

## Lecture 11 (10/2/20)

- Reading: Ch4; 125<sub>(Fig 4-10)</sub>, 134-136<sub>(structure determination)</sub>
- Problems: Ch4 (text); 10, 15

### NEXT

- Reading: Ch4; 127-130 (Collagen)  
Ch1; 27-29 (Enzymes)  
Ch5; 157-158, 160-161, 166<sub>(bottom)</sub>
- Problems: Ch1 (text); 16  
Ch4 (text); 1, 4, 5, 6  
Ch4 (study-guide); 6 (*do you know the facts?*)

## Lecture 11 (10/2/20)

### OUTLINE

#### Protein Characterization

##### A. Quaternary structure

1. How determined;
  - a. native size
  - b. subunit size

2. Ultracentrifugation

##### B. Tertiary structure

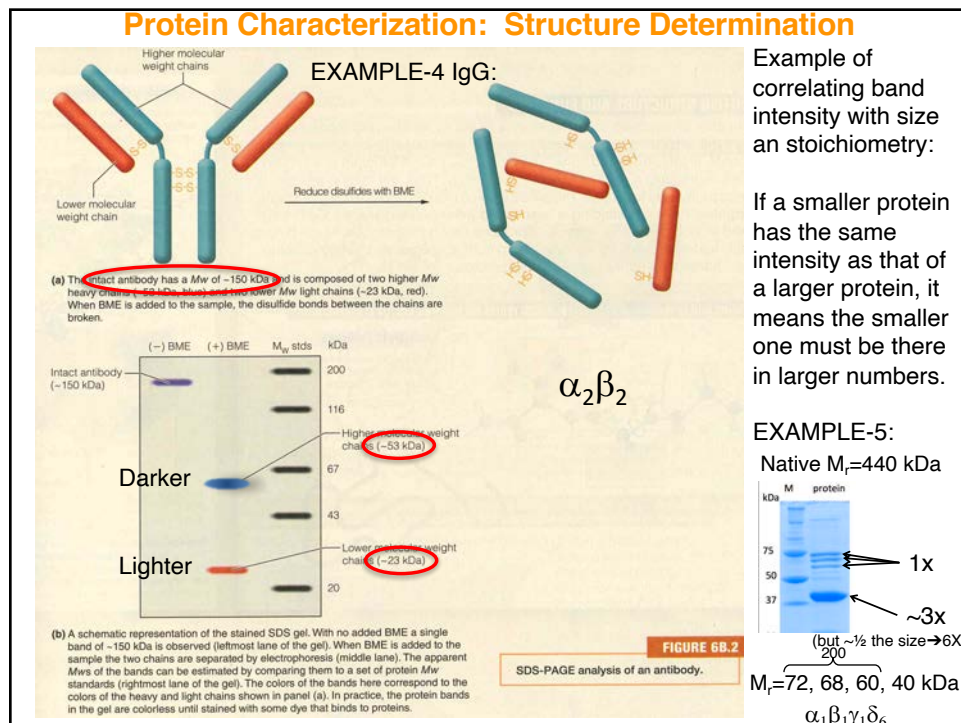
1. X-ray diffraction/crystallography
2. NMR spectroscopy
3. Comparison: NMR *versus* X-ray crystallography

##### C. Secondary structure

1. Circular dichroism (CD)

# Protein Characterization

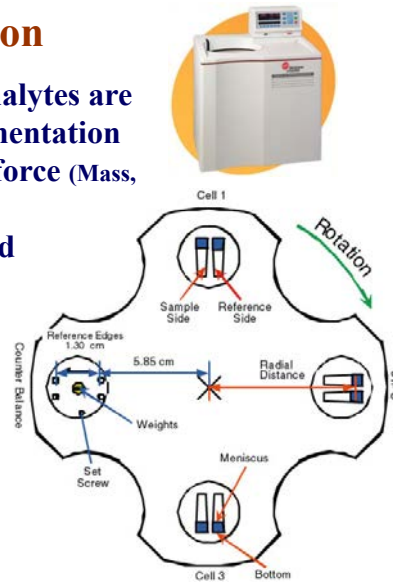
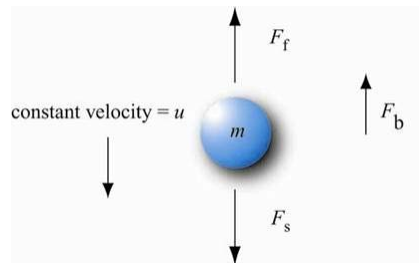
## Quaternary Structure



## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation

- ✳ In analytical ultracentrifugation, analytes are separated on the basis of their sedimentation when they experience a centripetal force (Mass, Shape, Density):  $\sim 1.4 \text{ g/cm}^3$
- ✳ Usually carried out at speeds around 60,000 rpm



There is a force due to buoyancy  $F_b$  and a frictional force on the molecule  $F_f$ , both which oppose its movement through the medium  $F_s$  (centrifugal force or sedimentary force).

## Protein Characterization: Structure Determination

### TWO Types:

- Analytical Ultracentrifugation – Sedimentation Velocity
- Analytical Ultracentrifugation – Sedimentation Equilibrium

- ✳ Or, described mathematically...  $F_s + F_b + F_f = \text{Sed. Vel. (rate)}$

Solvent density in g/mL

Specific Volume (occupied by 1 g of solute ( $\text{cm}^3/\text{g}$ ))

angular velocity


sedimentation velocity

radius from center of spin

Sedimentation coefficient (units of sec)

coefficient of friction ( $\text{g/mole}\cdot\text{sec}$ )

$$\frac{M(1-v\rho)}{Nf} = \frac{u_s}{\omega^2 r} \equiv s$$



Theodor Svedberg

$M$  is number average molecular weight ( $mN$ ).  
If all species are the same, then  $M = M_r$

## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Velocity

$$\frac{M(1-\bar{v}\rho)}{Nf} = \frac{\mu_s}{\omega^2 r} \equiv s$$

Sedimentation  
coefficient  
(units of sec)

☀ Units of sedimentation are seconds: 1 Svedberg =  $1\text{S} = 10^{-13}\text{ s}$

$1 - \bar{v}\rho$  is called the buoyancy factor – and since  $1/\bar{v}$  approximates  $\rho$ , then if the (Say, “Vee-bar-rho”)

density of the molecule is close to that of the solution  $1 - \bar{v}\rho = 0$  and the  $s = 0$ .

☀ If the molecule is less dense than solution then  $1 - \bar{v}\rho < 0$  and the molecule floats. (lipids or fats in a blood sample)

☀ Proteins are more dense than solution and  $1 - \bar{v}\rho > 0$ , and the molecule sediments.

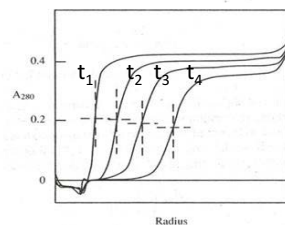
## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Velocity

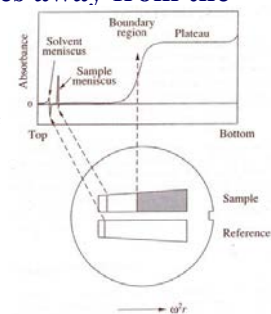
☀ The most basic type of ultracentrifugation experiment is to measure the rate at which the molecule moves away from the center of rotation

☀ What is actually measured is the movement of the **boundary** between dissolved molecule and ‘empty’ buffer

☀ Based on mass, shape, and density



Note shape of curves-  
less steep over time because  
of diffusion



## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Velocity

Protein	Molecular Mass (kD)	Partial Specific Volume, $\bar{V}_{20,w}$ ( $\text{cm}^3 \cdot \text{g}^{-1}$ )	Sedimentation Coefficient, $S_{20,w}$ (S)
Lipase (milk)	6.7	0.714	1.14
Ribonuclease A (bovine pancreas)	12.6	0.707	2.00
Cytochrome c (bovine heart)	13.4	0.728	1.71
Myoglobin (horse heart)	16.9	0.741	2.04
$\alpha$ -Chymotrypsin (bovine pancreas)	21.6	0.736	2.40
Crotoxin (rattlesnake)	29.9	0.704	3.14
Concanavalin B (jack bean)	42.5	0.730	3.50
Diphtheria toxin	70.4	0.736	4.60
Cytochrome oxidase ( <i>P. aeruginosa</i> )	89.8	0.730	5.80
Lactate dehydrogenase H (chicken)	150	0.740	7.31
Catalase (horse liver)	222	0.715	11.20
Fibrinogen (human)	340	0.725	7.63
Hemocyanin (squid)	612	0.724	19.50
Glutamate dehydrogenase (bovine liver)	1015	0.750	26.60
Turnip yellow mosaic virus protein	3013	0.740	48.80

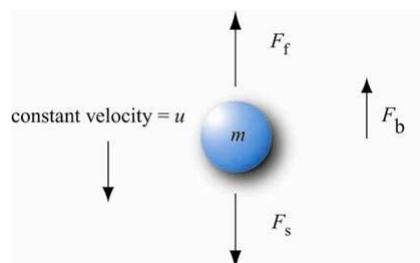
Source: Smith, M.H., in Sober, H.A. (Ed.), *Handbook of Biochemistry and Molecular Biology* (2nd ed.), p. C-10, CRC Press (1970).

$$\frac{M(1-\bar{v}\rho)}{Nf} = \frac{\mu_s}{\omega^2 r} \equiv s$$

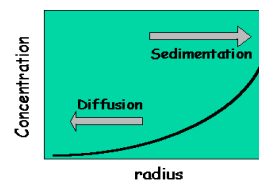
## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Equilibrium

- ✳ In sedimentation equilibrium, an equilibrium is established between **sedimentation away** from the center of rotation and **diffusion towards** the center of rotation (spin at much lower speeds) so we get no boundary between solute and meniscus

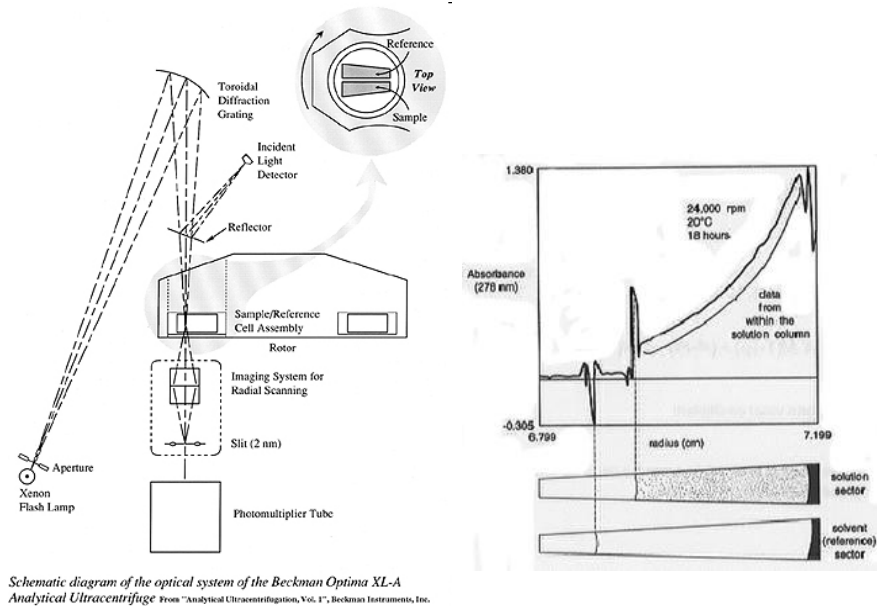


This is anything that measures concentration!



## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Equilibrium



## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Equilibrium

#### Determining Molecular Weight

Can be described by equation:  $C_A(r) = C_{A,0} e^{\sigma(r^2 - r_0^2)/2}$  where  $\sigma = \frac{M(1 - \bar{v}\rho)\omega^2}{RT}$

This equation can be expressed in terms of M:

$$M = \frac{2kT}{(1 - \bar{v}\rho)\omega^2} \ln \frac{C_A}{C_0(r^2 - r_0^2)}$$



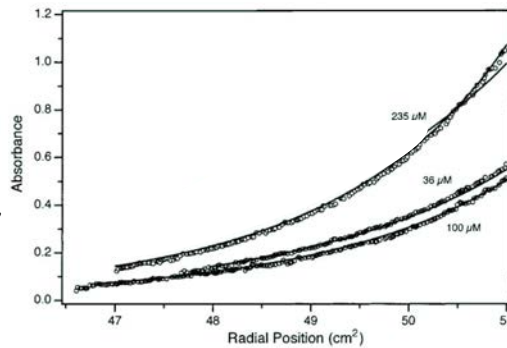
## Protein Characterization: Structure Determination

### Analytical Ultracentrifugation – Sedimentation Equilibrium

$$M = \frac{2kT}{(1 - \bar{v}\rho)\omega^2} \ln \frac{C_A}{C_0(r^2 - r_0^2)}$$

Thus, we can get M.

At most radii, its migrating “heavier” due to association.



$$A_2 \cong 2A$$

$$K_d = [A]^2/[A_2]$$

But recall, this  $M = mN$ , and depends on the number average molecular weight. So, if there are species with different molecular weights, as in a dissociation equilibrium of an oligomeric protein, this  $M$  will be sensitive to that dissociation.

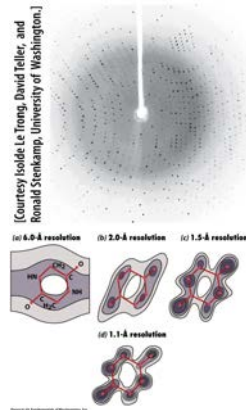
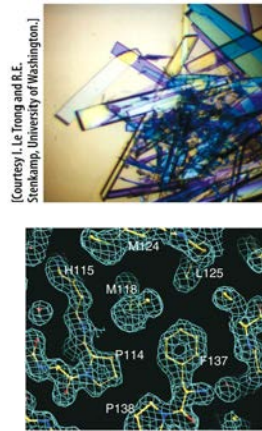
## Protein Characterization

### Tertiary Structure

## Protein Characterization: Structure Determination

### Analysis of 3° structure\*

- X-ray crystallography



- NMR



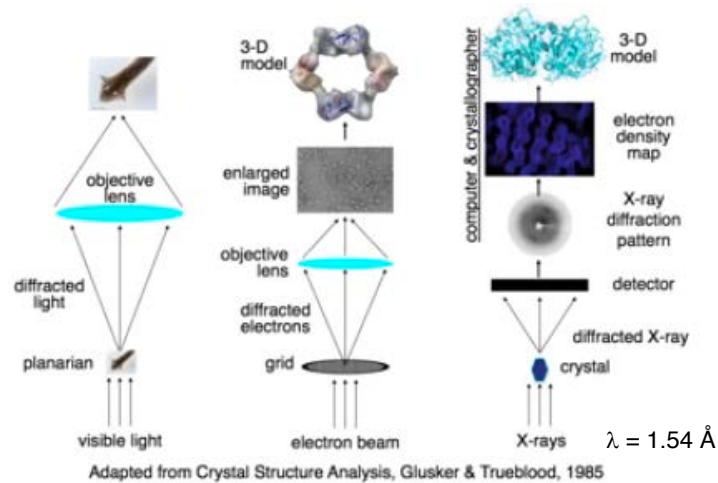
\*and 4° structure

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## Protein Characterization: Structure Determination

### X-ray Crystallography for Protein Structure

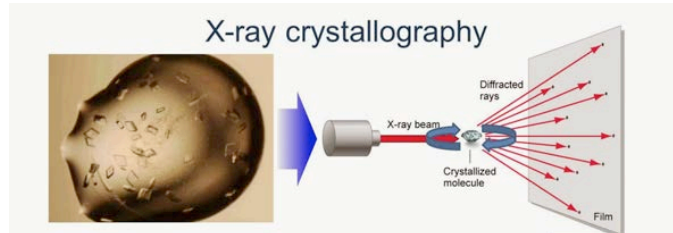
Comparison to other methods:



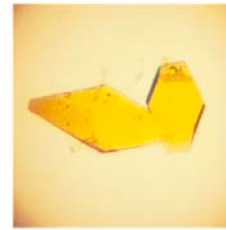


## Protein Characterization: Structure Determination

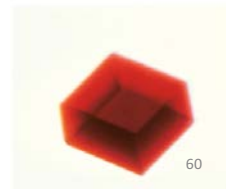
### X-ray Crystallography for Protein Structure



(a)



(b)



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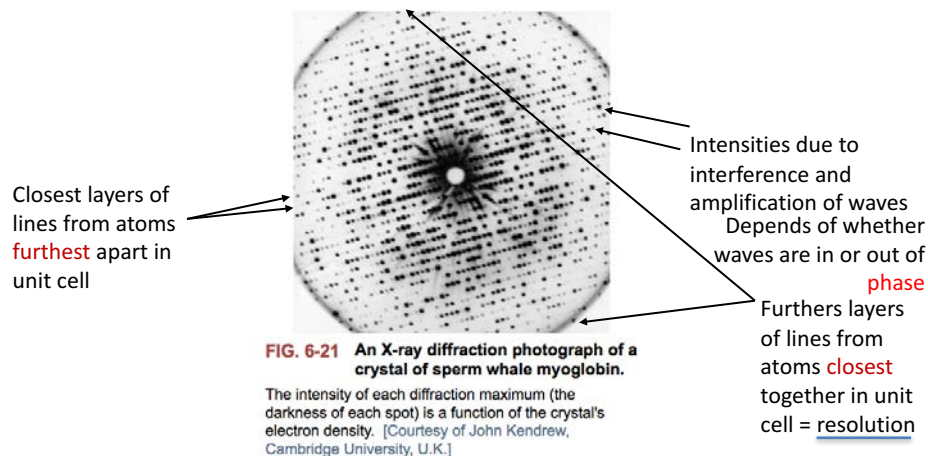
X-ray crystallography [Primer](#)

[https://www.youtube.com/watch?v=62kdCOHcN\\_U](https://www.youtube.com/watch?v=62kdCOHcN_U)

[For dummies](#) <https://www.youtube.com/watch?v=QuCRBxjk3fg>

## Protein Characterization: Structure Determination

### X-ray Crystallography for Protein Structure



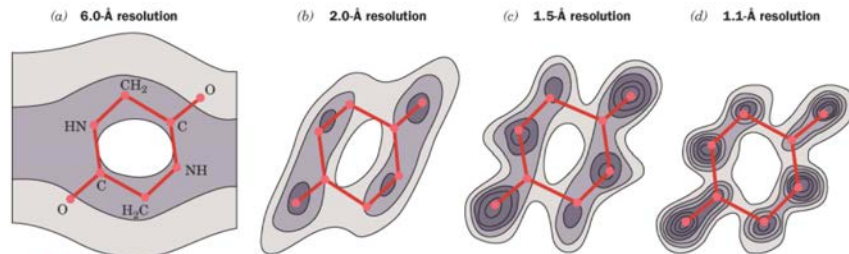
[Relationship of spots intensity and unit cell](#)

[<https://www.youtube.com/watch?v=fZ0m8wustVkj>]

## Protein Characterization: Structure Determination

### X-ray Crystallography for Protein Structure

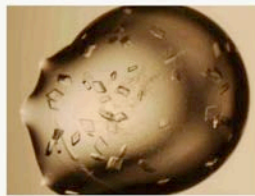
# Resolution

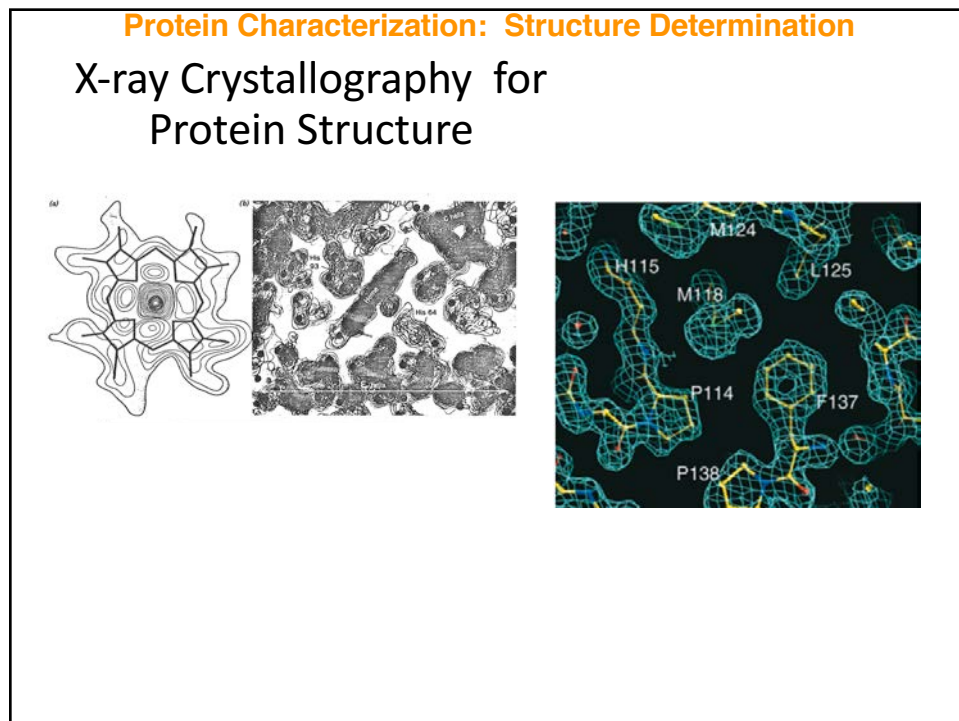
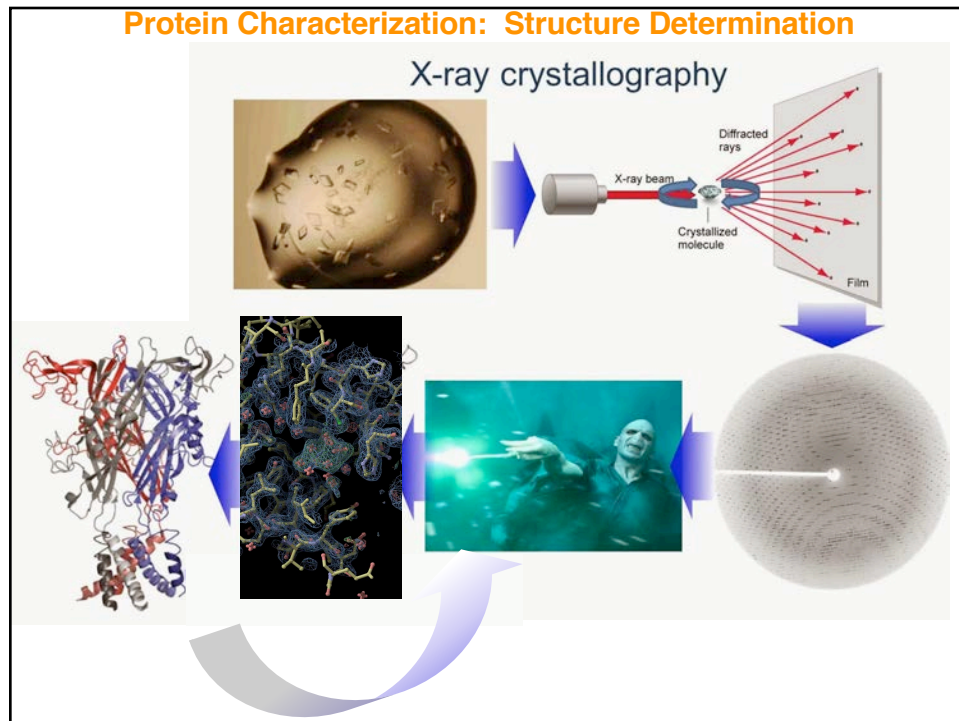


**FIG. 6-23** Electron density maps of diketopiperazine at different resolution levels. Hydrogen atoms are not visible in these maps because of their low electron density. [After Hodgkin, D.C., *Nature* 188, 445 (1960).]

## Protein Characterization: Structure Determination

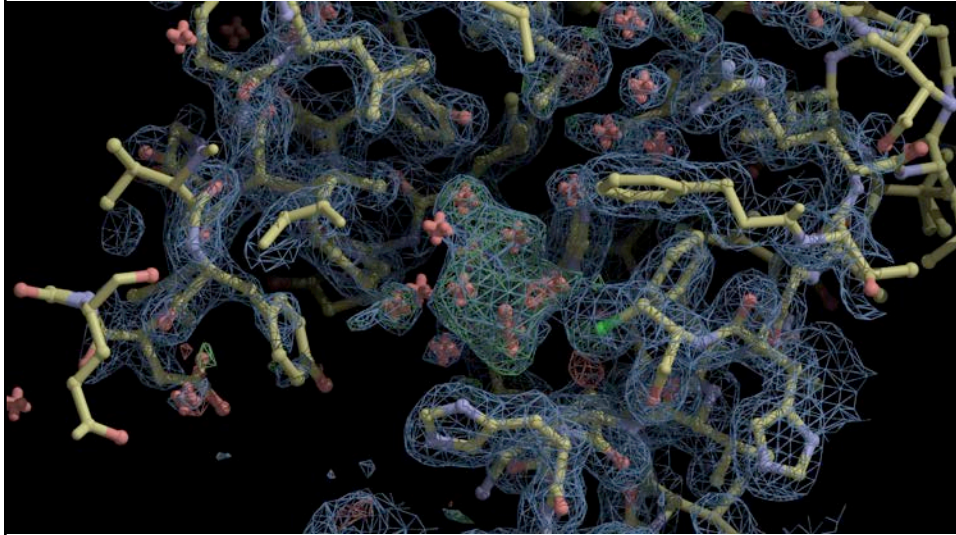
### X-ray crystallography





### Protein Characterization: Structure Determination

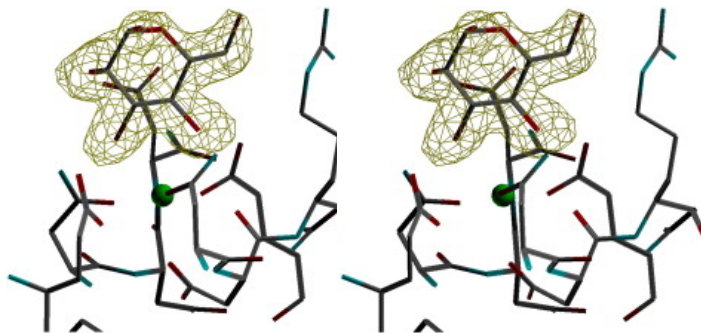
Typical X-ray crystallography structure



The electron density map from the x-ray diffraction data is shown as a blue cage, and the model of the protein that “fits” this density is modeled inside of it.

### Protein Characterization: Structure Determination

X-ray Crystallography for  
Protein Structure



Stereo View

## Protein Characterization: Structure Determination

### NMR for Protein Structure

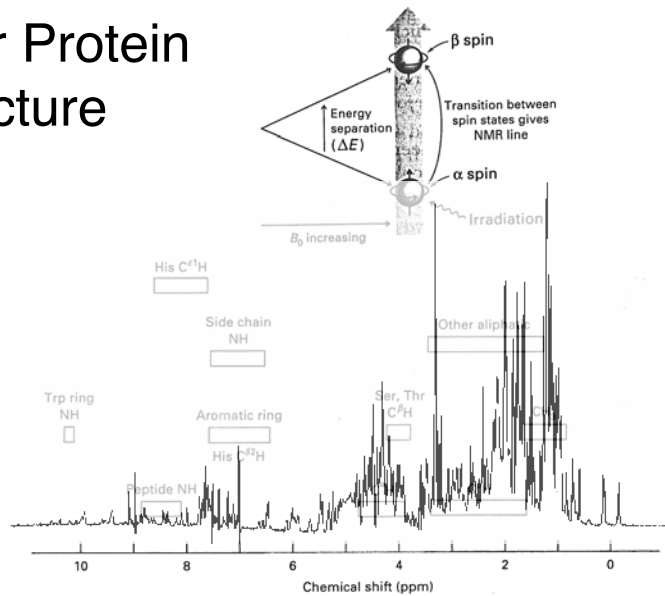


FIGURE 6.26

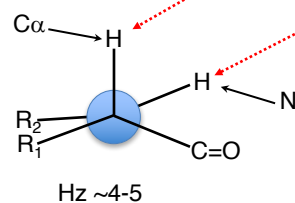
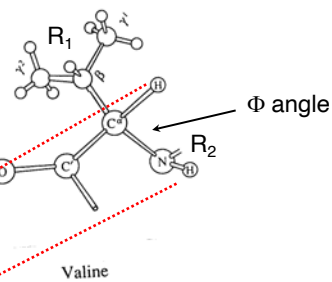
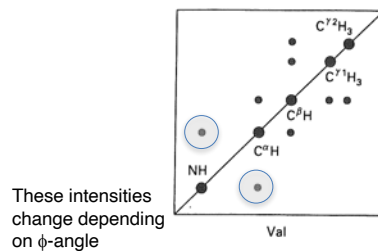
The range of  $^1\text{H}$ -NMR chemical shifts observed for different hydrogen atoms of peptides in the random coil conformation.

## Protein Characterization: Structure Determination

### NMR for Protein Structure

Correlation 2D spectroscopy (COSY)

J-coupling



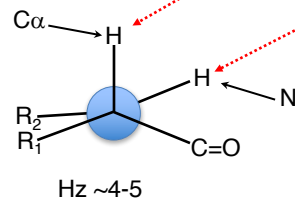
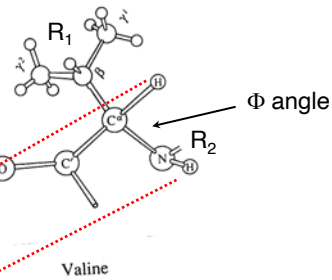
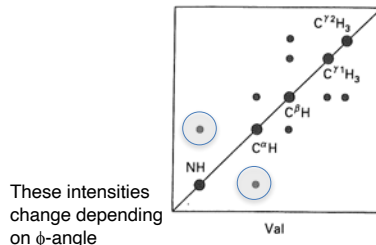
Can also get  $\phi$  &  $\psi$  angles from secondary chemical shifts (observed from known random coil shifts)(TALOS & ANGLOR)

## Protein Characterization: Structure Determination

### NMR for Protein Structure

Correlation 2D spectroscopy (COSY)

J-coupling

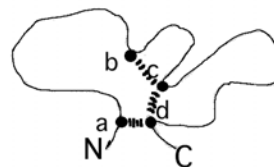
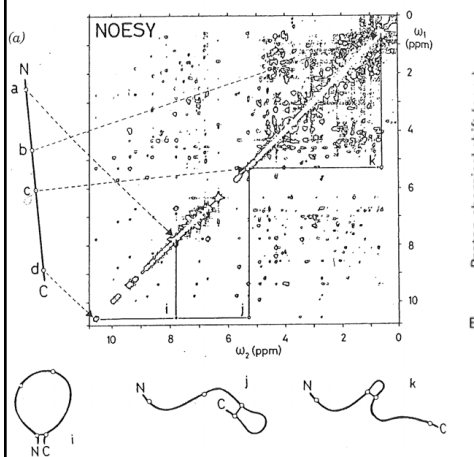


Can also get  $\phi$  &  $\psi$  angles from secondary chemical shifts (observed from known random coil shifts)(TALOS)

## Protein Characterization: Structure Determination

### NMR for Protein Structure

Nuclear-Overhauser Effect Spectroscopy (NOESY)

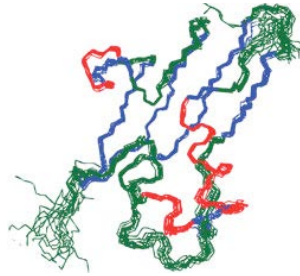


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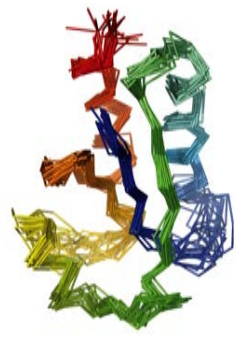


## Protein Characterization: Structure Determination

### Typical NMR structure



Blue =  $\beta$ -sheet, red =  $\alpha$ -helix, green = loops without 2° structure.

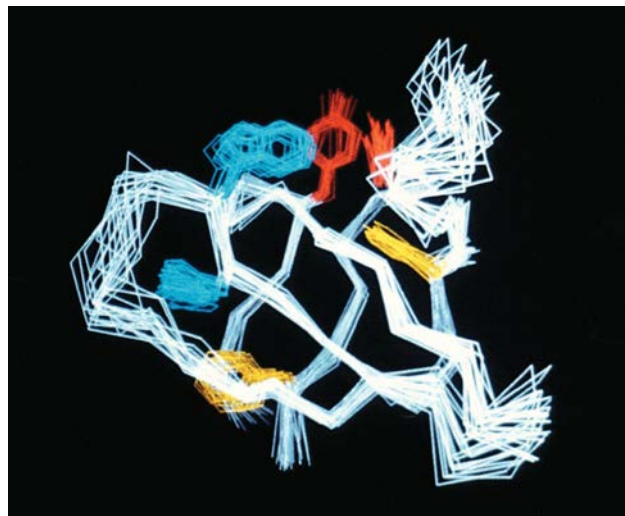


Blue = N-term, red = C-term  
"Rainbow coloring."

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## Protein Characterization: Structure Determination

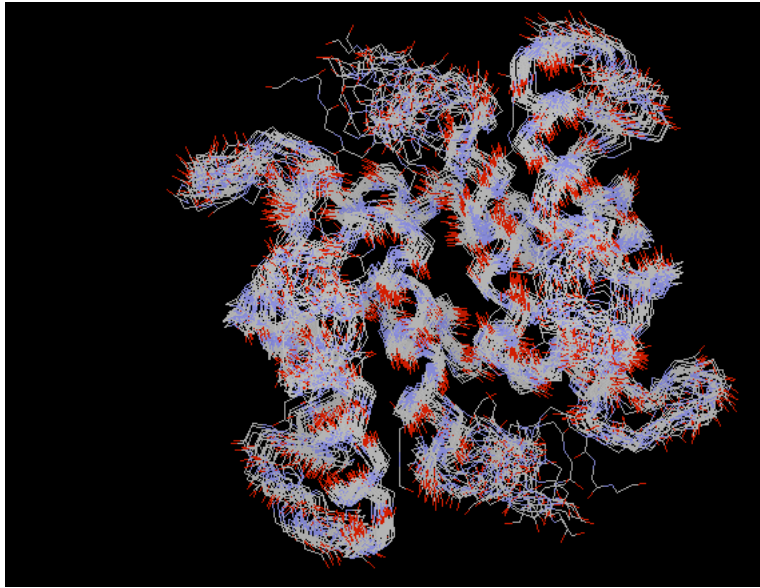
### NMR Structure of a Protein



Courtesy Stuart Schrieber, Harvard University

### Protein Characterization: Structure Determination

Typical NMR structure



Notice there are many overlapping structures that all fit the NMR data. Where it is tight, you have higher resolution and where it is loose you have parts of the molecule that are more mobile<sup>74</sup>

### Protein Characterization: Structure Determination

## Protein Characterization

### Tertiary Structure

Compare/Contrast X-ray crystallography and NMR:

- 1) Crystal vs. solution structures the same; not significant crystal constraints
- 2) NMR not as high resolution
- 3) NMR better at predicting regions that are dynamic; X-ray uses “B-factors” or even does not show, i.e., “disordered”
- 4) X-ray cannot distinguish “rotomers” of Asn, Gln, Thr; NMR is unambiguous
- 5) X-ray much better at larger structures; NMR has assignment problem only good for up to 30-40 kDa