ORIGINAL PAPER

What's Behind the Decline? The Role of White Matter in Brain Aging

Jason D. Hinman · Carmela R. Abraham

Accepted: 23 March 2007/Published online: 20 April 2007 © Springer Science+Business Media, LLC 2007

Abstract The specific molecular events that underlie the age-related loss of cognitive function are poorly understood. Although not experimentally substantiated, age-dependent neuronal loss has long been considered central to age-related cognitive decline. More recently, age-related changes in brain white matter have taken precedence in explaining the steady decline in cognitive domains seen in non-diseased elderly. Characteristic alterations in the ultrastructure of myelin coupled with evidence of inflammatory processes present in the white matter of several different species suggest that specific molecular events within brain white matter may better explain observed pathological changes and cognitive deficits. This review focuses on recent evidence highlighting the importance of white matter in deciphering the course of "normal" brain aging.

Keywords Age-related cognitive decline · Myelin · Oligodendrocyte · Calpain · CNP

Abbreviations

ARCD	Age-related cognitive decline
AD	Alzheimer's disease
PD	Parkinson's disease
MRI	Magnetic resonance imaging
FA	Fractional anisotropy
R_2	Transverse relaxation rates
CNS	Central nervous system

Special issue in honor of Naren Banik.

J. D. Hinman · C. R. Abraham (🖂)

NFT	Neurofibrillary tangle
iNOS	Inducible nitric oxide synthase
MHC	Major histocompatibility complex
CAOs	Complement activated oligodendrocytes
C3aR	Complement C3a receptor
PNS	Peripheral nervous system
GFAP	Glial acidic fibrillary protein
ACT	α1-Antichymotrypsin
MBP	Myelin basic protein
PLP	Proteolipid protein
MAG	Myelin-associated glycoprotein
CNP	2',3' Cyclic nucleotide phosphodiesterase
MOSP	Myelin oligodendrocyte specific protein
EAE	Experimental allergic encephalomyelitis
MS	Multiple sclerosis

Age-related cognitive decline (ARCD)

Severe impairment of memory and executive function, otherwise known as dementia, can be the result of a number of pathologic processes, including several neurodegenerative diseases, the most prevalent of which is Alzheimer's disease (AD). In addition, even in the absence of a clear pathologic process, normal healthy adults also experience a steady decline in cognitive ability whether measured longitudinally [64] or cross-sectionally [83, 90]. This age-related cognitive decline (ARCD) is not evenly distributed amongst cognitive tasks, such that vocabulary and tasks requiring semantic knowledge as well as autobiographical and emotional memory remain relatively intact while processing speed, working memory, and learning decline. These studies are fraught with issues of individual variability, by necessity are correlational and

Departments of Biochemistry and Medicine, Boston University School of Medicine, 715 Albany Street, K620, Boston, MA 02118, USA e-mail: cabraham@bu.edu

potentially confounded by the considerable incidence of age-related diseases known to adversely affect cognitive performance [33]. Yet somewhat remarkably, a consistent decline in cognitive ability within certain domains is maintained from population to population [4, 11, 12, 17, 19, 64, 90].

Although the pathophysiology behind the major causes of dementia is comparatively well-understood, the biology underlying ARCD is only recently being elucidated. In most general terms, ARCD has been attributed to changes in prefrontal cortex and the integrity of anterior white matter circuits leading to non-selective recruitment of irrelevant brain regions in impaired individuals [33, 50]. Our group, as part of a larger program project, has focused on understanding the molecular changes underlying ARCD, using the rhesus monkey as a model, with the most intriguing findings being those changes present in brain white matter. It is our belief that age-related changes in white matter have not received adequate attention in the context of playing a potentially causative role in cognitive decline and hence, the discussion to follow will focus on the importance of changes in brain white matter, beginning with those at the gross level and culminating with those occurring at the molecular level, as key events in understanding ARCD.

Imaging age-related changes in brain volume

Magnetic resonance imaging (MRI) has provided a useful non-invasive tool for analyzing gross age-related changes in brain volume. In aged human subjects, several studies demonstrate a decline in the volume of brain white matter, particularly in frontal lobe areas [8, 30, 61, 76]. Recent advances in MRI technology allow for more accurate volumetric measurements by better distinguishing between gray and white matter as well as capitalizing on the unique features of compact white matter regions. The use of diffusion tensor MRI to measure fractional anisotropy (FA), the degree to which water molecules in a tissue are allowed to diffuse, is particularly useful in assessment of white matter tract integrity, as increased anisotropy correlates with markers of myelination [45, 101]. In healthy elderly adults, FA is reduced compared to young, particularly in the anterior or frontal regions of the brain including the corpus callosum [32, 61, 79, 80]. In a more elegant analysis, a group of behaviorally tested monkeys showing impaired executive function with age also show significant reductions in FA in the anterior corpus callosum, the superior longitudinal fasciculus II and the cingulum bundle; all corticocortical pathways vital for executive function [52]. Using a different methodology, by transformation of the T2 relaxation times into transverse relaxation rates (R_2), a more sensitive measure of brain myelination differences is generated [25] such that myelination increases R_2 while myelin breakdown decreases R_2 . In a large population (n = 252) of healthy adults ranging from 19 to 82 years of age, frontal lobe white matter R_2 shows a quadratic regression relationship with age, indicating that white matter volume increases until the fifth decade of life, after which it declines steadily [7, 8], implying that myelination is highly sensitive to time-dependent changes. These age-dependent decreases in white matter volume generally occur without significant change in gray matter volume [30].

The aged neuron: lost or diseased?

The importance of age-dependent neuronal loss is obvious and its potential role in ARCD should also be evident. As a result, many early studies, particularly prior to the advent of MRI, focused on neuronal counts and morphology changes, for the most part ignoring changes in glial cell number or morphology [13, 20, 35]. However, with the advent of precise stereologic methodology [98] and better preservation of brain tissue, studies reporting no loss of neurons with age began to appear [5, 16, 96]. Newer methods were designed to account for the fact that young brains tend to shrink more during preparation thus leading to a conclusion of greater neuronal density in young subjects. Accounting for the difference in brain shrinkage, volumetric estimates of gray and white matter in human post-mortem brain tissue suggest no significant loss of neurons with age and a significant age-related loss of up to 11% in white matter [31]. This paradigm shift was supported in part by work in the aged rhesus monkey, indicating changes in the myelinated axons of white matter might be more important to aging [68, 71]. Indeed, detailed counts of neurons from primary visual cortex (Brodmann area 17) and prefrontal cortex (area 46) in the monkey demonstrate there is no change in neuron number with age (Fig. 1) [67, 70]. Furthermore, in the hippocampus, the center of working memory, the number of CA1 pyramidal cell neurons remains stable across age [99]. One recent report indicated that in a cognitively relevant location in the frontal cortex (area 8A), focal neuronal death occurs with age [91], however, this report has yet to be confirmed. Overall, simple age-related neuronal loss is not an adequate suspect in explaining ARCD [26].

On the whole, other than in Layer I of the cortex, where neurons lose apical dendritic branches and occasionally synapses [62, 63], neuronal number and structure are largely unaffected by age. It is important to note that in prefrontal cortex area 46 of aged rhesus monkeys, the



Fig. 1 Lack of evidence for neuronal loss in the aged rhesus monkey. Comparative analysis of mean neuronal counts from 250 μ m strips of primary visual cortex (area 17) and prefrontal cortex (area 46) in monkeys of varying ages reveals no significant change in total neuronal number per unit volume. Adapted from Peters et al. [67, 70]

observed dendritic and synaptic loss correlate with the degree of cognitive impairment [74]. Furthermore, in the monkey, neurons do show age-related changes in their expression of neurotransmitters and receptors [78]. Recent microarray analysis of gene expression in area 46 in aged monkeys, indicates widespread transcriptional changes occur in neurons, including the degree to which they are susceptible to apoptosis [14]. Thus, while overall neuronal number may not change with age, various alterations in the health of neurons should be expected and cannot be discounted in any discussion of age-related changes in the central nervous system (CNS).

The contribution of amyloid burden

In humans, ~50% of those over the age of 85 develop profound memory impairment as a result of AD pathology, including the formation of senile amyloid plaques and neurofibrillary tangles (NFTs) [18]. The high incidence of AD in humans coupled with the virtual absence of AD-like pathology in wild-type laboratory animals such as the mouse, rat, and rhesus monkey, make it difficult to discern which age-related changes are associated with ARCD versus AD. While amyloid plaques can be found in rhesus monkeys, they are generally of the diffuse, non-pathogenic form and their distribution is markedly different from that seen in AD [34, 92]. Furthermore, plaque density in behaviorally characterized monkeys does not correlate with cognitive deficits [15, 89]. Additionally, NFTs are not typically found in aged monkeys and the degree of agerelated cognitive impairment is not consistent with AD [47]. For these reasons, AD-like phenomena have not been a focus of understanding mechanisms of ARCD and can be considered a disparate pathologic process.

Evidence of age-related white matter inflammation

In the monkey, the number of activated microglial cells, identified by their up-regulation of the major histocompatibility complex (MHC) class II proteins, increases dramatically with age specifically in regions of brain white matter and the increase correlates with cognitive impairment [87]. Similar increases in activated microglia have been noted in mice [56]. These phagocytic cells are known to produce a multitude of pro-inflammatory molecules [57], among them inducible nitric oxide synthase (iNOS). The resulting increase in nitric oxide presumably accounts for the concomitant increase in detection of nitrotyrosine residues in the white matter of aged monkeys [87]. Activated microglia are known to be active and phagocytic during phases of demyelination in the Long Evans shaker rat [103], illustrating the consequences of an increasing number of these cells with age. In the optic nerve of aged monkeys, increased numbers of microglia and hypertrophied astrocytes can be detected under the electron microscope, and the microglia cells appear to have phagocytosed myelin debris [81].

Interestingly, recent work in the rhesus monkey indicates that early components of the complement cascade (C3 and C4) can be found covalently bound to the myelin membrane in both young and old animals (J. A. Duce et al. 2006, submitted). This binding is associated with a significant increase in aged animals in the number of complement activated oligodendrocytes (CAOs), a pathological hallmark of white matter inflammation seen in multiple sclerosis (MS) [84] and other neurodegenerative diseases [102]. Complement component 3a (C3a) is a potent chemotactic factor and presumably as a result of robust expression of the C3a receptor (C3aR), activated microglia can be found in close association with these CAOs and in mice lacking C3aR expression, white matter inflammation is attenuated [10]. Thus, fixation of complement components to myelin may be a critical inflammatory trigger in the aging brain.

In addition to the activation of microglia, reactive astrocytosis can be found in white matter regions throughout the aged monkey brain, evidenced by an increased cell size of glial fibrillary acidic protein (GFAP)-positive cells [88]. An increase in GFAP mRNA and protein expression can also be detected in the brains of aged humans and rodents, though the total number of astrocytes does not change appreciably [27, 59]. Reactive astrocytes, like microglia, are known to produce abundant pro-inflammatory molecules including various proteases and inhibitors [22]. Of particular interest is the protease inhibitor α 1-antichymotrypsin (ACT), a serine protease inhibitor and acute phase protein. In AD, ACT expression is highly up-regulated at the RNA level [2] with reactive astrocytes being the major

source of ACT in the brain [1]. Microarray analysis of brain white matter from young and old rhesus monkeys reveals that ACT is also up-regulated during normal aging (Duce and Abraham, unpublished results).

Myelin changes in the aged brain

In part, based on the volume of evidence from gross pathology, imaging studies, and histologic assessments implicating changes in white matter as central to the mechanism of ARCD, a number of studies focus on microscopic examination of aged myelin. In the peripheral nervous system (PNS) of aged Fischer 344 rats, no change in the number of myelinated fibers was observed, though the ultrastructure of the myelin sheath, evidenced by ballooning and infolding, was disrupted with age [46]. Using histologic stains for myelin content [41], demonstrated diffuse myelin pallor throughout the human brain, particularly in the intracortical fibers thought to be responsible for high-level cognitive function. Tang et al. [95] and Pakkenberg and Gundersen [62] have both demonstrated a loss of white matter from the human brain with age, of up to 15%, as well as a decrease in the length of myelinated fibers. In a population of Danish individuals, the total length of myelinated fibers was seen to decrease by 10% per decade and 45% across the lifetime [54]. This decrease in fiber length indicates a much greater degree of myelin loss than indicated by studies of white matter volume.

Disruption of the myelin sheath, either by activated proinflammatory cells or by inappropriate protein maintenance within the oligodendrocyte (or a combination of both), has been described in aging rat [46, 93], monkey [66, 86], and human [3] brains. Four distinct phenomena are known to occur as myelin ages and these have been best described at the ultrastructural level as occurring throughout the CNS of the aged monkey (Fig. 2) [65]. First, localized splitting of the major denseline to accommodate dense cytoplasm from the oligodendrocyte can be seen in some sheaths (A). Second, myelin sheaths can balloon out, separating the intraperiod line, which then surrounds a fluid-filled space (B). Some thick myelin sheaths demonstrate a double sheath (C), with one compact set of lamellae surrounded by another. Finally, in both primates and rodents, so-called redundant myelin sheaths have been described [65, 69, 73] (D). These sheaths have far too many lamellae in proportion to the caliber of the axon. The significance of these ultrastructural changes on the function of the axon remains unclear, although the changes correlate with deficits in cognitive function in both primary visual cortex [69] and prefrontal cortex [72], suggesting that the disruption may interrupt normal axonal conduction and thus impair cognition.



Fig. 2 Age-related changes in myelin structure. There are four important features of age-related change in myelin ultrastructure [72]. These include, localized splitting of the MDL to accommodate dense cytoplasm of the oligodendrocyte (A), ballooning of myelin sheaths at the IPL forming a fluid-filled space (B), double myelin sheaths (C), and so-called "redundant" myelin sheaths (D). Adapted from Peters et al. [69, 73] and Peters and Sethares [72]

Recent studies have suggested that these age-related changes in myelin membranes alter axonal protein organization at or near the node of Ranvier. In both aged Fischer 344 rats and rhesus monkeys, voltage-gated potassium channels of the *Shaker* family (specifically Kv1.2), normally localized to the juxtaparanode beneath compact myelin, are seen to mislocalize into the paranodal region with age [37]. This reorganization occurs together with the presence of thick myelin sheaths with excessive paranodal loops of myelin. It is not clear from this study whether these changes are detrimental or restorative to axonal conduction, as the functionality of Kv1 channels in myelin is not well characterized. However it does demonstrate the ability of myelin to have retrograde effects on neuronal health.

The role of inappropriate protein degradation

Another common consequence of brain aging is the increased detection of ubiquitin. Particularly notable in agerelated neurodegenerative diseases, ubiquitin becomes associated with the NFT of AD, the Lewy body of Parkinson's disease (PD), the inclusions of amyotrophic lateral sclerosis, etc. [51]. In fact, one of the genes associated with autosomal recessive juvenile PD, parkin, codes for an enzyme in the ubiquitin pathway [44, 85]. Ubiquitin, a small, highly conserved 8 kDa protein modifier is attached to proteins destined for degradation by the proteasomal pathway (reviewed by Fang and Weissman [23]). Likewise, ubiquitin can be removed by the action of deubiquitinases and the addition of ubiquitin may serve multiple cellular functions besides protein degradation [29].

In the aging brain, immunohistochemical staining for ubiquitin reveals localization to myelin and lysosome-related bodies in neurons [51]. Using the electron microscope together with immunolabeling, age-related ubiquitin deposits in humans and dogs can be seen within focal enlargements of myelin sheaths [55]. Likewise [21], report dense ubiquitin-positive inclusions within glial cells and ubiquitin staining in swollen myelin sheaths as well as documenting significantly more ubiquitin immunoreactivity in white matter than in gray. In aged dogs, white matter degeneration is marked by deposits of non-degraded ubiquitin-positive conjugates in myelin [24]. Interestingly, though ubiquitin conjugation serves primarily to target proteins for degradation, ubiquitin-protein conjugates are consistently found in the aged brain and in neurodegenerative diseases, suggesting a failure of proteasomal function with age, leaving large non-degraded, and potentially nonfunctional protein complexes to accumulate. The toxicity of these complexes may be particularly relevant in white matter and myelin, where the normal rate of protein turnover is already considerably lower than that seen in whole brain [48].

Age-related changes in myelin biochemistry

The ultimate effect of age-related increases in white matter inflammation and myelin disruption must trickle down to observable alterations in myelin lipids and protein expression. On this topic, a number of reports spanning several decades exist, demonstrating evidence of both agerelated increases and decreases in various proteins and lipids that comprise myelin. At this time, the lack of wellcontrolled studies prevents a thorough discussion of agerelated changes in myelin lipids. It is essential to add, however that sulfatide-null mice demonstrate an increasing prevalence of redundant, uncompacted or degenerating myelin as well as alterations in nodal and paranodal ion channel localization as they age [38, 53]. These changes are very similar to changes observed in aged rhesus monkey myelin ultrastructure and axonal reorganization. There has been no focused analysis of changes in CNS sulfatide over time, though in a global analysis of myelin lipids sulfatide was among the myelin components that were

decreased in frontal and temporal white matter regions in a cohort of human brains aged 20-100 [94].

With respect to total amounts of myelin, measured either with histologic stains or by various purification methods, there is a consensus of significant myelin loss over time [9, 86, 100]. A majority of these older studies indicate no change in the individual protein composition of myelin but were carried out prior to clarification of the myelin proteome [9, 100]. Myelin basic protein (MBP) has shown to be decreased in elderly with no cognitive impairment (NCI) [6, 97] as well as in AD [77], though in a study of behaviorally characterized rhesus monkeys, MBP, proteolipid protein (PLP), and myelin-associated glycoprotein (MAG) were unchanged while the levels of myelin oligodendrocyte specific protein (MOSP) and 2''3'-cyclic nucleotide phosphodiesterase (CNP) were increased with age [86].

In addition to examining the age-related changes in the total amount of myelin proteins, exploring what posttranslational modifications occur to these proteins with age is equally important. As an example, MBP isolated from MS human brain and demyelinating mouse models indicate that multiple post-translational modifications, including deimination (resulting in citrullinated proteins) and dephosphorylation, may be essential to altering myelin structure and function [42] and are commonly observed in age-related neurodegenerative disease [39, 58]. In the case of CNP, there is evidence from a cohort of aged rhesus monkeys that CNP accumulation is related to the increased detection of ubiquitinated species of CNP in myelin from aged animals (Hinman, Chen, Hollander, and Abraham in preparation). Additionally, both CNP and MOSP showed evidence of limited proteolysis, which in the case of CNP was attributable to an age-dependent increase in activated calpain-1 [36, 86]. This calpain-mediated limited proteolysis of CNP can also be observed in vitro by incubation of white matter homogenates from a young monkey prepared in the absence of protease inhibitors. The degradation is enhanced by the addition of 500 µM calcium and can be inhibited by a specific calpain inhibitor (Fig. 3a). Interestingly, the appearance of the major 40 kDa proteolytic fragment of CNP appears to correlate with age in a limited sample of aged rhesus monkeys (Fig. 3b). Already implicated in myelin breakdown in MS and its animal model, experimental allergic encephalomyelitis (EAE) [82], the exact proteolytic mechanism of calpain in the degradation of myelin proteins has yet to be determined, however its role in limited proteolysis is likely essential to understanding myelin breakdown. Further work is necessary to understand the potential significance of limited proteolytic fragments of CNP and other myelin proteins such as MOSP.



Fig. 3 Age-dependent limited proteolysis of CNP by calpain-1. Incubation of TX-100 soluble homogenate from a 9-year-old (young) monkey for 24 h at 37°C in the presence of TBS (lane 3) results in the same pattern of CNP proteolysis as that seen in a 26-year-old monkey at time zero (lane 13 from *left*; *arrow* indicates major 40 kDa CNP fragment). This pattern can be induced at time zero by the addition of 500 μ M Ca⁺⁺ (lane 4) and abolished by the addition

of 5 mM EGTA (lanes 7–9) or 0.35 U of calpastatin (lanes 10–12) (A). Long exposures of immunoblots for CNP in TX-100 soluble fractions of temporal lobe subcortical white matter reveals the presence of a limited proteolytic fragment of CNP migrating at ~40 kDa. Quantitative assessment of the level of this major 40 kDa fragment observed in aged monkeys demonstrates a statistically significant correlation with age throughout brain white matter (**B**)

The apparent paradox of simultaneous protein accumulation and increased proteolysis can be explained by considering the relationship between failed proteasomal degradation and the recruitment of alternative proteolytic mechanisms such as calpain. This relationship has been described in the context of P-glycoprotein and p107 such that both ubiquitination and calpain activation are necessary for the degradation of the target molecules [40, 60]. In the nervous system, the parkin gene codes for a ubiquitin ligase which appears to induce calpain-mediated degradation of alpha-synuclein suggesting further that cytotoxic states such as alpha-synuclein accumulation is a stimulus for cooperation between ubiquitin and calpain-mediated proteolysis [43].

Changes such as those described in the rhesus monkey, occurring in critical lower abundance myelin proteins (e.g., CNP and MOSP), may be triggered by the activation of inflammatory processes and underlie myelin disruption. Interestingly, mice overexpressing CNP show evidence of redundant myelin and intramyelinic vacuoles similar to those seen in aged monkeys [28], while mice lacking CNP show an age-dependent reactive gliosis and disorganized paranodal profiles [49, 75], features also noted in aged monkeys. While the precise role of CNP in myelin has yet to be determined, these similarities argue strongly for a prominent role for CNP (and the microtubule cytoskeleton) in the long-term maintenance of the myelin membrane.

Summary

The study of ARCD has matured recently, moving from a focus on age-related neuronal loss as being directly causative, to a greater understanding of the contribution of molecular changes in brain white matter. In recent years, research of this nature has shifted focus from characterizing gross or morphologic changes with age to understanding what molecular events underlie age-related white matter inflammation and myelin membrane disruption. Considering the abundance of evidence from both humans, rodents, as well as the monkey, declines in white matter integrity appear at least as important as neuronal changes in understanding ARCD. In particular, age-related changes in myelin protein degradation and the recruitment of

alternative mechanisms of proteolysis appear as principal events in the brain aging process, occurring in advance of age-related changes in neurons. This evidence should help to guide future research in brain aging and influence potential interventional targets.

Acknowledgments The authors thank our fellow collaborators in the Aging Program Project (NIA AG00001) particularly Drs. Peters, Rosene, Moss, Leubke, Hollander, Duce, and Chen.

References

- Abraham CR (2001) Reactive astrocytes and alpha1-antichymotrypsin in Alzheimer's disease. Neurobiol Aging 22(6):931– 936
- Abraham CR, Selkoe DJ, Potter H (1988) Immunochemical identification of the serine protease inhibitor alpha 1-antichymotrypsin in the brain amyloid deposits of Alzheimer's disease. Cell 52(4):487–501
- 3. Albert M (1993a) Neuropsychological and neurophysiological changes in healthy adult humans across the age range. Neurobiol Aging 14(6):623–625
- Albert MS (1993b) Neuropsychological and neurophysiological changes in healthy adult humans across the age range. Neurobiology Aging 14:623–625
- Anderson JM, Hubbard BM, Coghill GR, Slidders W (1983) The effect of advanced old age on the neurone content of the cerebral cortex. Observations with an automatic image analyser point counting method. J Neurol Sci 58(2):235–246
- Ansari KA, Loch J (1975) Decreased myelin basic protein content of the aged human brain. Neurology 25(11):1045–1050
- Bartzokis G, Beckson M, Lu PH, Nuechterlein KH, Edwards N, Mintz J (2001) Age-related changes in frontal and temporal lobe volumes in men: a magnetic resonance imaging study. Arch Gen Psychiatry 58(5):461–465
- Bartzokis G, Cummings JL, Sultzer D, Henderson VW, Nuechterlein KH, Mintz J (2003) White matter structural integrity in healthy aging adults and patients with Alzheimer disease: a magnetic resonance imaging study. Arch Neurol 60(3):393–398
- Berlet HH, Volk B (1980) Studies of human myelin proteins during old age. Mech Ageing Dev 14(1–2):211–222
- Boos L, Campbell IL, Ames R, Wetsel RA, Barnum SR (2004) Deletion of the complement anaphylatoxin C3a receptor attenuates, whereas ectopic expression of C3a in the brain exacerbates, experimental autoimmune encephalomyelitis. J Immunol 173(7):4708–4714
- Brayne C, Gill C, Paykel ES, Huppert F, O'Connor DW (1995) Cognitive decline in an elderly population–a two wave study of change. Psychol Med 25(4):673–683
- Brayne C, Spiegelhalter DJ, Dufouil C, Chi LY, Dening TR, Paykel ES, O'Connor DW, Ahmed A, McGee MA, Huppert FA (1999) Estimating the true extent of cognitive decline in the old old. J Am Geriatr Soc 47(11):1283–1288
- Brody H (1955) Organization of the cerebral cortex. III. A study of aging in the human cerebral cortex. J Comp Neurol 102(2):511–516
- Chen C, Duce JA, Hollander W, Rosene DL, Abraham CR Agerelated changes in gene expression in area 46 of the aged rhesus monkey (in preparation)
- Cork LC (1993) Plaques in prefrontal cortex of aged, behaviorally-tested rhesus monkeys: incidence, distribution, and relationship to task performance. Neurobiol Aging 14(6):675– 676

- Cragg BG (1975) The density of synapses and neurons in normal, mentally defective ageing human brains. Brain 98(1):81–90
- Cullum S, Huppert FA, McGee M, Dening T, Ahmed A, Paykel ES, Brayne C (2000) Decline across different domains of cognitive function in normal ageing: results of a longitudinal population-based study using CAMCOG. Int J Geriatr Psychiatry 15(9):853–862
- Cummings JL (2004) Alzheimer's disease. N Engl J Med 351(1):56–67
- Deary IJ, Leaper SA, Murray AD, Staff RT, Whalley LJ (2003) Cerebral white matter abnormalities and lifetime cognitive change: a 67-year follow-up of the Scottish Mental Survey of 1932. Psychol Aging 18(1):140–148
- Devaney KO, Johnson HA (1980) Neuron loss in the aging visual cortex of man. J Gerontol 35(6):836–841
- Dickson DW, Wertkin A, Kress Y, Ksiezak-Reding H, Yen SH (1990) Ubiquitin immunoreactive structures in normal human brains. Distribution and developmental aspects. Lab Invest 63(1):87–99
- Eddleston M, Mucke L (1993) Molecular profile of reactive astrocytes-implications for their role in neurologic disease. Neuroscience 54(1):15–36
- Fang S, Weissman AM (2004) A field guide to ubiquitylation. Cell Mol Life Sci 61(13):1546–1561
- Ferrer I, Pumarola M, Rivera R, Zujar MJ, Cruz-Sanchez F, Vidal A (1993) Primary central white matter degeneration in old dogs. Acta Neuropathol (Berl) 86(2):172–175
- 25. Ferrie JC, Barantin L, Saliba E, Akoka S, Tranquart F, Sirinelli D, Pourcelot L (1999) MR assessment of the brain maturation during the perinatal period: quantitative T2 MR study in premature newborns. Magn Reson Imaging 17(9):1275–1288
- Finch CE (2003) Neurons, glia, and plasticity in normal brain aging. Neurobiol Aging 24(Suppl 1):S123–S127; Discussion S131
- Goss JR, Finch CE, Morgan DG (1991) Age-related changes in glial fibrillary acidic protein mRNA in the mouse brain. Neurobiol Aging 12(2):165–170
- Gravel M, Peterson J, Yong VW, Kottis V, Trapp B, Braun PE (1996) Overexpression of 2',3'-cyclic nucleotide 3'-phosphodiesterase in transgenic mice alters oligodendrocyte development and produces aberrant myelination. Mol Cell Neurosci 7(6):453–466
- Guterman A, Glickman MH (2004) Deubiquitinating enzymes are IN/(trinsic to proteasome function). Curr Protein Pept Sci 5(3):201–211
- Guttmann CR, Jolesz FA, Kikinis R, Killiany RJ, Moss MB, Sandor T, Albert MS (1998) White matter changes with normal aging. Neurology 50(4):972–978
- Haug H, Eggers R (1991) Morphometry of the human cortex cerebri and corpus striatum during aging. Neurobiol Aging 12(4):336–338; Discussion 352–335
- 32. Head D, Buckner RL, Shimony JS, Williams LE, Akbudak E, Conturo TE, McAvoy M, Morris JC, Snyder AZ (2004) Differential vulnerability of anterior white matter in nondemented aging with minimal acceleration in dementia of the Alzheimer type: evidence from diffusion tensor imaging. Cereb Cortex 14(4):410–423
- Hedden T, Gabrieli JD (2004) Insights into the ageing mind: a view from cognitive neuroscience. Nat Rev Neurosci 5(2):87–96
- Heilbroner PL, Kemper TL (1990) The cytoarchitectonic distribution of senile plaques in three aged monkeys. Acta Neuropathol (Berl) 81(1):60–65
- Henderson G, Tomlinson BE, Gibson PH (1980) Cell counts in human cerebral cortex in normal adults throughout life using an image analysing computer. J Neurol Sci 46(1):113–136

- 36. Hinman JD, Duce JA, Siman RA, Hollander W, Abraham CR (2004) Activation of calpain-1 in myelin and microglia in the white matter of the aged rhesus monkey. J Neurochem 89(2):430–441
- Hinman JD, Peters A, Cabral H, Rosene DL, Hollander W, Rasband MN, Abraham CR (2006) Age-related molecular reorganization at the node of Ranvier. J Comp Neurol 495(4):351–362
- 38. Ishibashi T, Dupree JL, Ikenaka K, Hirahara Y, Honke K, Peles E, Popko B, Suzuki K, Nishino H, Baba H (2002) A myelin galactolipid, sulfatide, is essential for maintenance of ion channels on myelinated axon but not essential for initial cluster formation. J Neurosci 22(15):6507–6514
- 39. Ishigami A, Ohsawa T, Hiratsuka M, Taguchi H, Kobayashi S, Saito Y, Murayama S, Asaga H, Toda T, Kimura N, Maruyama N (2005) Abnormal accumulation of citrullinated proteins catalyzed by peptidylarginine deiminase in hippocampal extracts from patients with Alzheimer's disease. J Neurosci Res 80(1):120–128
- 40. Jang JS, Choi YH (1999) Proteolytic degradation of the retinoblastoma family protein p107: a putative cooperative role of calpain and proteasome. Int J Mol Med 4(5):487–492
- Kemper TL (1994) Neuroanatomical and neuropathological changes during aging and dementia. In: Alber ML, Knoefel JE (eds) Clinical neurology and aging. Oxford University Press, New York, Oxford, pp 3–67
- 42. Kim JK, Mastronardi FG, Wood DD, Lubman DM, Zand R, Moscarello MA (2003a) Multiple sclerosis: an important role for post-translational modifications of myelin basic protein in pathogenesis. Mol Cell Proteomics 2(7):453–462
- 43. Kim SJ, Sung JY, Um JW, Hattori N, Mizuno Y, Tanaka K, Paik SR, Kim J, Chung KC (2003b) Parkin cleaves intracellular alpha-synuclein inclusions via the activation of calpain. J Biol Chem 278(43):41890–41899
- 44. Kitada T, Asakawa S, Hattori N, Matsumine H, Yamamura Y, Minoshima S, Yokochi M, Mizuno Y, Shimizu N (1998) Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. Nature 392(6676):605–608
- 45. Klingberg T, Vaidya CJ, Gabrieli JD, Moseley ME, Hedehus M (1999) Myelination and organization of the frontal white matter in children: a diffusion tensor MRI study. Neuroreport 10(13):2817–2821
- 46. Knox CA, Kokmen E, Dyck PJ (1989) Morphometric alteration of rat myelinated fibers with aging. J Neuropathol Exp Neurol 48(2):119–139
- 47. Lai ZC, Moss MB, Killiany RJ, Rosene DL, Herndon JG (1995) Executive system dysfunction in the aged monkey: spatial and object reversal learning. Neurobiol Aging 16(6):947–954
- Lajtha A, Toth J, Fujimoto K, Agrawal HC (1977) Turnover of myelin proteins in mouse brain in vivo. Biochem J 164(2):323– 329
- 49. Lappe-Siefke C, Goebbels S, Gravel M, Nicksch E, Lee J, Braun PE, Griffiths IR, Nave KA (2003) Disruption of Cnp1 uncouples oligodendroglial functions in axonal support and myelination. Nat Genet 33(3):366–374
- Logan JM, Sanders AL, Snyder AZ, Morris JC, Buckner RL (2002) Under-recruitment and nonselective recruitment: dissociable neural mechanisms associated with aging. Neuron 33(5):827–840
- 51. Lowe J, Mayer RJ, Landon M (1993) Ubiquitin in neurodegenerative diseases. Brain Pathol 3(1):55–65
- 52. Makris N, Papadimitriou GM, van der Kouwe A, Kennedy DN, Hodge SM, Dale AM, Benner T, Wald LL, Wu O, Tuch DS, Caviness VS, Moore TL, Killiany RJ, Moss MB, Rosene DL (2006) Frontal connections and cognitive changes in normal

aging rhesus monkeys: a DTI study. Neurobiol Aging 2006, Sep 6; [Epub ahead of print]

- 53. Marcus J, Honigbaum S, Shroff S, Honke K, Rosenbluth J, Dupree JL (2006) Sulfatide is essential for the maintenance of CNS myelin and axon structure. Glia 53(4):372–381
- Marner L, Nyengaard JR, Tang Y, Pakkenberg B (2003) Marked loss of myelinated nerve fibers in the human brain with age. J Comp Neurol 462(2):144–152
- 55. Migheli A, Attanasio A, Pezzulo T, Gullotta F, Giordana MT, Schiffer D (1992) Age-related ubiquitin deposits in dystrophic neurites: an immunoelectron microscopic study. Neuropathol Appl Neurobiol 18(1):3–11
- Mouton PR, Long JM, Lei DL, Howard V, Jucker M, Calhoun ME, Ingram DK (2002) Age and gender effects on microglia and astrocyte numbers in brains of mice. Brain Res 956(1):30–35
- Nakanishi H (2003) Microglial functions and proteases. Mol Neurobiol 27(2):163–176
- Nicholas AP, Sambandam T, Echols JD, Tourtellotte WW (2004) Increased citrullinated glial fibrillary acidic protein in secondary progressive multiple sclerosis. J Comp Neurol 473(1):128–136
- 59. Nichols NR, Day JR, Laping NJ, Johnson SA, Finch CE (1993) GFAP mRNA increases with age in rat and human brain. Neurobiol Aging 14(5):421–429
- 60. Ohkawa K, Asakura T, Takada K, Sawai T, Hashizume Y, Okawa Y, Yanaihara N (1999) Calpain inhibitor causes accumulation of ubiquitinated P-glycoprotein at the cell surface: possible role of calpain in P-glycoprotein turnover. Int J Oncol 15(4):677–686
- 61. O'Sullivan M, Jones DK, Summers PE, Morris RG, Williams SC, Markus HS (2001) Evidence for cortical "disconnection" as a mechanism of age-related cognitive decline. Neurology 57(4):632–638
- Pakkenberg B, Gundersen HJ (1997) Neocortical neuron number in humans: effect of sex and age. J Comp Neurol 384(2):312– 320
- 63. Pakkenberg B, Pelvig D, Marner L, Bundgaard MJ, Gundersen HJ, Nyengaard JR, Regeur L (2003) Aging and the human neocortex. Exp Gerontol 38(1–2):95–99
- 64. Park DC, Smith AD, Lautenschlager G, Earles JL, Frieske D, Zwahr M, Gaines CL (1996) Mediators of long-term memory performance across the life span. Psychol Aging 11(4):621–637
- 65. Peters A (2002a) The effects of normal aging on myelin and nerve fibers: a review. J Neurocytol 31(8–9):581–593
- 66. Peters A (2002b) Structural changes in the normally aging cerebral cortex of primates. Prog Brain Res 136:455–465
- Peters A, Leahu D, Moss MB, McNally KJ (1994) The effects of aging on area 46 of the frontal cortex of the rhesus monkey. Cereb Cortex 4(6):621–635
- Peters A, Morrison JH, Rosene DL, Hyman BT (1998a) Feature article: are neurons lost from the primate cerebral cortex during normal aging? Cereb Cortex 8(4):295–300
- Peters A, Moss MB, Sethares C (2000) Effects of aging on myelinated nerve fibers in monkey primary visual cortex. J Comp Neurol 419(3):364–376
- Peters A, Nigro NJ, McNally KJ (1997) A further evaluation of the effect of age on striate cortex of the rhesus monkey. Neurobiol Aging 18(1):29–36
- Peters A, Rosene DL, Moss MB, Kemper TL, Abraham CR, Tigges J, Albert MS (1996) Neurobiological bases of age-related cognitive decline in the rhesus monkey. J Neuropathol Exp Neurol 55(8):861–874
- 72. Peters A, Sethares C (2002) Aging and the myelinated fibers in prefrontal cortex and corpus callosum of the monkey. J Comp Neurol 442(3):277–291

- Peters A, Sethares C, Killiany RJ (2001) Effects of age on the thickness of myelin sheaths in monkey primary visual cortex. J Comp Neurol 435(2):241–248
- 74. Peters A, Sethares C, Moss MB (1998b) The effects of aging on layer 1 in area 46 of prefrontal cortex in the rhesus monkey. Cereb Cortex 8(8):671–684
- Rasband MN, Tayler J, Kaga Y, Yang Y, Lappe-Siefke C, Nave KA, Bansal R (2005) CNP is required for maintenance of axonglia interactions at nodes of Ranvier in the CNS. Glia 50(1):86– 90
- Resnick SM, Pham DL, Kraut MA, Zonderman AB, Davatzikos C (2003) Longitudinal magnetic resonance imaging studies of older adults: a shrinking brain. J Neurosci 23(8):3295–3301
- 77. Roher AE, Weiss N, Kokjohn TA, Kuo YM, Kalback W, Anthony J, Watson D, Luehrs DC, Sue L, Walker D, Emmerling M, Goux W, Beach T (2002) Increased A beta peptides and reduced cholesterol and myelin proteins characterize white matter degeneration in Alzheimer's disease. Biochemistry 41(37):11080–11090
- Rosene DL, Nicholson TJ (1999) Neurotransmitter receptor changes in the hippocampus and cerebral cortex in normal aging. Cereb Cortex 14:111–128
- 79. Salat DH, Tuch DS, Greve DN, van der Kouwe AJ, Hevelone ND, Zaleta AK, Rosen BR, Fischl B, Corkin S, Rosas HD, Dale AM (2005a) Age-related alterations in white matter micro-structure measured by diffusion tensor imaging. Neurobiol Aging 26(8):1215–1227
- Salat DH, Tuch DS, Hevelone ND, Fischl B, Corkin S, Rosas HD, Dale AM (2005b) Age-related changes in prefrontal white matter measured by diffusion tensor imaging. Ann NY Acad Sci 1064:37–49
- Sandell JH, Peters A (2002) Effects of age on the glial cells in the rhesus monkey optic nerve. J Comp Neurol 445(1):13–28
- Schaecher KE, Shields DC, Banik NL (2001) Mechanism of myelin breakdown in experimental demyelination: a putative role for calpain. Neurochem Res 26(6):731–737
- Schaie KW (1996) Intellectual development in adulthood: the Seattle longitudinal study. Cambridge University Press, Cambridge
- 84. Schwab C, McGeer PL (2002) Complement activated C4d immunoreactive oligodendrocytes delineate small cortical plaques in multiple sclerosis. Exp Neurol 174(1):81–88
- 85. Shimura H, Hattori N, Kubo S, Mizuno Y, Asakawa S, Minoshima S, Shimizu N, Iwai K, Chiba T, Tanaka K, Suzuki T (2000) Familial Parkinson disease gene product, parkin, is a ubiquitin-protein ligase. Nat Genet 25(3):302–305
- 86. Sloane JA, Hinman JD, Lubonia M, Hollander W, Abraham CR (2003) Age-dependent myelin degeneration and proteolysis of oligodendrocyte proteins is associated with the activation of calpain-1 in the rhesus monkey. J Neurochem 84(1):157–168
- Sloane JA, Hollander W, Moss MB, Rosene DL, Abraham CR (1999) Increased microglial activation and protein nitration in white matter of the aging monkey. Neurobiol Aging 20(4):395– 405

- 88. Sloane JA, Hollander W, Rosene DL, Moss MB, Kemper T, Abraham CR (2000) Astrocytic hypertrophy and altered GFAP degradation with age in subcortical white matter of the rhesus monkey. Brain Res 862(1–2):1–10
- 89. Sloane JA, Pietropaolo MF, Rosene DL, Moss MB, Peters A, Kemper T, Abraham CR (1997) Lack of correlation between plaque burden and cognition in the aged monkey. Acta Neuropathol (Berl) 94(5):471–478
- 90. Small SA, Stern Y, Tang M, Mayeux R (1999) Selective decline in memory function among healthy elderly. Neurology 52(7):1392–1396
- 91. Smith DE, Rapp PR, McKay HM, Roberts JA, Tuszynski MH (2004) Memory impairment in aged primates is associated with focal death of cortical neurons and atrophy of subcortical neurons. J Neurosci 24(18):4373–4381
- 92. Struble RG, Price DL Jr, Cork LC, Price DL (1985) Senile plaques in cortex of aged normal monkeys. Brain Res 361(1– 2):267–275
- 93. Sugiyama I, Tanaka K, Akita M, Yoshida K, Kawase T, Asou H (2002) Ultrastructural analysis of the paranodal junction of myelinated fibers in 31-month-old-rats. J Neurosci Res 70(3):309–317
- 94. Svennerholm L, Bostrom K, Jungbjer B, Olsson L (1994) Membrane lipids of adult human brain: lipid composition of frontal and temporal lobe in subjects of age 20 to 100 years. J Neurochem 63(5):1802–1811
- 95. Tang Y, Nyengaard JR, Pakkenberg B, Gundersen HJ (1997) Age-induced white matter changes in the human brain: a stereological investigation. Neurobiol Aging 18(6):609–615
- Terry RD, DeTeresa R, Hansen LA (1987) Neocortical cell counts in normal human adult aging. Ann Neurol 21(6):530–539
- 97. Wang DS, Bennett DA, Mufson EJ, Mattila P, Cochran E, Dickson DW (2004) Contribution of changes in ubiquitin and myelin basic protein to age-related cognitive decline. Neurosci Res 48(1):93–100
- West MJ (1993a) New stereological methods for counting neurons. Neurobiol Aging 14(4):275–285
- West MJ (1993b) Regionally specific loss of neurons in the aging human hippocampus. Neurobiol Aging 14(4):287–293
- 100. Wiggins RC, Gorman A, Rolsten C, Samorajski T, Ballinger WE Jr, Freund G (1988) Effects of aging and alcohol on the biochemical composition of histologically normal human brain. Metab Brain Dis 3(1):67–80
- 101. Wimberger DM, Roberts TP, Barkovich AJ, Prayer LM, Moseley ME, Kucharczyk J (1995) Identification of "premyelination" by diffusion-weighted MRI. J Comput Assist Tomogr 19(1):28–33
- 102. Yamada T, Akiyama H, McGeer PL (1990) Complement-activated oligodendroglia: a new pathogenic entity identified by immunostaining with antibodies to human complement proteins C3d and C4d. Neurosci Lett 112(2–3):161–166
- 103. Zhang SC, Goetz BD, Carre JL, Duncan ID (2001) Reactive microglia in dysmyelination and demyelination. Glia 34(2):101– 109