<u>ENG BE 209</u>

COURSE SYLLABUS

SPRING 2013

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Introduction to the molecular mechanisms of cell function in the context of cutting-edge applications in bioengineering and medicine. Biological concepts include: molecular building blocks, energetics, reaction kinetics, nucleic acids and DNA repair, transcription, translation, regulation and cytoskeleton. Applications include bioenergy, biomanufacturing, antibiotics, diabetes, protein therapeutics, gene circuit engineering, & tissue engineering. Quantitative principles, computational methods, and experimental methods will be integrated into lectures and labs.

LECTURE/SEMINAR/EXAMINATION

<u>Required Text:</u> Essential Cell Biology, B. Alberts et. al. 2009 <u>Tuesday 11.30 am–2.30 pm (auxiliary date: even weeks: Friday 10 am-1 pm)</u> <u>Location (see map)</u>

Room 108b: Building 40, Dekanatsgebäude; Hörsaal 2 (HS2): Building 91, MTZ <u>Examination/Grading:</u> Active Attendance / written

Week	Time	Topic	Room
	<u>April</u>		
16	Tue 16	CH1 + CH2 + CH3	108b
17	Tue 23	CH4 + CH5	HS2
18	Tue 30, 12.00	CH6	HS2
	<u>May</u>		
18	Tue 07	EXAM SEMINAR 1: CH1-6	108b
19	Tue 14	CH7 + CH8	108b
22	Tue 28	CH9 + CH11	108b
	<u>June</u>		
23	Tue 04	CH12 + CH13	108b
24	Tue 11	CH14	108b
25	Tue 18, 12.00	EXAM SEMINAR 2: CH7-14	108b
26	Tue 25	CH15 + CH16	108b
	<u>July</u>		
27	Tue 02	CH17 + CH18	108b
28	Tue 09, 13.00	CH19 + CH20	108b
	Fri 12, 10.00	EXAM LABORATORY	108b
29	Tue 16	EXAM SEMINAR 3: CH15-20	108b

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Labs will provide practical experience with current methods used in molecular/cell biology and biotechnology. The first half of the course will cover protein folding, bioenergetics, cell growth, and microscopy. The second half will be devoted to a single multi-week module on genetic engineering and gene regulation. Students will build a bacterial gene expression system, learning recombinant DNA technology, cloning and gene expression measurement techniques. Labs will emphasize the experimental and analytical skills required in modern engineering and scientific research.

LABORATORY

<u>Required:</u> Laboratory Manual – White Lab coat <u>Tuesday:</u> 3.00 pm – 6.00 pm <u>Location:</u> MTZ, Fiedlerstrasse 42 (map: Bld. 91) <u>Examination/Grading:</u> Active Attendance / Written

Day	Content	Discipline Responsibility	Location
April 09	Laboratory 1 Analysis of Simulated Epidemic and Hand Contamination and Cytological Studies	Microbiology Prof. Jacobs	Mikroskopiersaal
April 16	Laboratory 2 General Histology	Anatomy Prof. Kasper	Mikroskopiersaal
April 23	Laboratory 3 Quantitative Determination and Spectrums of Haemoglobins in Blood	Biochemistry Dr. Scheibe	Institut für Physiologische Chemie
April 30	Laboratory 4 Determination of Activity of Lactate Dehydrogenase in Optical Test	Biochemistry Dr. Hempel	Institut für Physiologische Chemie
May 07	Laboratory 5 Molecular Biological Diagnosis of Cystic Fibrosis with the Help of PCR	Biochemistry Dr. Kreutzmann	Institut für Physiologische Chemie
May 14	Laboratory 6 Characterization of DNA using Restriction Endonucleases	Biochemistry Dr. Kreutzmann	Institut für Physiologische Chemie
May 28	Laboratory 7 – Group I Bacterial Transformation	Immunology Prof. Roers Anett Skupin	Institut für Immunologie
June 04	Laboratory 7 – Group II Bacterial Transformation	Immunology Prof. Roers Anett Skupin	Institut für Immunologie
June 11	Laboratory 8 – Group I Plasmid Preparation from <i>E. coli</i> and Sequencing of Plasmid DNA	Immunology Prof. Roers Anett Skupin	Institut für Immunologie
June 18	Laboratory 8 – Group II Plasmid Preparation from <i>E. coli</i> and Sequencing of Plasmid DNA	Immunology Prof. Roers Anett Skupin	Institut für Immunologie
June 25	Laboratory 9 – Group I Transient Transfection of Eukaryotic Cells Western Blotting Technique	Immunology Prof. Roers Anett Skupin	Institut für Immunologie
July 02	Laboratory 9 – Group II Transient Transfection of Eukaryotic Cells Western Blotting Technique	Immunology Prof. Roers Anett Skupin	Institut für Immunologie