

Lecture 25 (11/16/20) Nucleic Acids

TODAY

- Reading: Ch25; 990-995, 1005-1012
Ch26; 1035-1038
Ch27; 1077-1085, 1092-1096
- Problems: Ch25 (text); 1-3,5-7,10,13-16,12
Ch25 (study-guide: applying); 1,4
Ch25 (study-guide: facts); 3,4,6
Ch26 (text); 1,2,5,6,12
Ch26 (study-guide: applying); 1
Ch26 (study-guide: facts); 1,3,5
Ch27 (text); 6,7,9
Ch27 (study-guide: applying); 1,3,5

NEXT

- Reading: Ch27; 1088-1091, 1096-1108
- Problems: Ch27 (text); 5,8,10,11,13,16,17
Ch27 (study-guide: applying); 2,3
Ch27 (study-guide: facts); 4,6

- A. The 4 S's of Nucleotides & NA
- B. Structure of the Information
- C. Recombinant DNA: Biochemical Basis of Biotechnology
 1. Restriction enzymes, DNA ligase
 2. Vectors and Inserts to make recombinant DNA (rDNA)
 3. Transformation of hosts
 4. Selection of transformants
 5. Expression
 6. Site-directed mutagenesis
- D. Replication
 1. Polymerases
 2. Fidelity
 - a. Polymerase recognition
 - b. Exonuclease
 - c. Mis-match repair
 - d. Post-replication repair
 - i. Direct reversal
 - ii. Base excision
 - iii. Nucleotide excision
 3. Sequence determination
 4. PCR
- E. Transcription
 1. RNA polymerase
 2. fidelity
- F. Translation
 1. Genetic code
 2. tRNA

DNA Replication

- DNA Replication
 - Polymerases
 - Activities
 - Structure
 - Mechanism
- Replication/polymerase fidelity
 - Exonuclease Activities
 - Mismatch repair
 - Post-replication repair
 - Reversal
 - Base Excision repair
 - Nucleotide Excision repair
- DNA Sequencing
- PCR

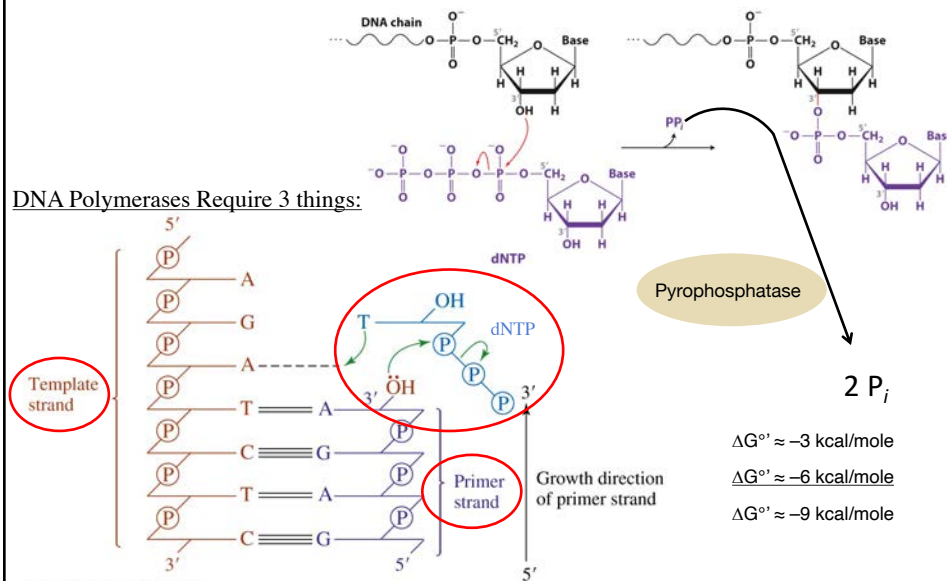
DNA Replication

Nucleic acid function: Central Dogma



DNA Replication

Addition at 3' end



Comparison of Polymerases

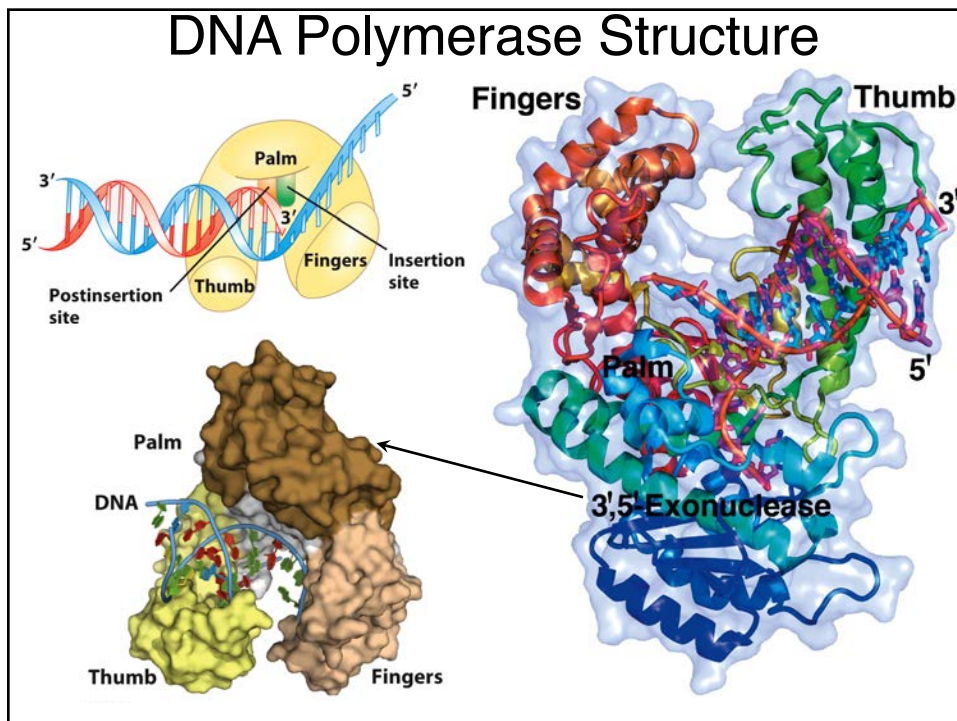
TABLE 25-1 Properties of *E. coli* DNA Polymerases

	Pol I	Pol II	Pol III
Mass (kD)	103	90 (α_4)	130 *
Molecules/cell	400	?	10–20
Turnover number ^a	20	5	1000
Structural gene	<i>polA</i>	<i>polB</i>	<i>polC</i>
Conditionally lethal mutant	+	–	+
Polymerization: 5' → 3'	+	+	+
? → Exonuclease: 3' → 5'	+	+	+
? → Exonuclease: 5' → 3'	+	–	–
Processivity	100	10,000	500,000 ^b

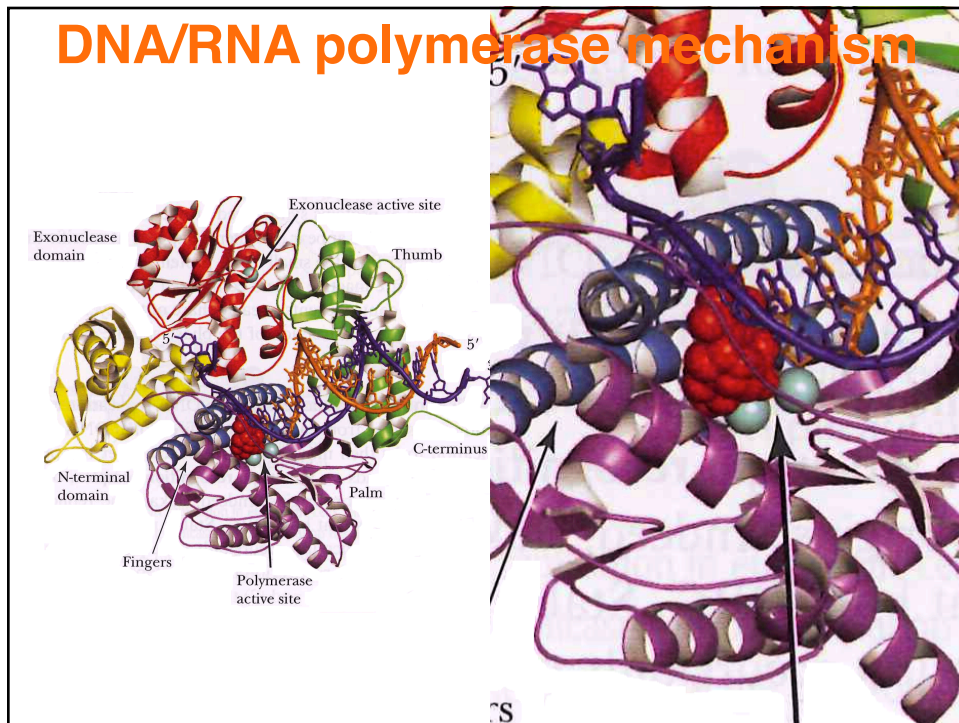
^adNTP polymerized sec⁻¹ at 37 °C.

^bnot including Okasaki fragments

*In a complex with 10 proteins (26 subunits) of >900 kD. Core is trimer of α , θ , ϵ



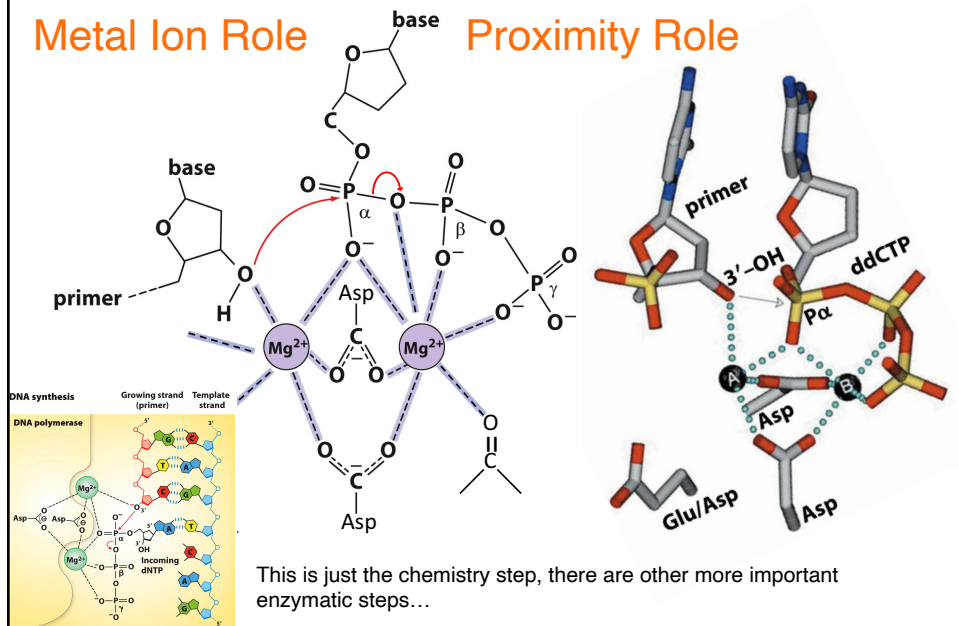
DNA/RNA polymerase mechanism



DNA/RNA polymerase mechanism

Metal Ion Role

Proximity Role



DNA Replication

- DNA Replication
 - Polymerases
 - Activities
 - Structure
 - Mechanism
- Replication/polymerase fidelity
 - Exonuclease Activities
 - Mismatch repair
 - Post-replication repair
 - Reversal
 - Base Excision repair
 - Nucleotide Excision repair
- DNA Sequencing
- PCR

Replication/polymerase Fidelity Geometry of Base Pairing Accounts for High Fidelity

- Errors in replication for *E. coli*:
 $1/10^9 - 1/10^{10}$ bp
 - 3×10^6 bp/genome $\times 1/10^{-10}$ mistakes/bp
 = 0.0003 mistakes/genome
 - 1 per 1,000–10,000 replications
- DNA polymerase active site excludes base pairs with incorrect geometry
 - At BOTH the **insertion site** and the **post insertion site**
 - BUT, DNA polymerases still insert wrong base 1/10,000 times.
 - Repair mechanisms fix these errors.

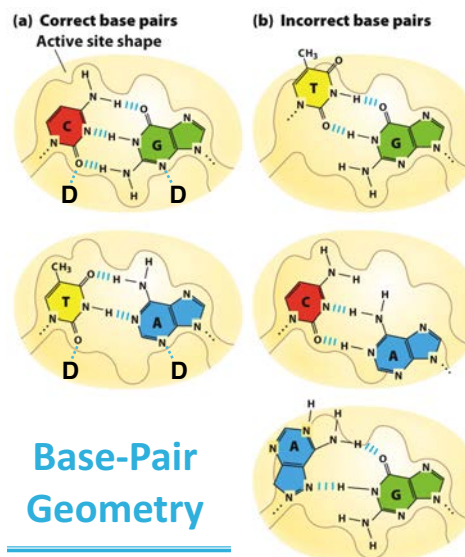
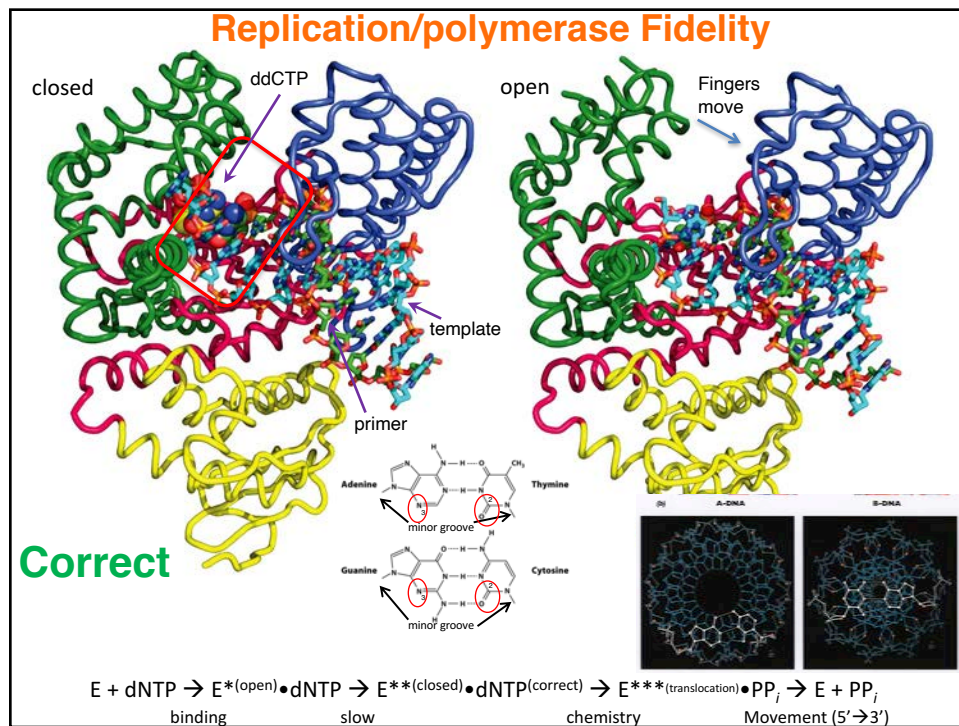
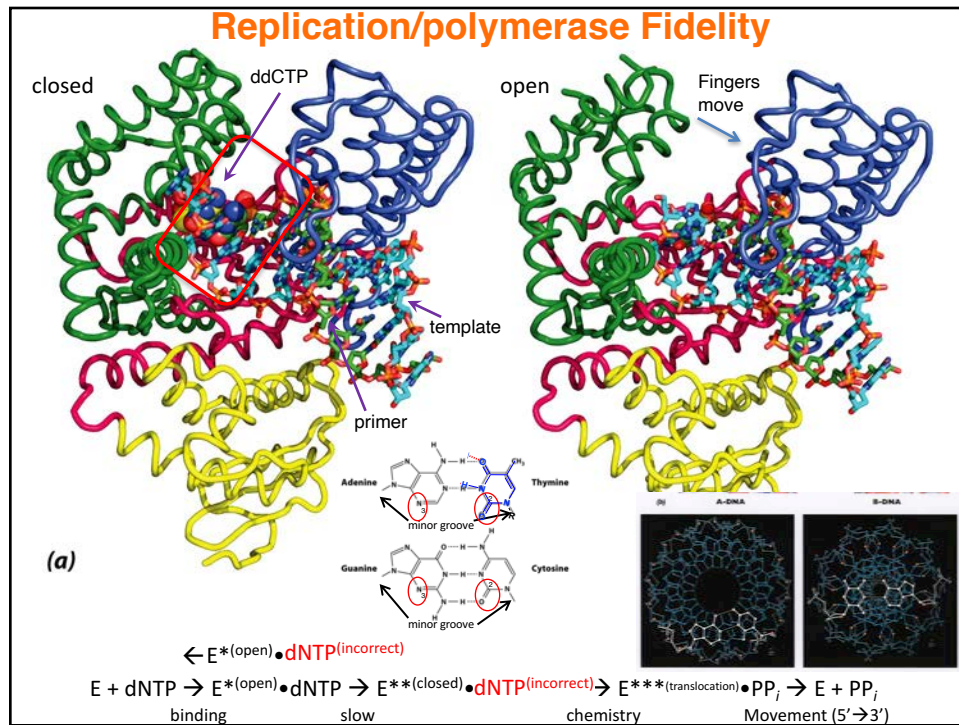
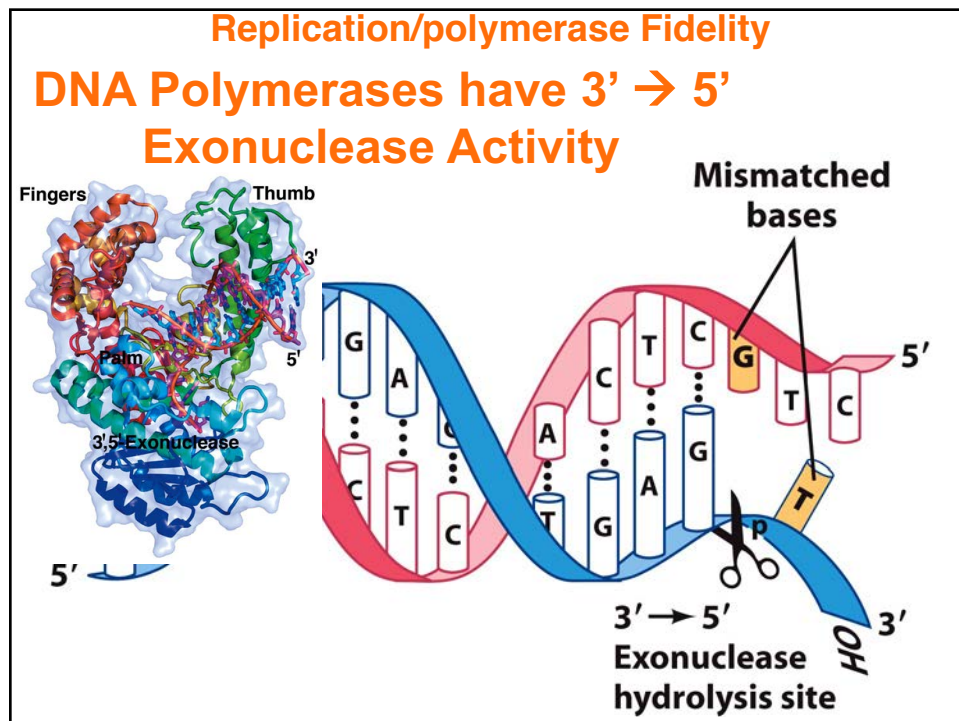
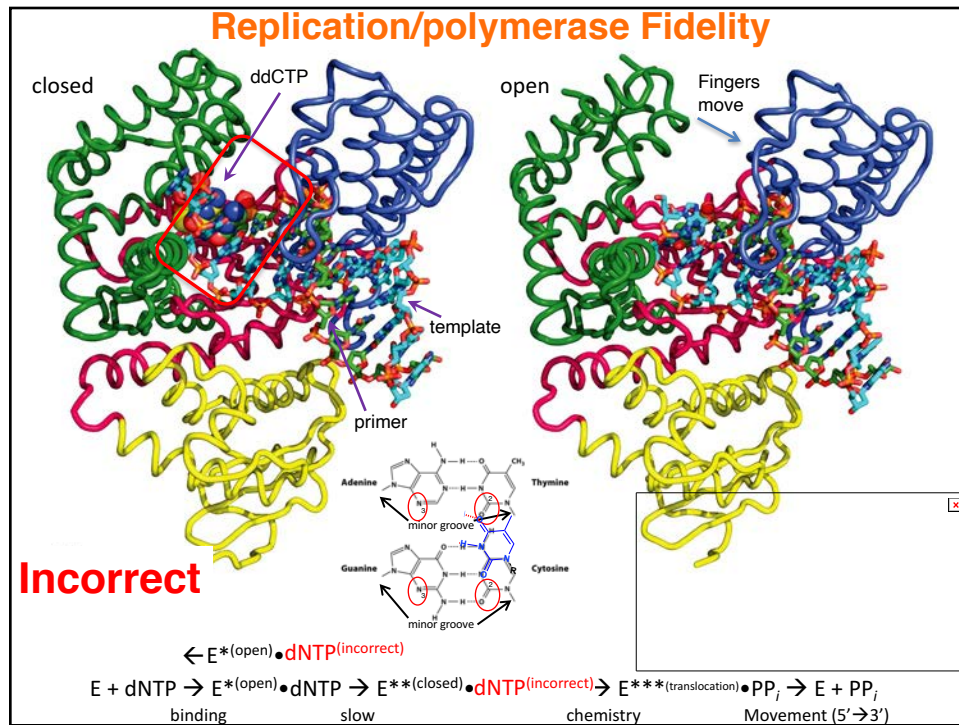
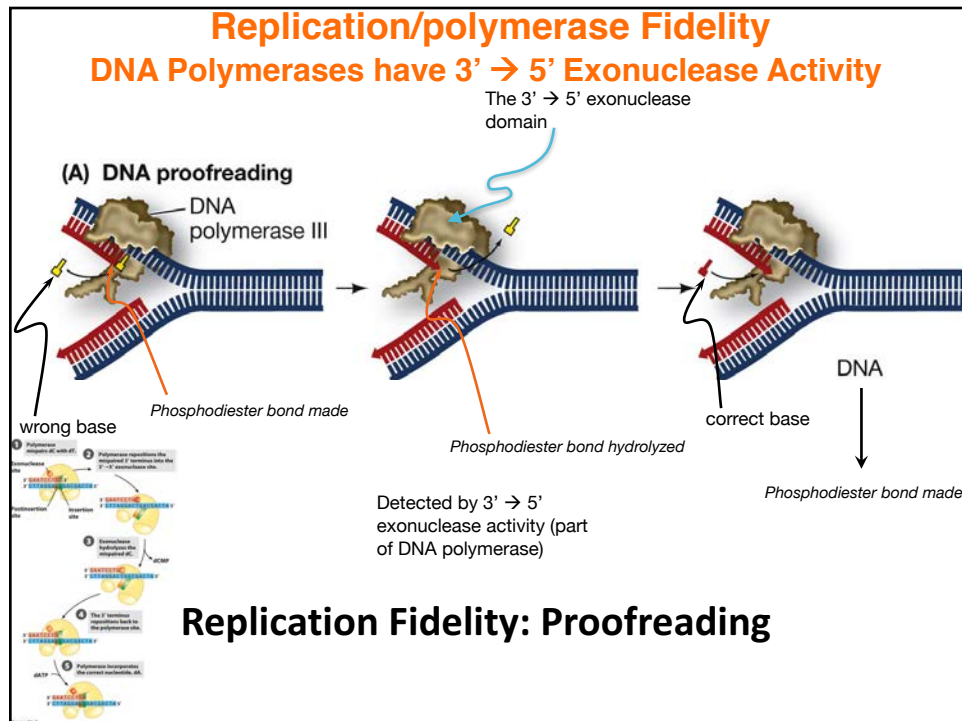


Figure 25-6







Replication/polymerase Fidelity

Comparison of Polymerases

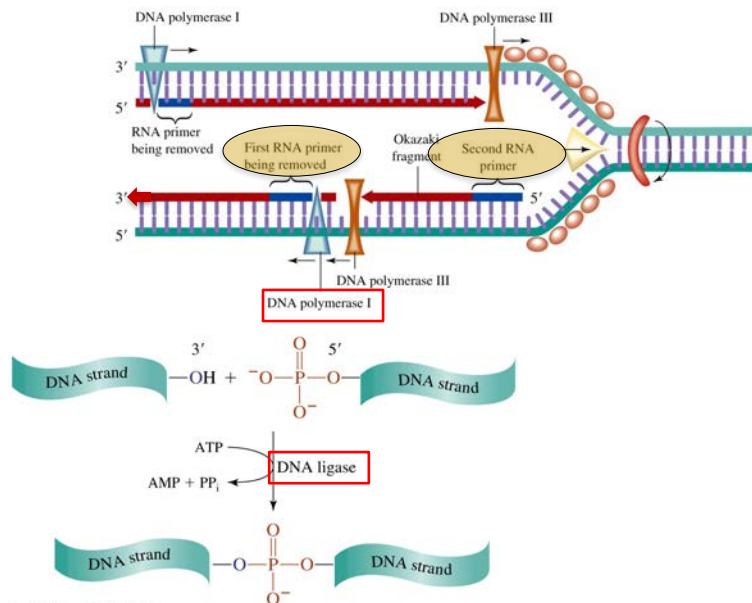
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DNA Replication Process



Pol I Has 5' → 3' Exonuclease Activity

Nick Translation

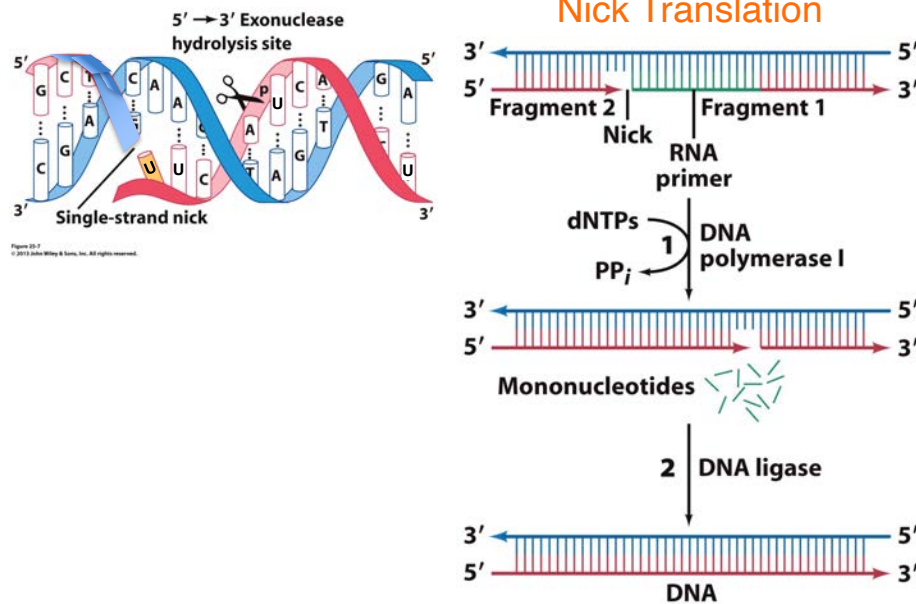


Figure 25-8
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Replication/polymerase Fidelity

Polymerases make mistakes in synthesis (polymerase 5'→3' activity) = 1/10,000

DNA polymerases have proofreading ability (3'→5' exonuclease activity) = 1/10,000

After replication fork, any mistakes are corrected by **mismatch repair** mechanism. This misses = 1/100

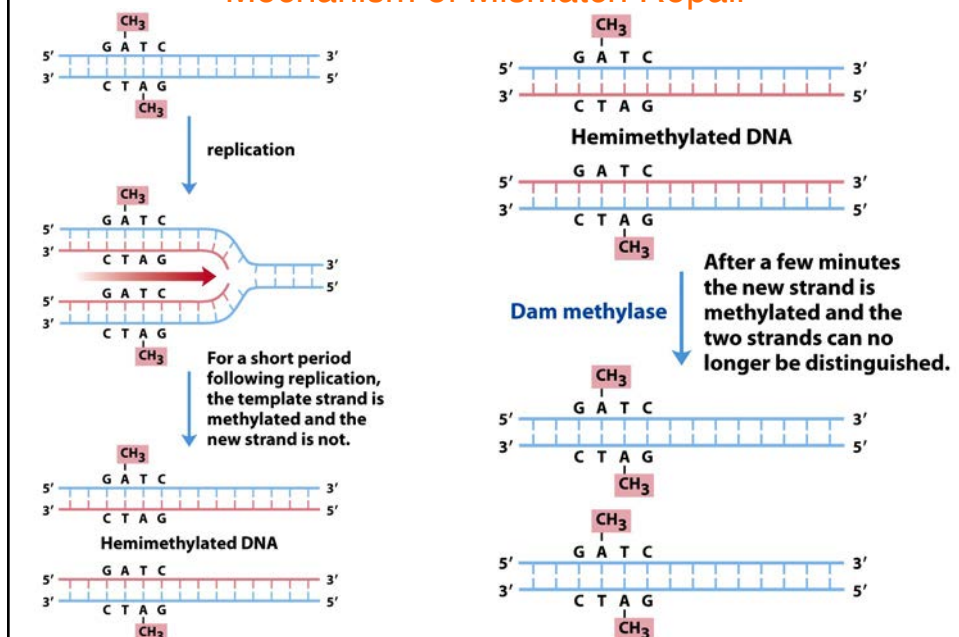
Overall for Replication = $1/10^{10}$

3×10^9 bp/haploid genome \times $1/10^{10}$ mistakes/bp = 0.3 mistakes/haploid genome

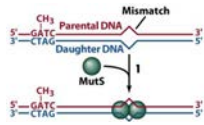
Plus DNA can be damaged in living cells. These damages are repaired by other mechanisms called Post-replication DNA Repair:

- Direct reversal of modification
- Base-excision repair
- Nucleotide excision repair

Replication/polymerase Fidelity Mechanism of Mismatch Repair



Replication/polymerase Fidelity Mechanism of Mismatch Repair



In *E. coli* a mismatch is recognized by MutS, which then recruits MutL.

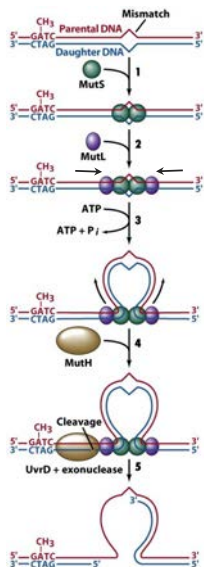
In an ATP dependent manner, the complex translocates along the double stranded DNA to generate a loop.

On encountering a hemi-methylated GATC sequence, it recruits MutH (an endonuclease) and MutH cuts the unmethylated DNA.

A helicase (UvrD) then separate the two strands and the exonuclease completely removes the defected strand.

The gap is then filled by DNA polymerase and DNA ligase.

Replication/polymerase Fidelity Mechanism of Mismatch Repair



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DNA Repair

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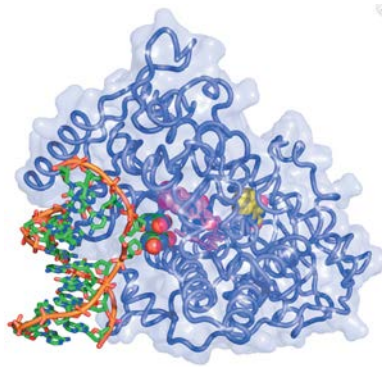
$$3 \times 10^9 \text{ bp/haploid genome} \times 1/10^{10} \text{ mistakes/bp} = 0.3 \text{ mistakes/haploid genome}$$

Plus DNA can be damaged in living cells. These damages are repaired by other mechanisms called Post-replication DNA Repair:

- **Direct reversal of modification (I)**
- **Base-excision repair (II)**
- **Nucleotide excision repair (III)**

DNA Repair

DNA Repair (I): Direct Reversal of Modification



DNA photolyase

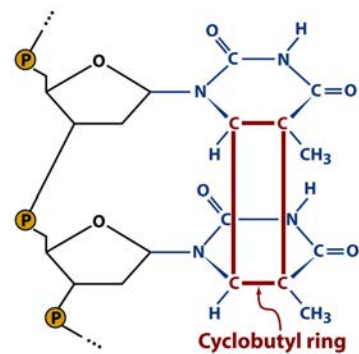
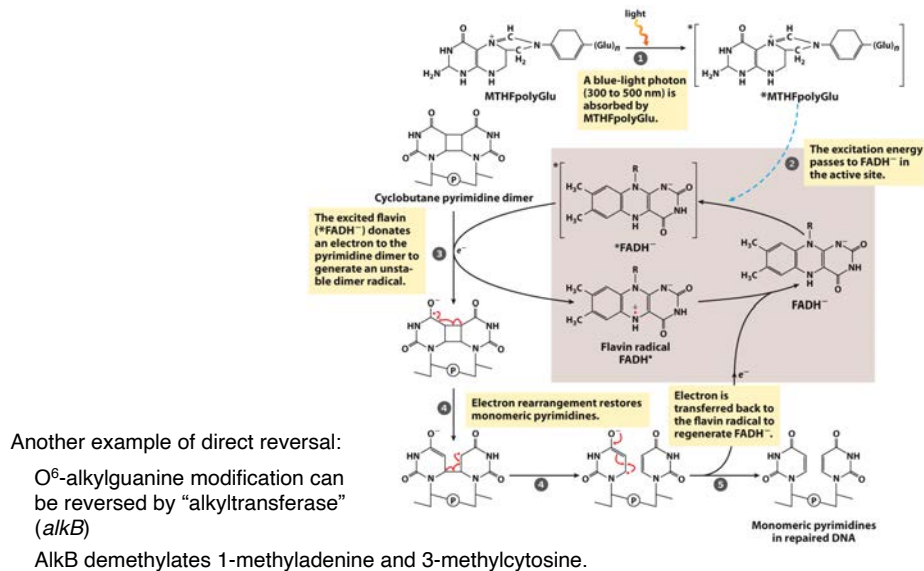


Figure 24-27 Fundamentals of Biochemistry, 2/e
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UV-crosslinking: pyrimidine dimers

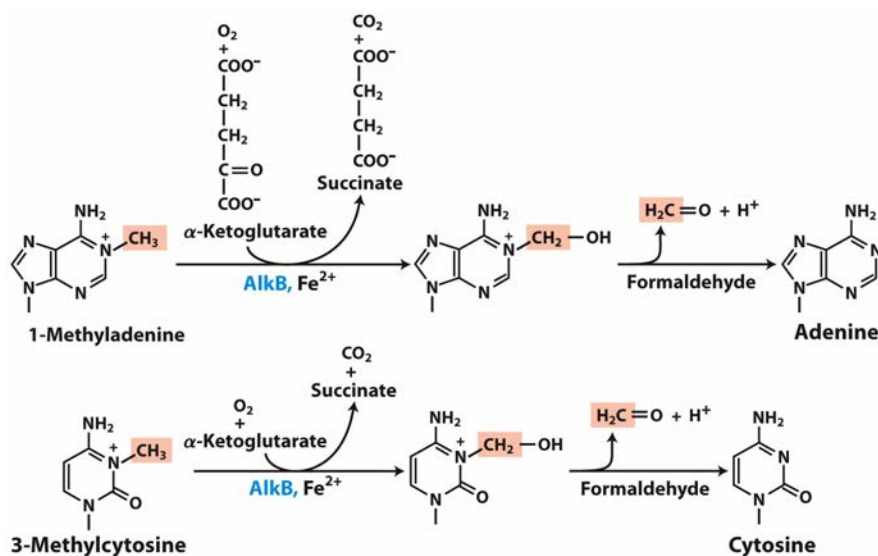
DNA Repair

Repair of Pyrimidine Dimers with Photolyase



DNA Repair

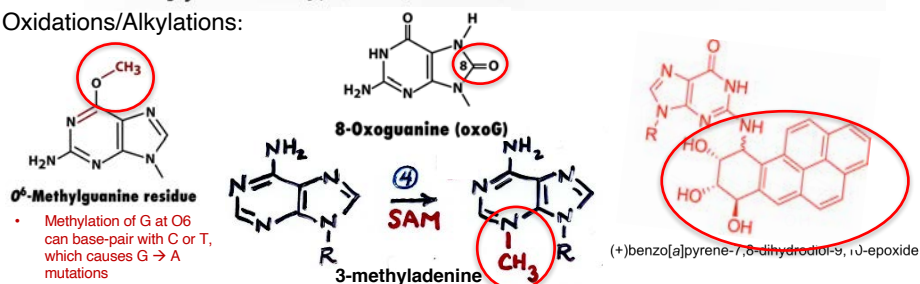
Direct Repair of Alkylated Bases by AlkB



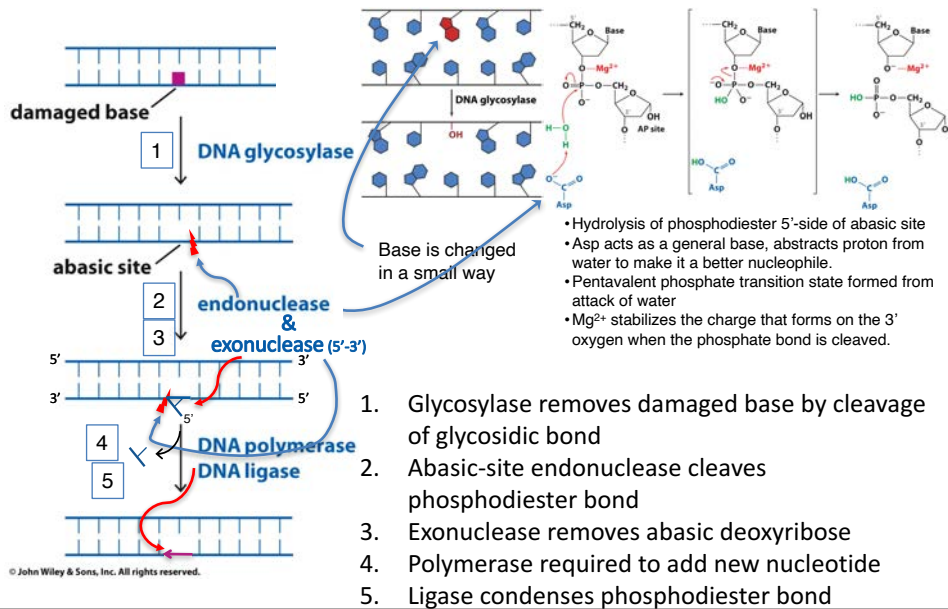
RECALL: Processes which chemically degrade DNA

Process	Events/cell/day
1. Spontaneous hydrolysis of glycosidic bonds [G ~ A ~ 20X C,T]	2-10 x 10 ³
2. Deamination of cytosine (and 5-MeC) [C ~ 5-MeC ~ 50X A > G]	100-500
➡ 3. Oxidation of guanine to 8-hydroxyguanine	100-500
➡ 4. Formation of 3-methyladenine and other bases	600
➡ 5. Photochemical pyrimidine dimer formation [cyclobutane type; 6,4-product; spore photoproduct]	variable

Oxidations/Alkylations:

**DNA Repair****DNA Repair (II): Base excision Repair**

abasic endo-nuclease mechanism



DNA Repair

STEP 1

DNA Repair (II): Base excision Repair

Depends on there being a specific Glycosylase

[Glycosylases exist for 8-Oxo-guanine (8-hydroxyG), uracil, hypoxanthine, 3-methyladenine]

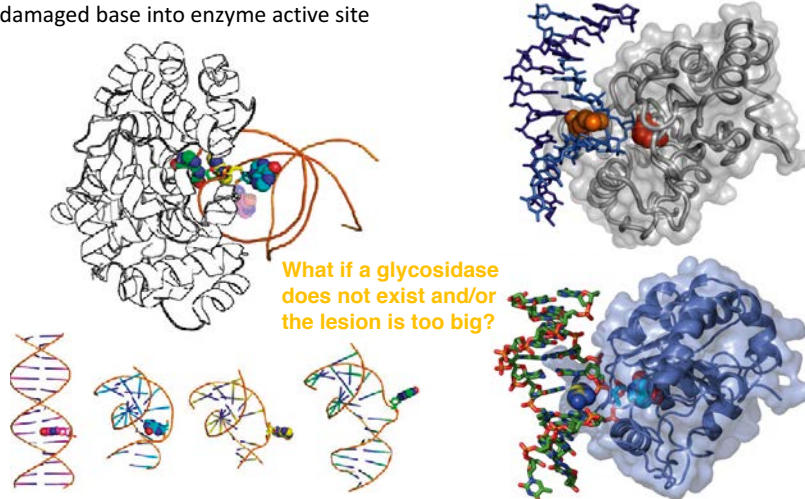
Oxo-guanine glycosylase

Bends DNA helix and extrudes

damaged base into enzyme active site

Uracil DNA glycosylase

Flips out base and fills hole with Arg

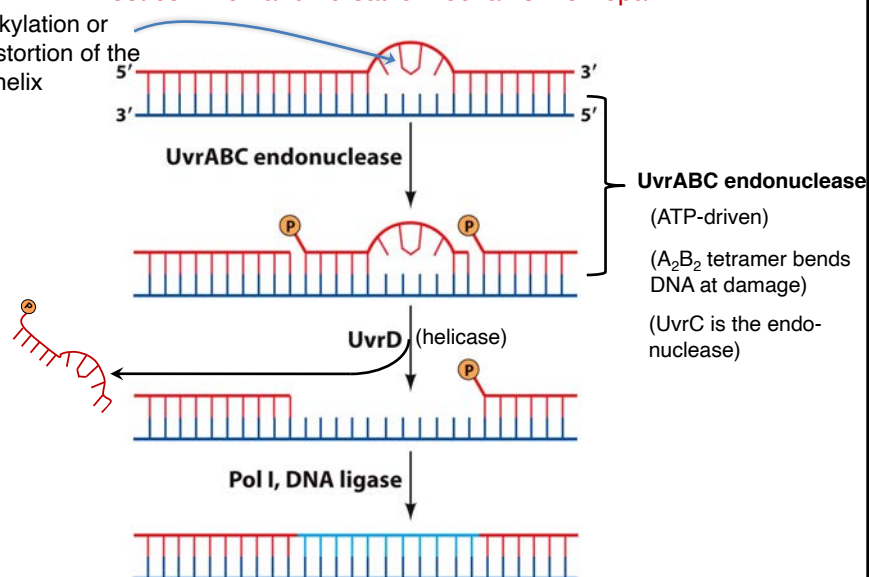


DNA Repair

DNA Repair (III): Nucleotide Excision Repair (NER)

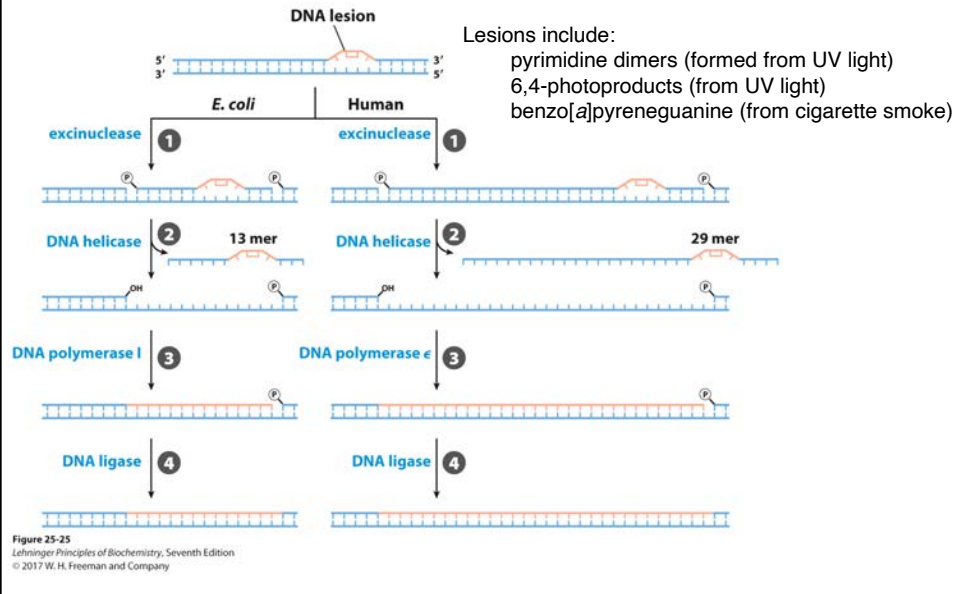
Most common and versatile mechanism of repair

Bulky alkylation or other distortion of the double helix



DNA Repair

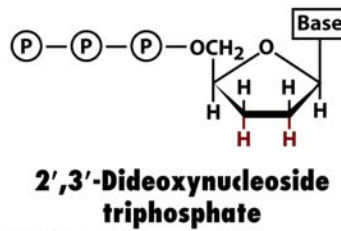
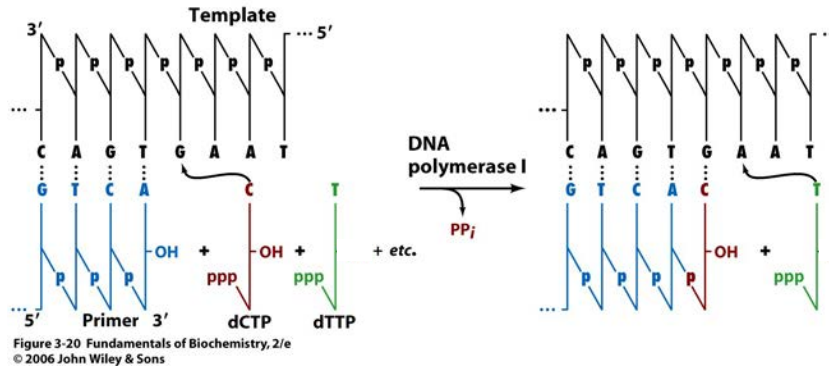
NER in Bacteria and Humans



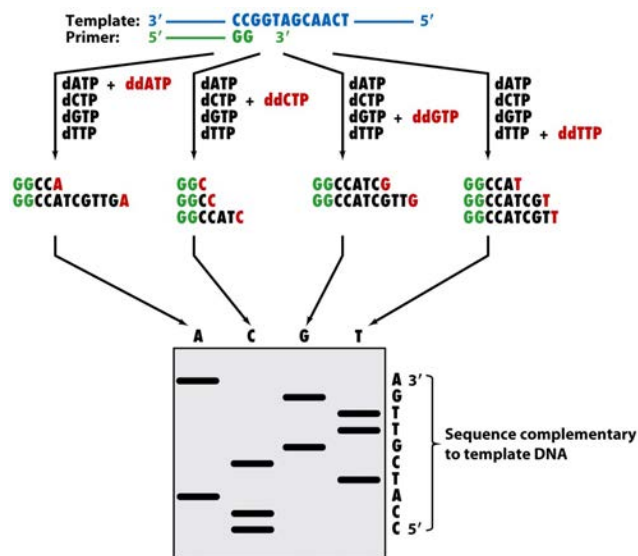
DNA Replication

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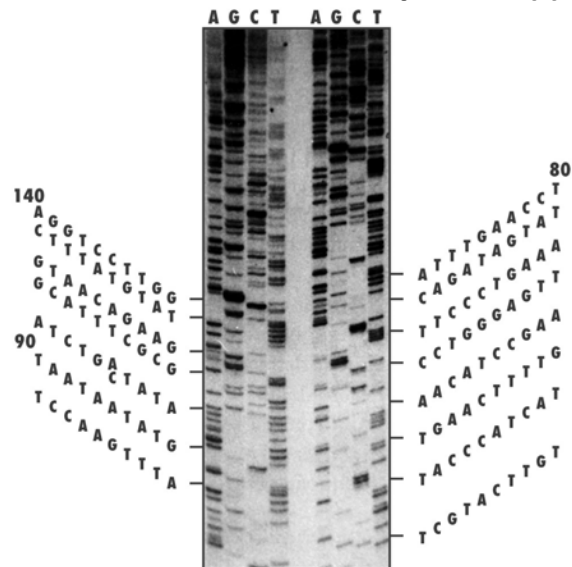
DNA Sequence Determination (Sanger)



DNA Sequencing

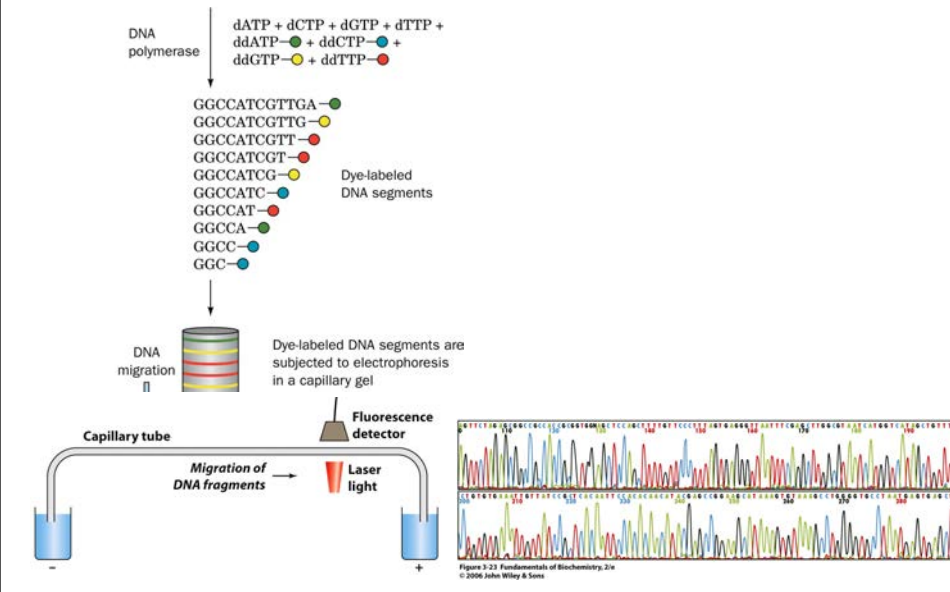


Visualize the DNA Sequence (I)



Chain-Terminator Sequencing Method

Template: 3' — CCGGTAGCAACT — 5'
 Primer: 5' — GG — 3'



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Molecular Cloning: Polymerase Chain Reaction

Besides using host to MAKE MORE copies of DNA sequences, they can be made by the **polymerase chain reaction (PCR)** technique (*in vitro*).

PCR is a cyclical process:

- DNA fragments are **denatured** by heating → single-stranded DNA
- Primers (one complimentary to each strand), plus dNTPs, and DNA polymerase are added → **annealing** and **extension** → new double-stranded DNA
- Repeat

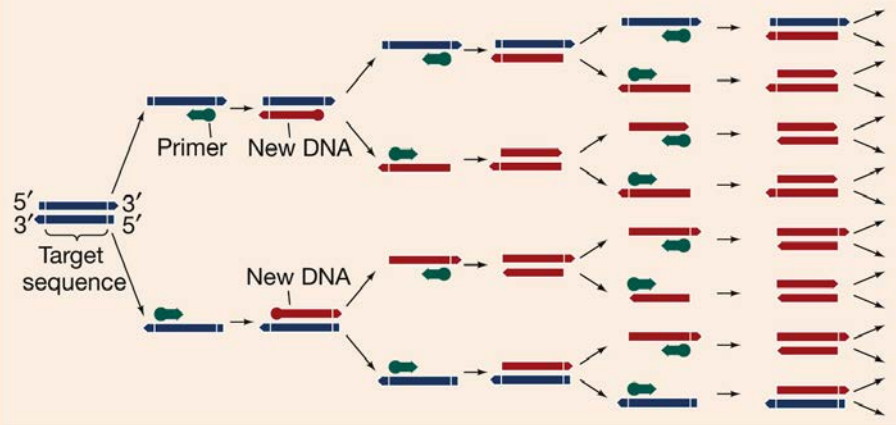
An initial problem with PCR was its temperature requirements.

The key insight was the use of an enzyme that could withstand the heat needed to denature the DNA → DNA polymerase from *Thermus aquaticus*, **Taq Polymerase** (insight of Kerri Mullis).

The amplified DNA can then be inserted into plasmids to create recombinant DNA and cloned in host cells. Synthetic DNA can be manipulated to create specific mutations in order to study the consequences of the mutation: Site-directed Mutagenesis.

Process Diagram: Polymerase Chain Reaction (PCR)

TOOLS FOR INVESTIGATING LIFE



Molecular Cloning: Polymerase Chain Reaction