keratin helix

One model-in text



Protein Structure-Secondary

The α Helix: examples

Fibrous Proteins:
the α -keratin helix

FIGURE 6.11
The coiled-coil structure of α -keratin.
(a) Structure of microfibrils formed by two different modes of interaction of protofibrils.
(b) Formation of a protofibril from two coiled coils.

Another model

Hierarchy:

- α -keratin
- dimer-coiled-coil
- Protofibril-coiled two dimers
- Microfibril-8 protofibrils

Protein Structure-Secondary

The α Helix: Summary

Amino terminus

5.4 Å
(3.6 residues)

Carboxyl terminus

(a)

(b)

δ^-

δ^+

(c)

(d)

Figure 4-4

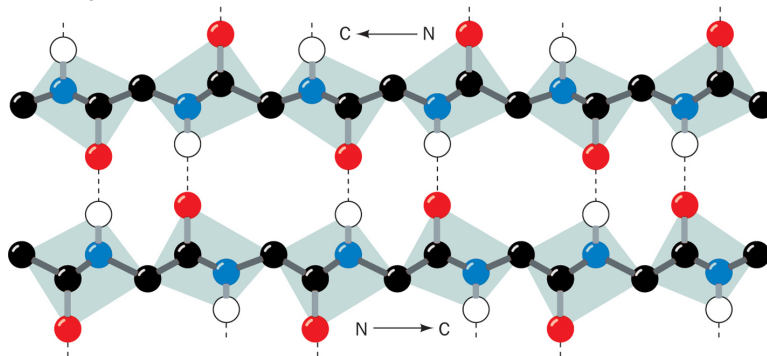
- Right handed helix
- 3.6 aa per turn
- 5.4 Å rise per turn
- Carbonyl of residue “n” is H-bonded to NH of n+4 residue
- Has tightly packed core of main-chain atoms
- R-groups project outward
- Has overall macro-dipole (N-term +; C-term -)
- Can be amphipathic

Secondary Structure

Protein Structure-Secondary β Sheets- antiparallel

Using his rules, Pauling predicted two basic structures:
 α -helix
 β -sheet, which he called a “back-and-forth” structure

(a) Antiparallel



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Protein Structure-Secondary

β Sheets- antiparallel

1

Right handed
(2.0₇)

2

Dimensions

-it's a sheet

Almost fully extended:

-3.4 Å rise/residue

-2 residues/repeat → Pitch is 6.8 Å

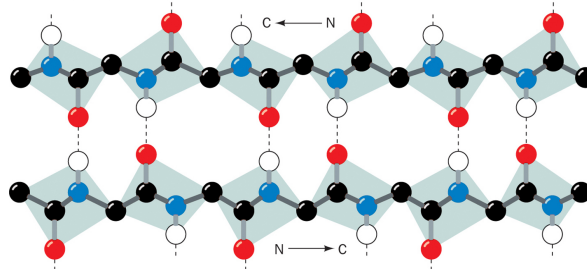
-length variable

3

Conformational angles

- $\phi = -139^\circ$

- $\psi = +135^\circ$



4

Electrostatics

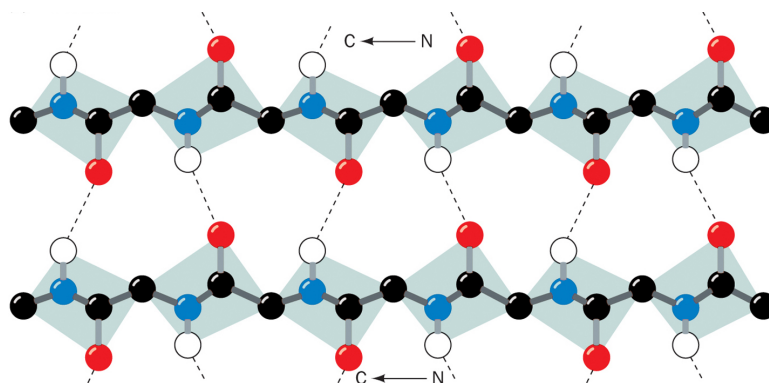
-H-bonds all satisfied
(as predicted)

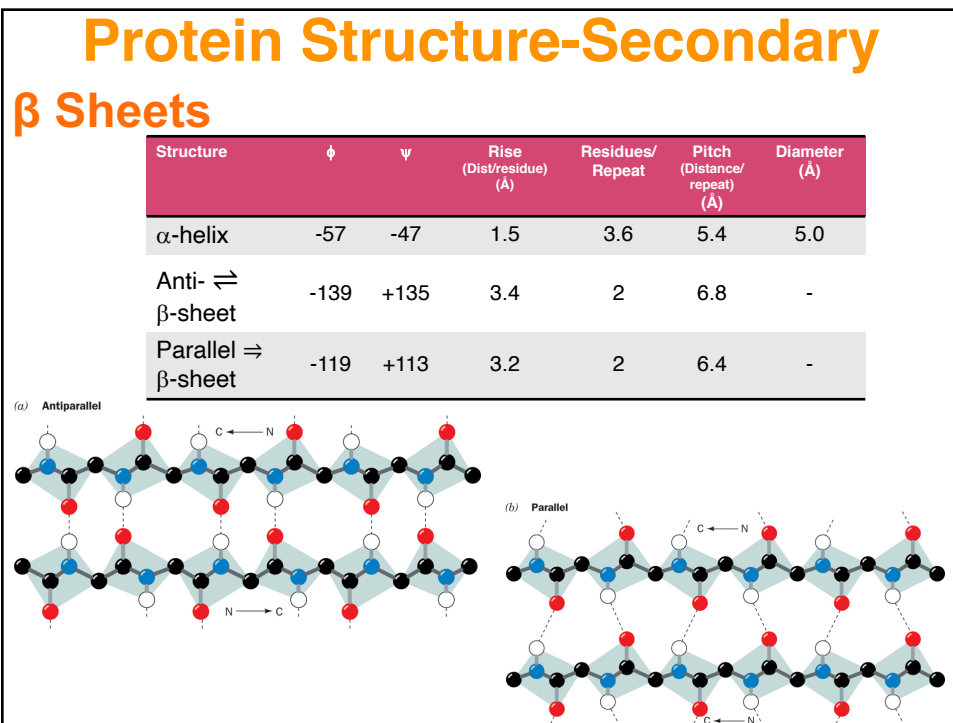
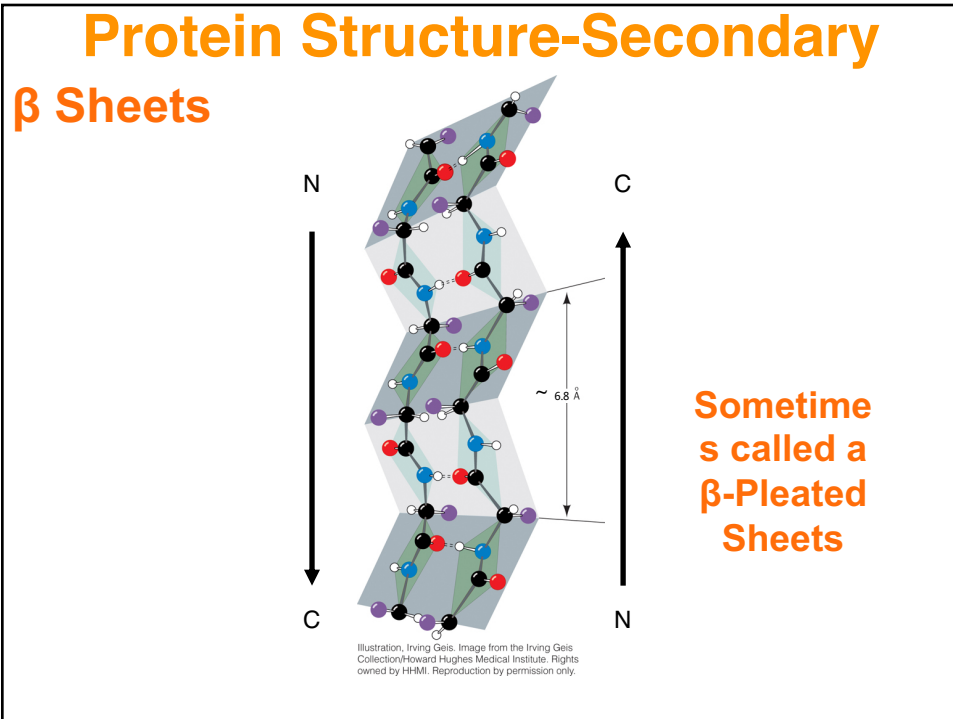
-no polarity

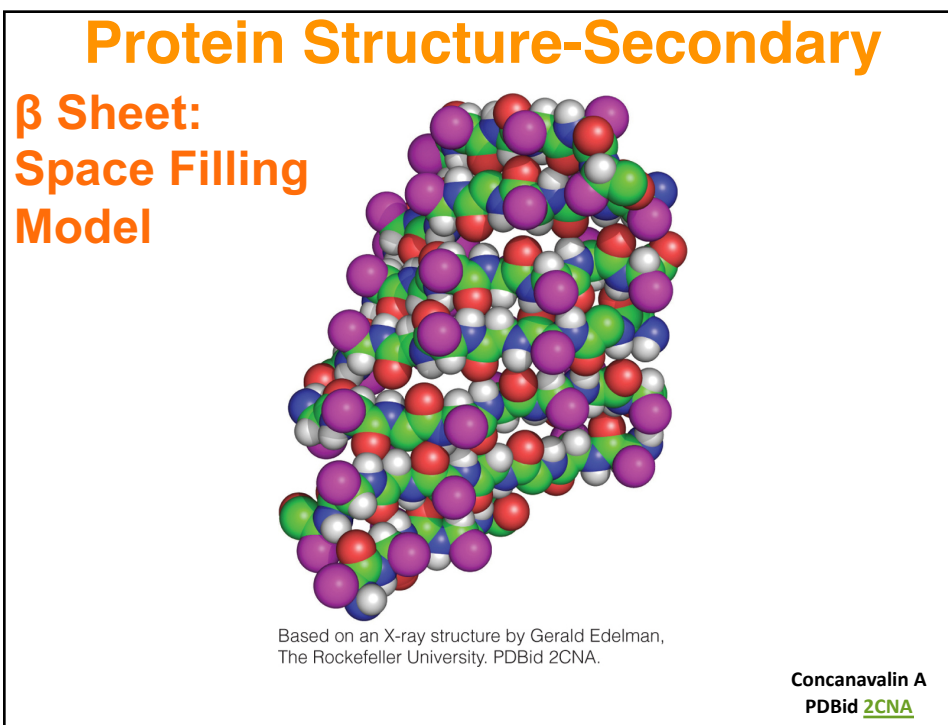
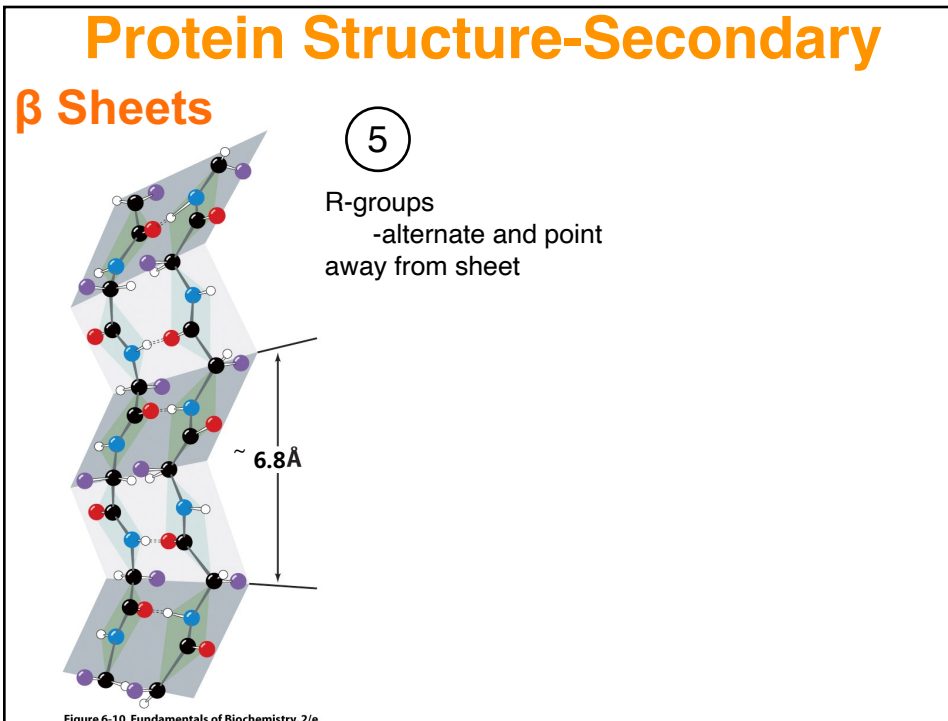
Protein Structure-Secondary

β Sheets- parallel

Pauling did not predict a β -sheet made of β -strands going the same direction.







Protein Structure-Secondary

The β Sheet: Propensities

Table 6-1 Propensities of Amino Acid and β Sheet Conformations

Residue	P_{β}
Ala	0.83
Arg	0.93
Asn	0.89
Asp	0.54
Cys	1.19
Gln	1.10
Glu	0.37
Gly	0.75
His	0.87
Ile	1.60
Leu	1.30
Lys	0.74
Met	1.05
Phe	1.38
Pro	0.55
Ser	0.75
Thr	1.19
Trp	1.37
Tyr	1.47
Val	1.70

Like:

- prefer large, bulky groups
- Val, Ile, Leu, Tyr, Trp, Phe

Don't Like:

- Pro (same reason)
- Glu/Asp/Lys (full charges too close)
- Gly (same reason)

Protein Structure-Secondary

The β Sheet: examples

Globular Proteins

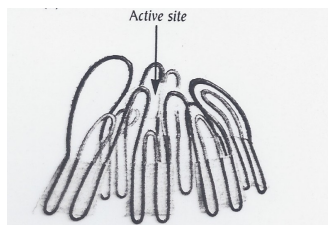
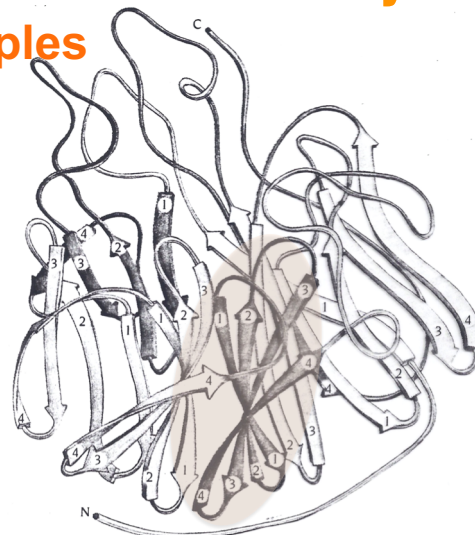


Figure 5.9 The six four-stranded motifs in a single subunit of neuraminidase form the six blades of a propeller-like structure. A schematic diagram of the subunit structure shows the propeller viewed from its side (a). An idealized propeller structure viewed from the side to highlight the position of the active site is shown in (b). The loop regions that connect the motifs (red in b) in combination with the loops that connect strands 2 and 3 within the motifs (green in b) form a wide funnel-shaped active site pocket. [(a) Adapted from P. Colman et al., *Nature* 326: 358-363, 1987.]



Neuraminidase

- 6 x 4-stranded β Sheets: make a funnel to active site
- right-handed twist

Protein Structure-Secondary

The β Sheet: examples

Fibrous Proteins: Silk fibroin (β -keratin)

Silk

all parallel β -sheet

Sequence repeats:



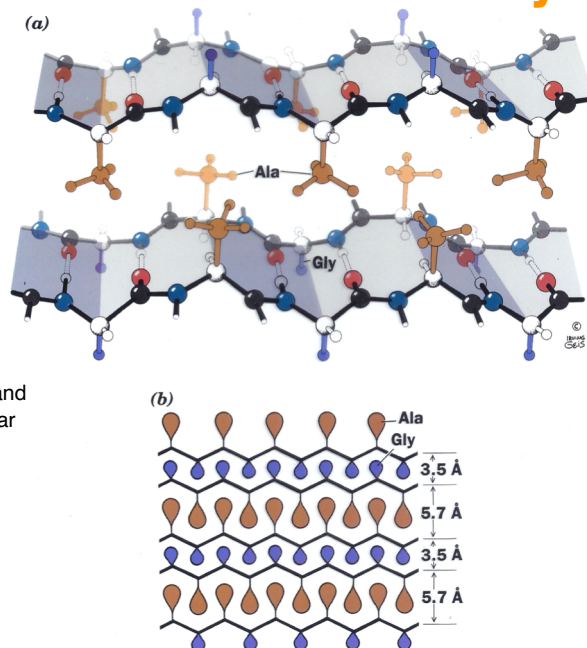
- Gly is every-other residue, and Ala as well
- Recall the alternating R-groups
- Therefore, Gly is all on one side of sheet, and Ala on the other side of sheet

Protein Structure-Secondary

The β Sheet: examples

Silk fibroin (β -keratin)

Silk's tensile strength is comparable to that of steel and about half as strong as Kevlar



Protein Structure-Secondary β Sheets

Amino Acid	α -Keratin (Wool)	Fibroin (Silk)
Gly	8.1	44.6
Ala	5.0	29.4
Ser	10.2	12.2
Glu + Gln	12.1	1.0
Cys	11.2	0
Pro	7.5	0.3
Arg	7.2	0.5
Leu	6.9	0.5
Thr	6.5	0.9
Asp + Asn	6.0	1.3
Val	5.1	2.2
Tyr	4.2	5.2
Ile	2.8	0.7
Phe	2.5	0.5
Lys	2.3	0.3
Trp	1.2	0.2
His	0.7	0.2
Met	0.5	0

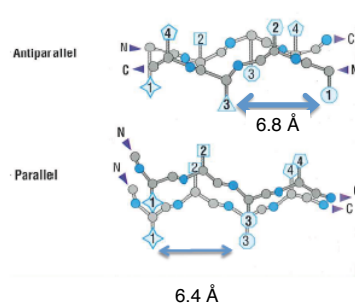
Note: The three most abundant amino acids in each protein are ~~indicated in red~~ boxed. Values given are in mole percent.

85% G, A, S

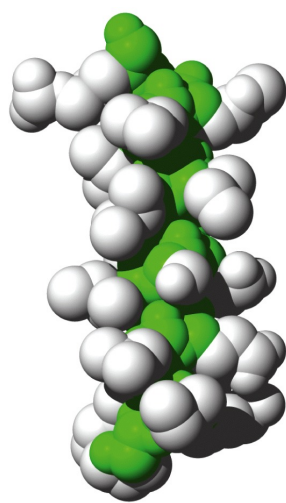
Protein Structure-Secondary

Summary of β Sheet

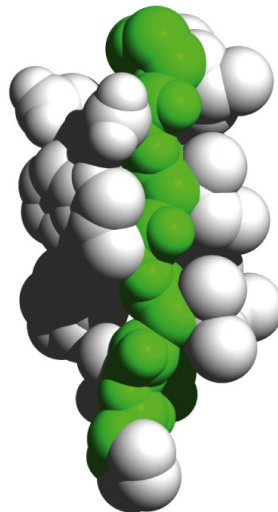
- Parallel or antiparallel
- ~ 7.0 Å between R groups
- ~ 3.5 Å between alpha carbons
- Alternating residues face opposite sides
- Extended structure
- Right-handed twist



Protein Structure-Secondary



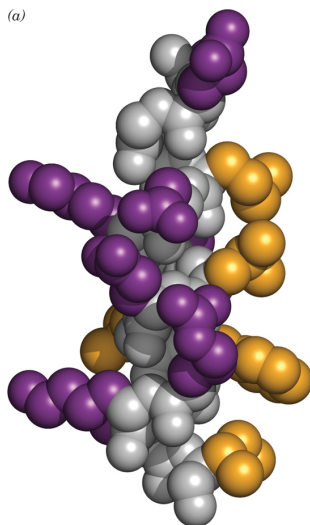
α -helix



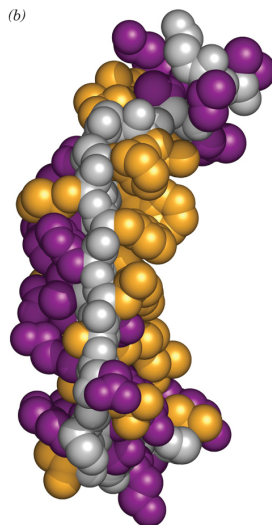
β -sheet

Protein Structure-Secondary

(a)



(b)



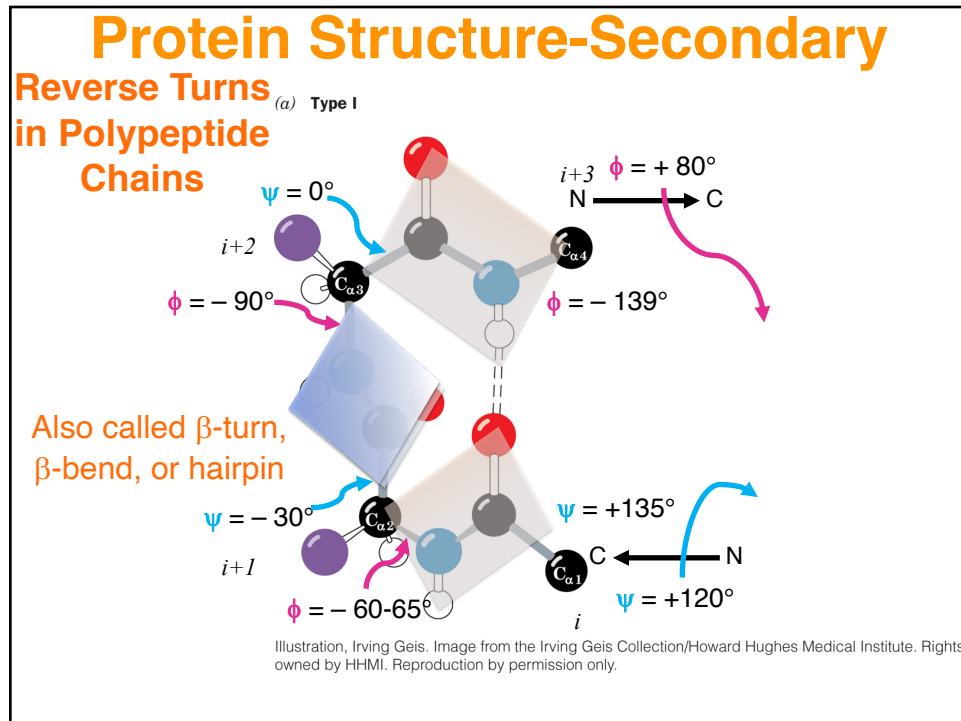
Purple=polar
Gold=hydrophobic

Recall that β -sheets
require strands from
different parts of the same
polymer, so....

How are the connected?

Based on X-ray structures by Ilme Schlichting, Max Planck Institut für Molekulare Physiologie, Dortmund, Germany, and Gerald Edelman, The Rockefeller University, PDBids 1A6M and 2CNA.

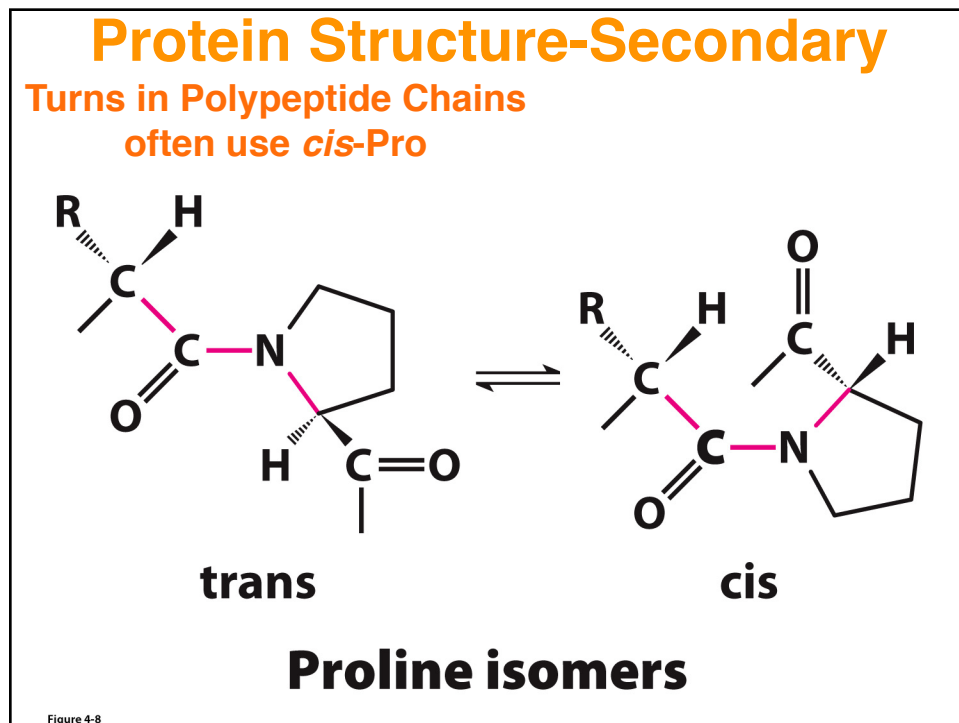
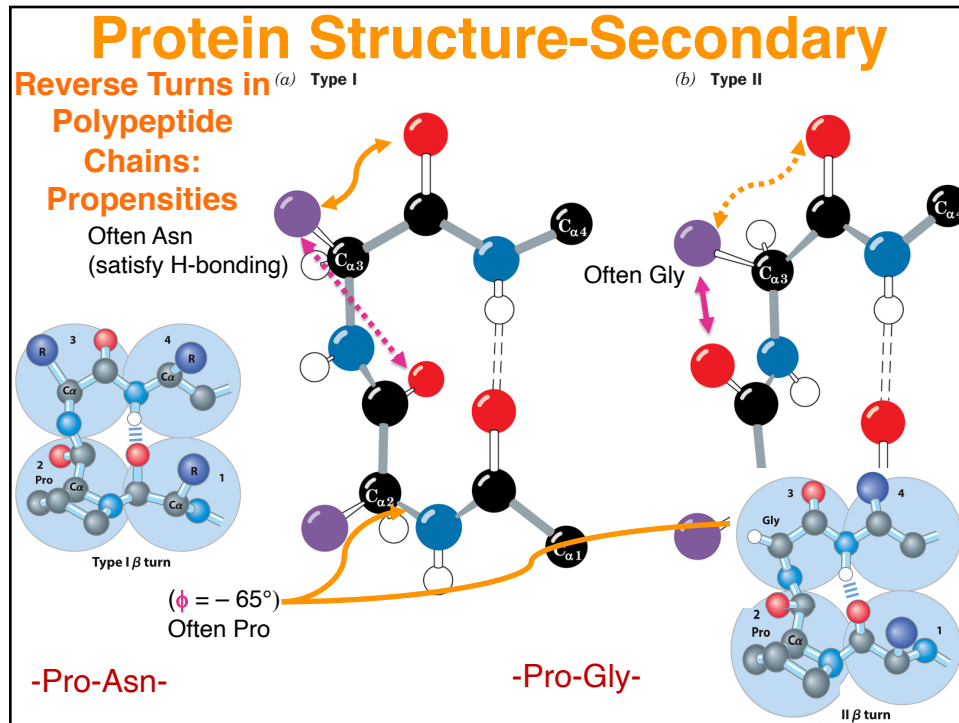
Oxy-Myoglobin and Concanavalin A
PDBids 1A6M and 2CNA



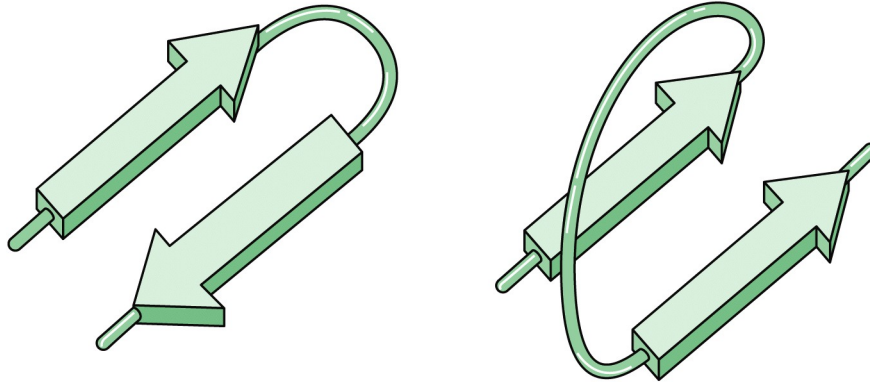
Protein Structure-Secondary

Structure	ϕ	ψ	Rise (Dist/residue) (Å)	Residues/Repeat	Pitch (Distance/repeat) (Å)	Diameter (Å)
α -helix	-57	-47	1.5	3.6	5.4	5.0
Anti- \Rightarrow β -sheet	-139	+135	3.4	2	6.8	-
Parallel \Rightarrow β -sheet	-119	+113	3.2	2	6.4	-
β -turn-Type I				4	0	-
<i>i</i> + 1	-60	-30	-			
<i>i</i> + 2	-90	0	-			
β -turn-Type II				4	0	-
<i>i</i> + 1	-60	120	-			
<i>i</i> + 2	80	0	-			

Start and stop with same angles



Protein Structure-Secondary



Reverse Turns are used for anti-parallel sheets, but how do parallel strands find each other to make a sheet?

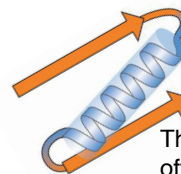
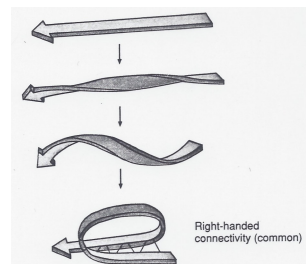
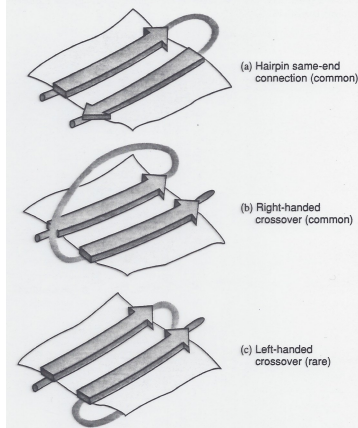
Protein Structure-Secondary

These strands are not straight as we saw, and this twist helps the conformation of these loops.

Figure 5.19

Summary

Three ways of making connections between β strands. (a) A hairpin same-end connection is commonly found for β strands in the antiparallel orientation. (b) A right-handed crossover connection is commonly found for β strands in the parallel orientation. (c) A left-handed crossover connection is rarely found.



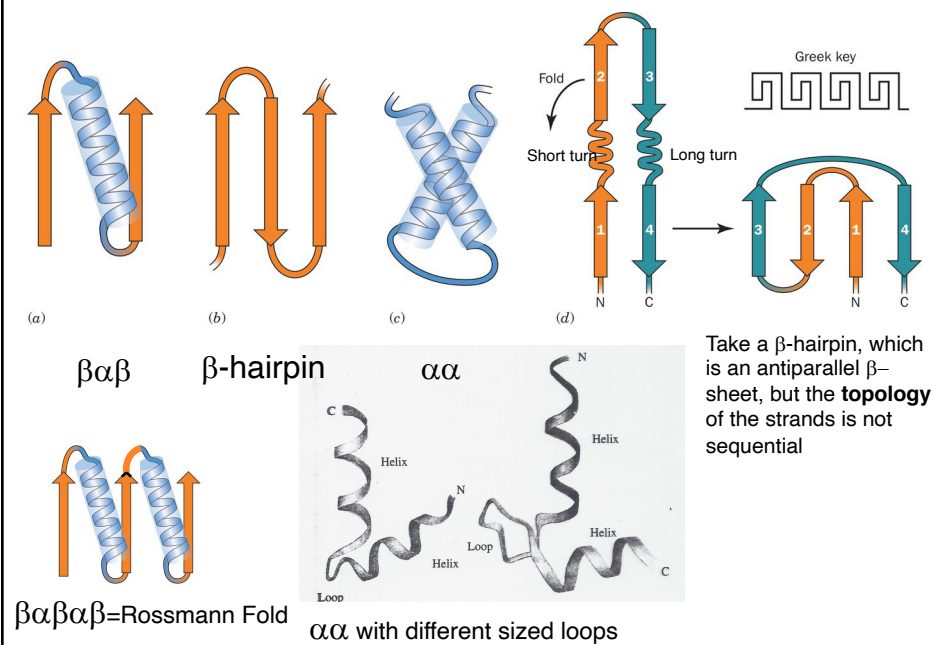
These loops are very often not just random conformations, but form α -helices

Protein Structure-Secondary

What is happening?

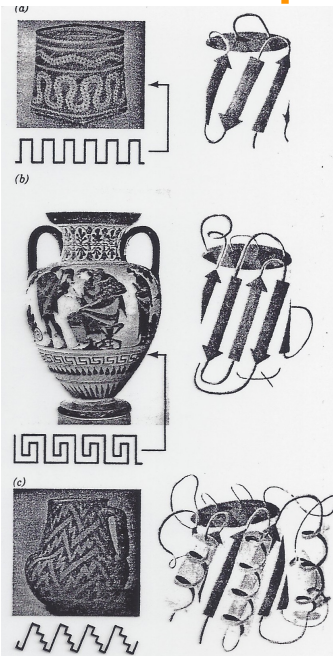
- Different pieces of 2° structure are mixing together.
- These are called “Motifs” or Super-secondary Structures
- What are the structures and names of some of the most common motifs?
 - $\beta\alpha\beta$
 - Rossmann Fold
 - β -hairpin
 - $\alpha\alpha$
 - Greek key
 - β -meander
 - β -barrel
 - $\alpha\beta$ -barrel

Protein Structure-Supersecondary



Protein Structure-Supersecondary

Due to the right-handed twist in the β -strands, as you add more strands the structure comes back on itself to form "barrels"



β -meander (β)₈

Greek Key (β)₈ (β -barrel)

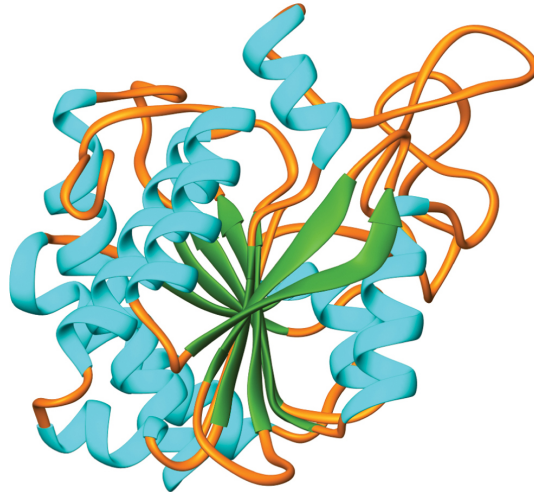
($\alpha\beta$)₈-Barrel

4 ($\beta\alpha\beta$) motifs
connected by 4 α -helices

FIGURE 7-49. Comparisons of the backbone folding patterns of protein β barrels (right) with geometric motifs commonly used to decorate Native American and Greek weaving and pottery (left). (a) Native American polychrome cane basket and the polypeptide backbone of rubredoxin from *Clostridium pasteurianum* showing its linked β meanders. [Museum of the American Indian, Heye Foundation.] (b) Red figured Greek amphora with its Greek key border area showing Cassandra and Ajax (about 450 B.C.) and the polypeptide backbone of human γ albumin with its "Greek key" pattern. [The Metropolitan Museum of Art, Fletcher Fund, 1956.] (c) Early Anasazi redware pitcher from New Mexico and the polypeptide backbone of chicken muscle triose phosphate isomerase showing its "lightning" pattern of overlapping β units. This so-called α/β barrel is also diagrammed in Fig. 7-19b. [Museum of the American Indian, Heye Foundation.] [After Richardson, J.S., *Nature* 268, 498 (1977).]

Tertiary Structure

Bovine Carboxypeptidase A

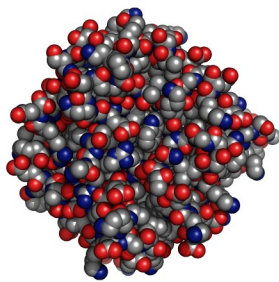


Based on an X-ray structure by William Lipscomb, Harvard University.
PDBid 3CPA.

Carboxypeptidase A
PDBid [3CPA](#)

Protein Structure-Tertiary

Picturing protein structure



(a)

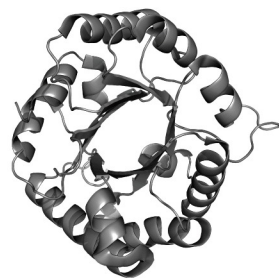
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Space-fill



(b)

Backbone trace



(c)

Ribbon

Triose-phosphate Isomerase (TIM)

137

Protein Structure-Tertiary

Picturing protein structure

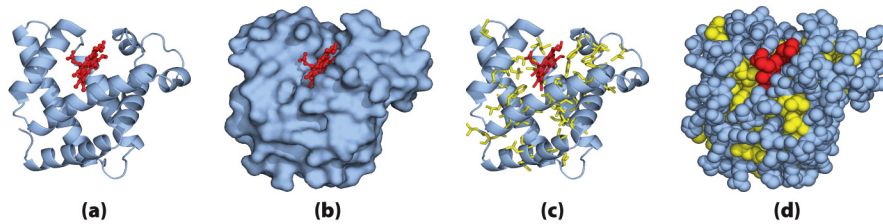


Figure 4-16
Lehninger Principles of Biochemistry, Seventh Edition
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Ribbon

Surface
contour

Ribbon with
side-chains

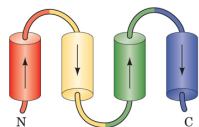
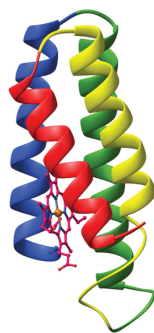
Space-
filling

Sperm-whale Myoglobin (Mb)

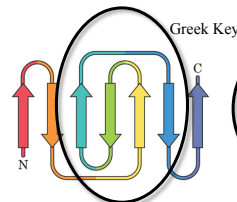
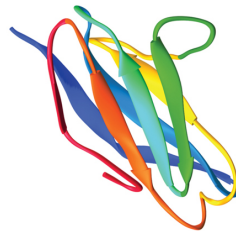
Protein Structure-Tertiary

Protein Classification:

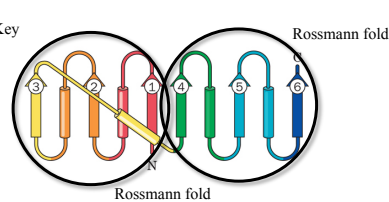
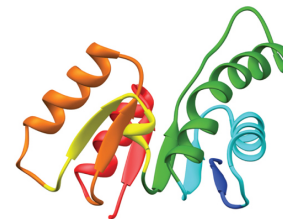
α , β , α/β , or barrels



Cytochrome *b*562
PDBid [256B](#)



Immunoglobulin-fold
Human immunoglobulin fragment
PDBid [7FAB](#)



Dogfish lactate dehydrogenase domain 2
PDBid [6LOH](#)